

DEPARTMENT OF CHEMISTRY

# GRADUATE COURSE IN MASS SPECTROMETRY: LECTURE 5



Separation methods (chromatography) – Mass spectrometry



Dr. James Wickens, 11<sup>th</sup> November 2015

# Chromatography

## History and milestones

Paper  
Chromatography

Partition  
Chromatography

New developments  
all the time

Column  
Chromatography

Tsvet

### The Nobel Prize in Chemistry 1952



Archer John Porter  
Martin

Prize share: 1/2



Richard Laurence  
Millington Synge

Prize share: 1/2

Erica  
Cremer

# Chromatography

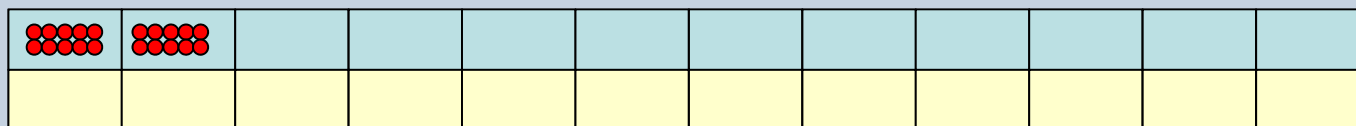
Mobile Phase



Stationary phase

# Chromatography

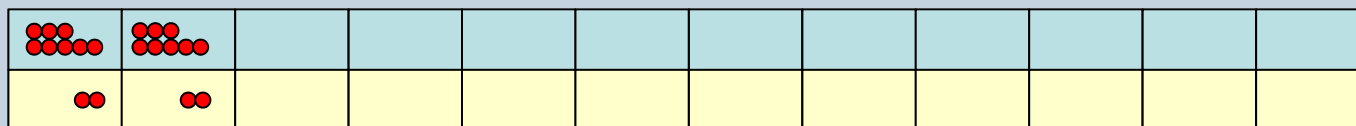
- Injection



80 : 20  
Mobile : Stationary

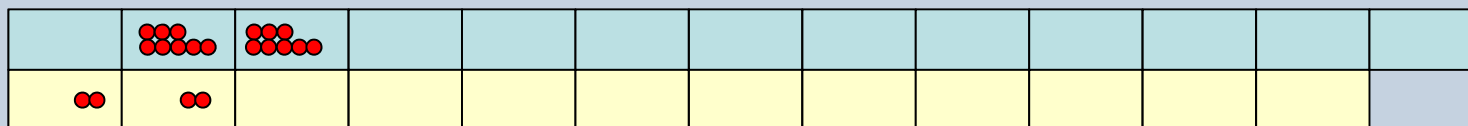
# Chromatography

- Equilibration



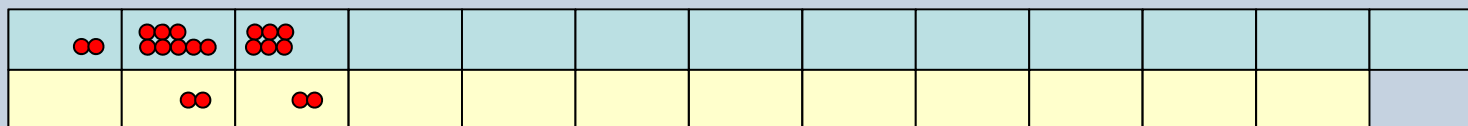
# Chromatography

- Mobile phase moves



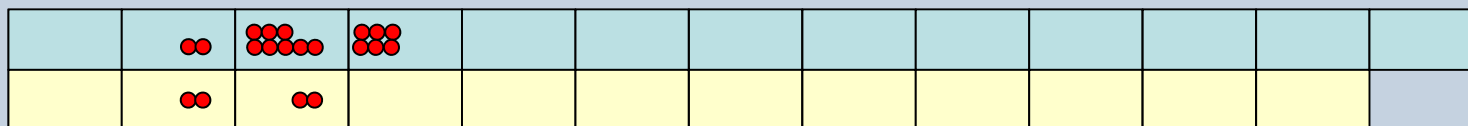
# Chromatography

- Equilibration



# Chromatography

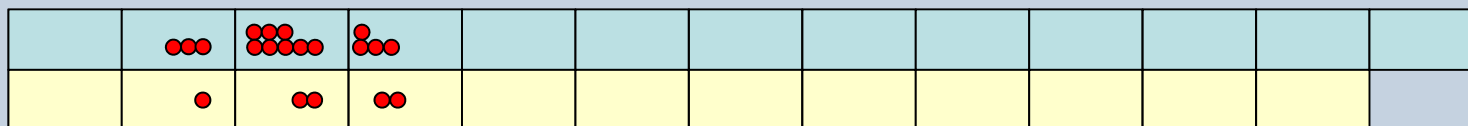
- Mobile phase moves





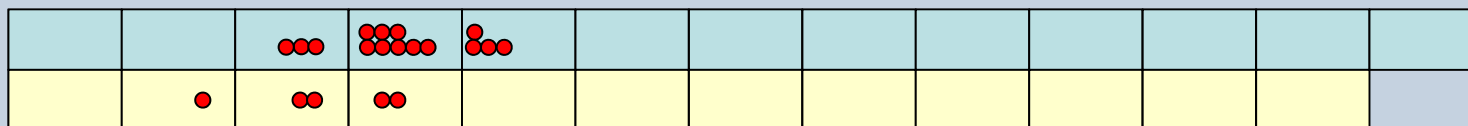
# Chromatography

- Equilibration



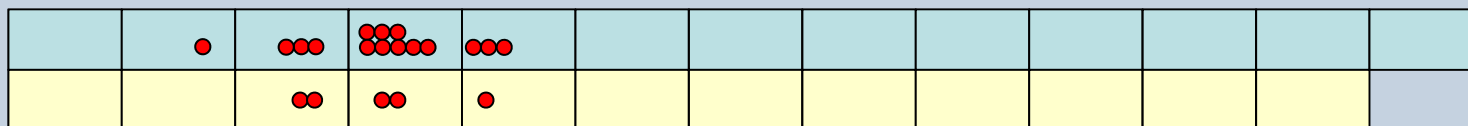
# Chromatography

- Mobile phase moves



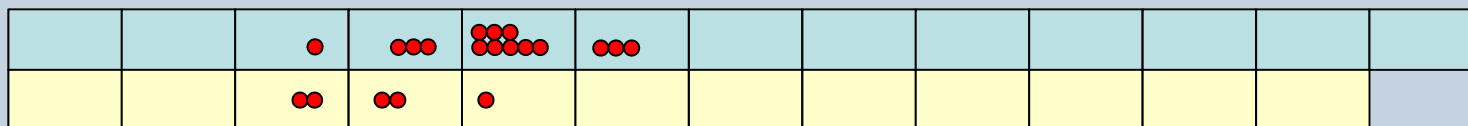
# Chromatography

- Equilibration



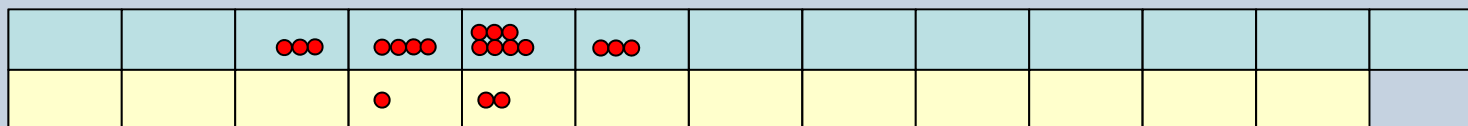
# Chromatography

- Mobile phase moves



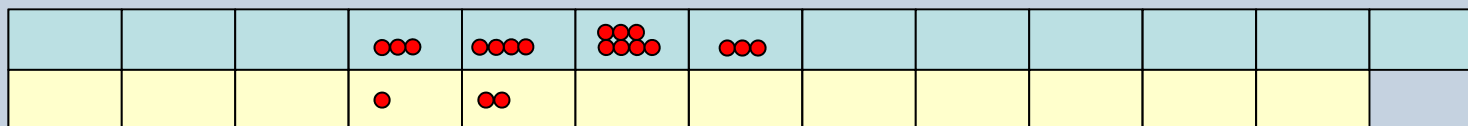
# Chromatography

- Equilibration



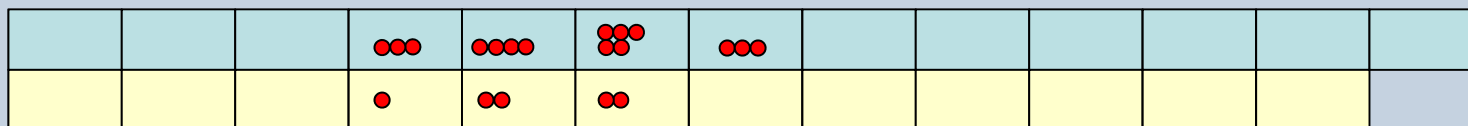
# Chromatography

- Mobile phase moves



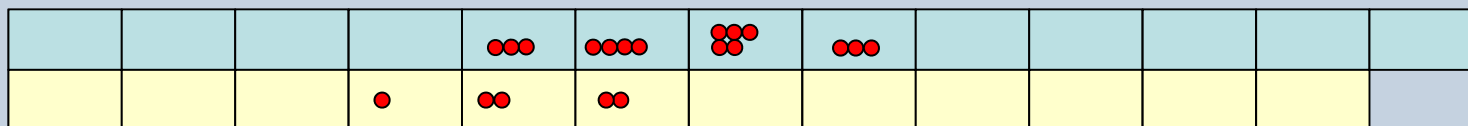
# Chromatography

- Equilibration



# Chromatography

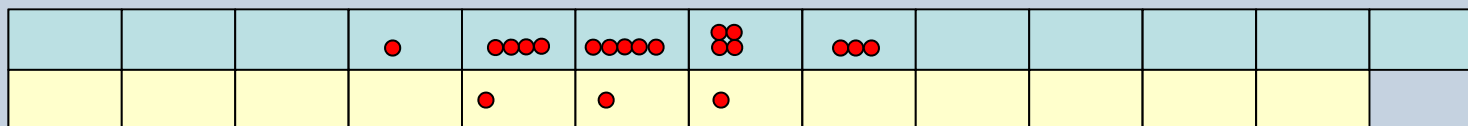
- Mobile phase moves



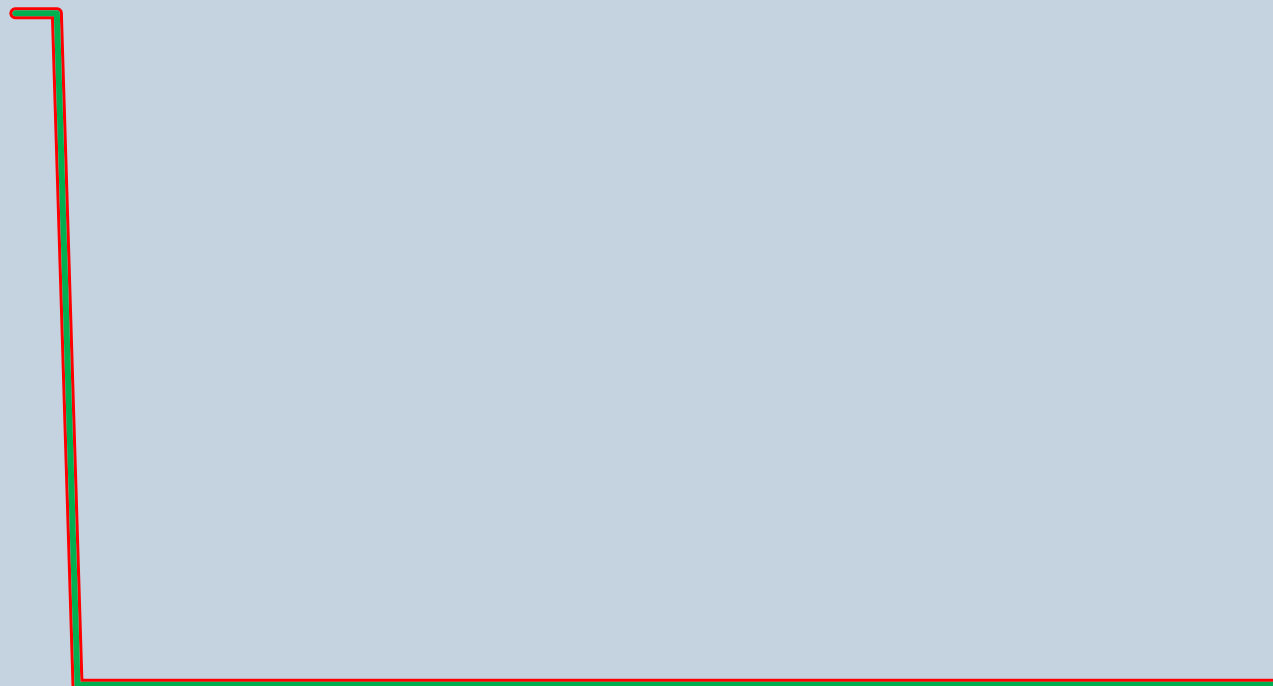


# Chromatography

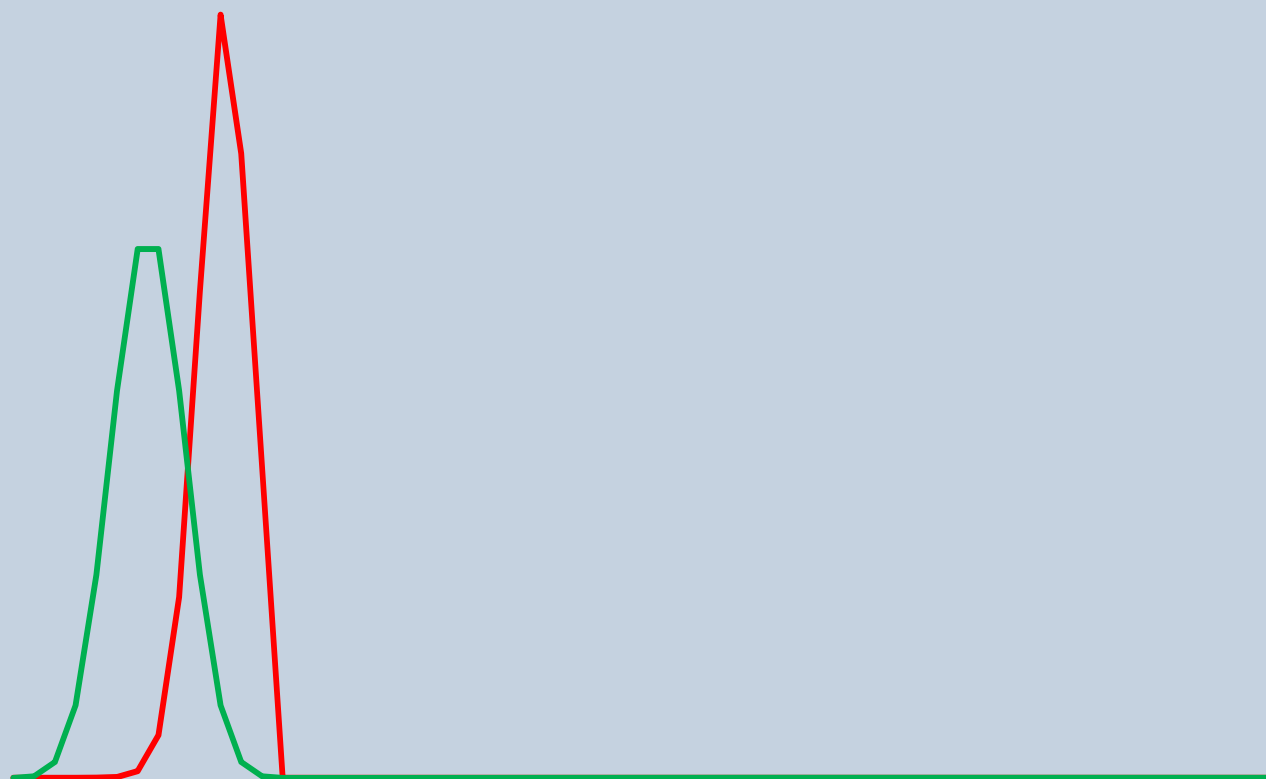
- Equilibration



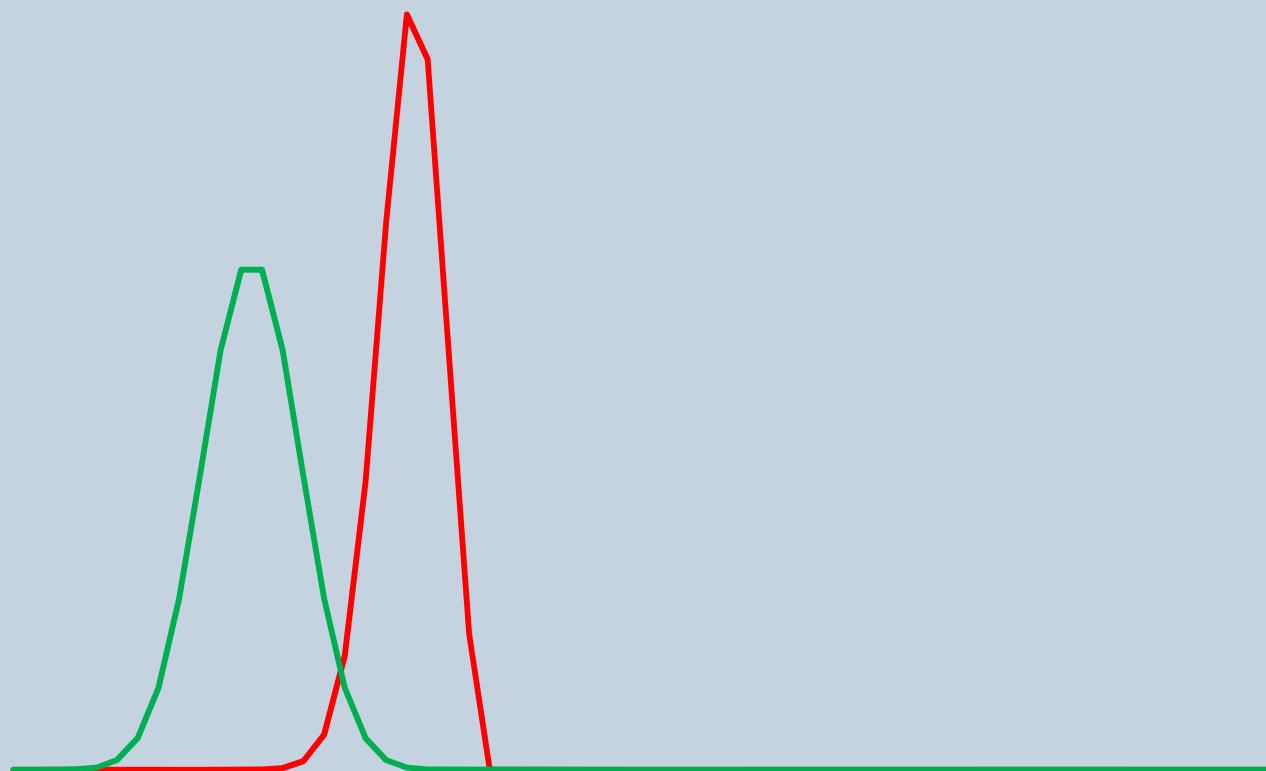
# Chromatography: Separation



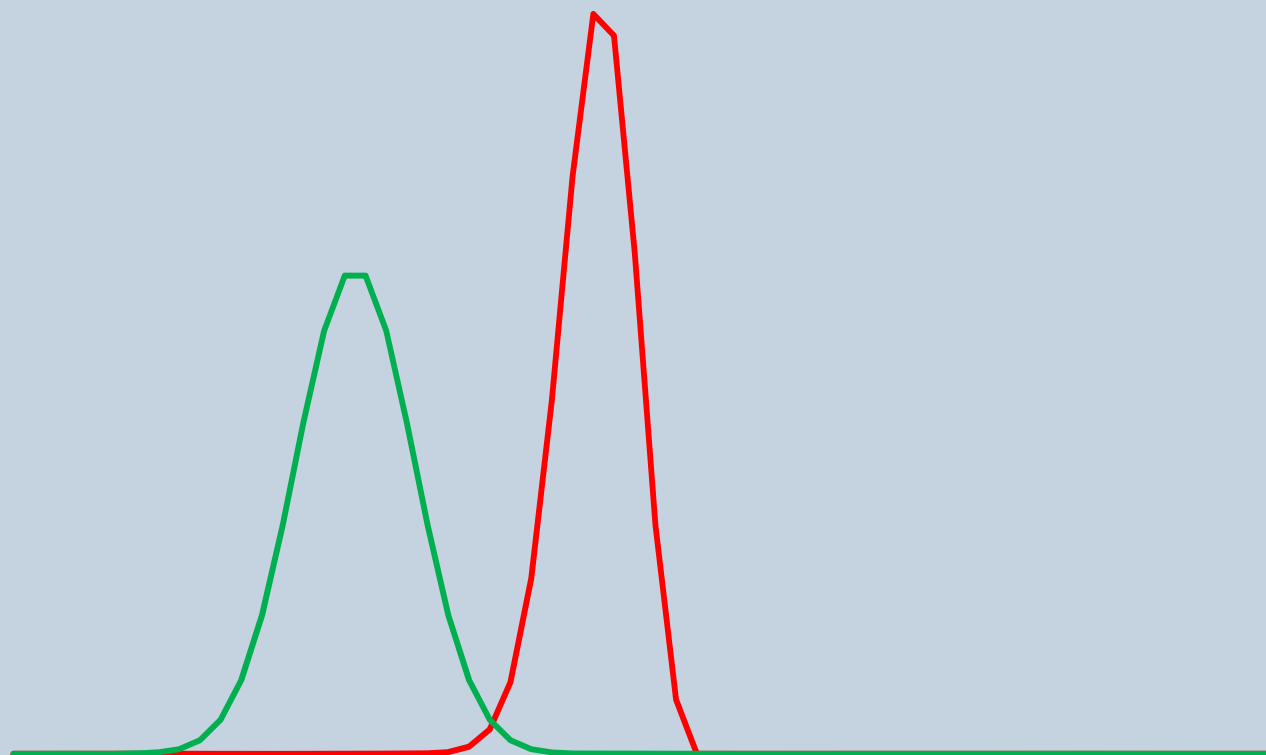
# Chromatography: Separation



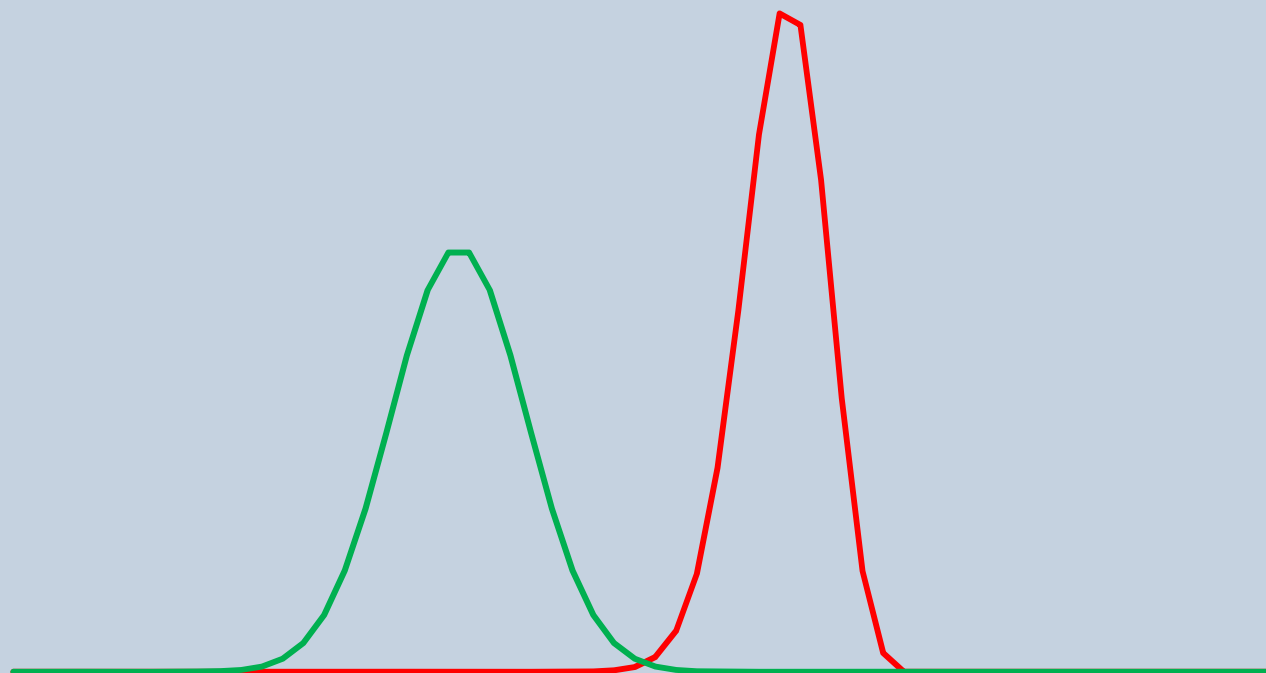
# Chromatography: Separation



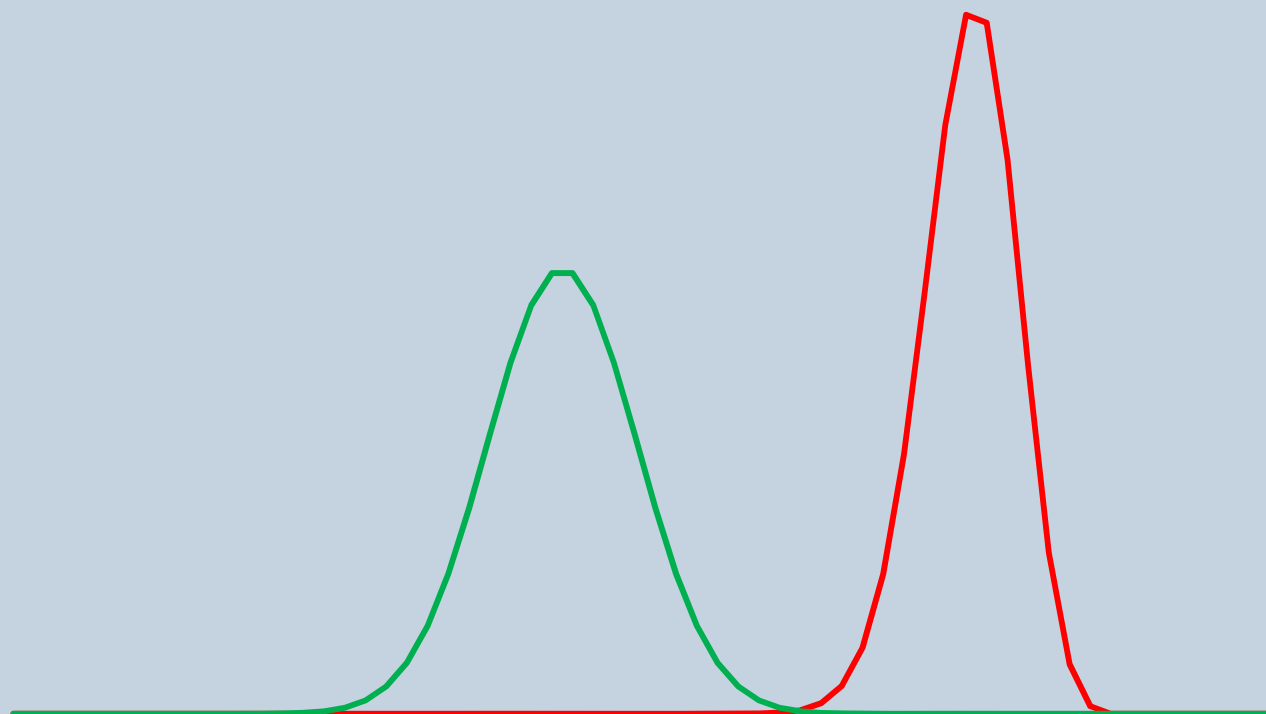
# Chromatography: Separation



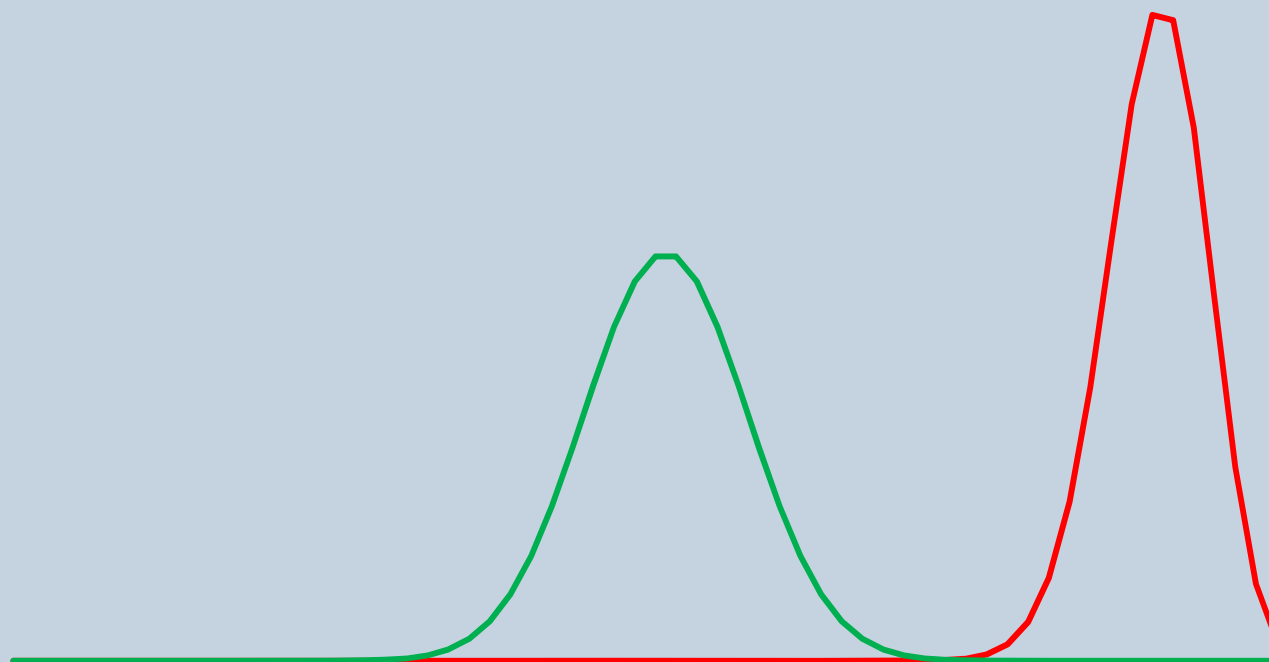
# Chromatography: Separation



# Chromatography: Separation



# Chromatography: Separation



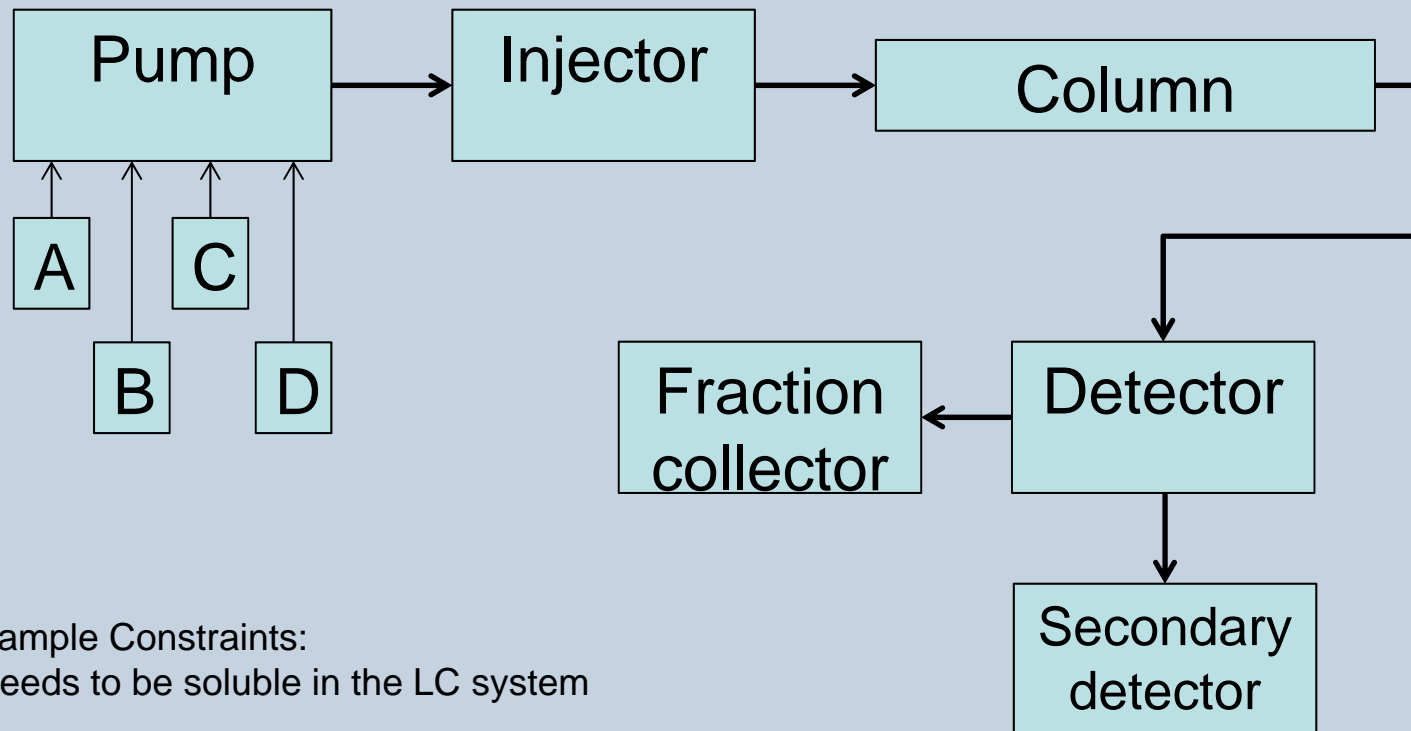


# Liquid Chromatography

- Basics
  - Experimental description, sample constraints etc.
- Scale
  - Preparative - Nano
- Hardware
  - LC
  - Physical description of the column
- Separation mechanisms
- MS interfaces

# Liquid Chromatography Basics

- High Performance Liquid Chromatography



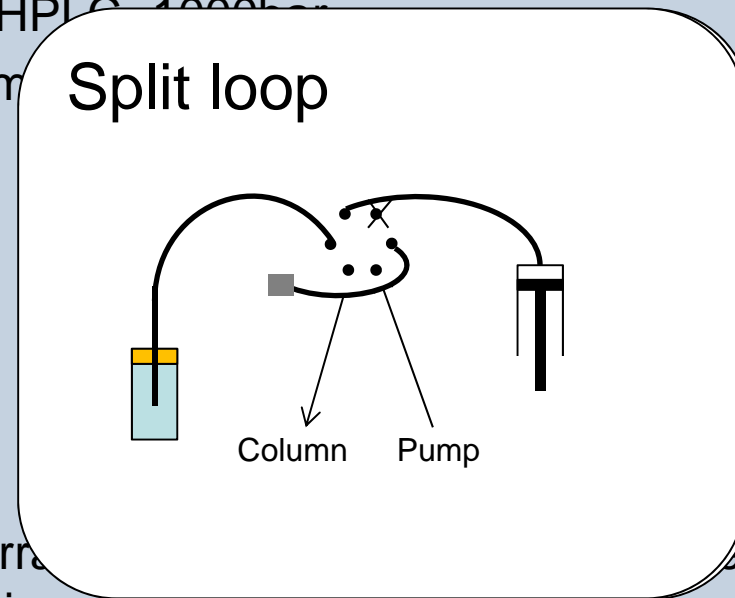
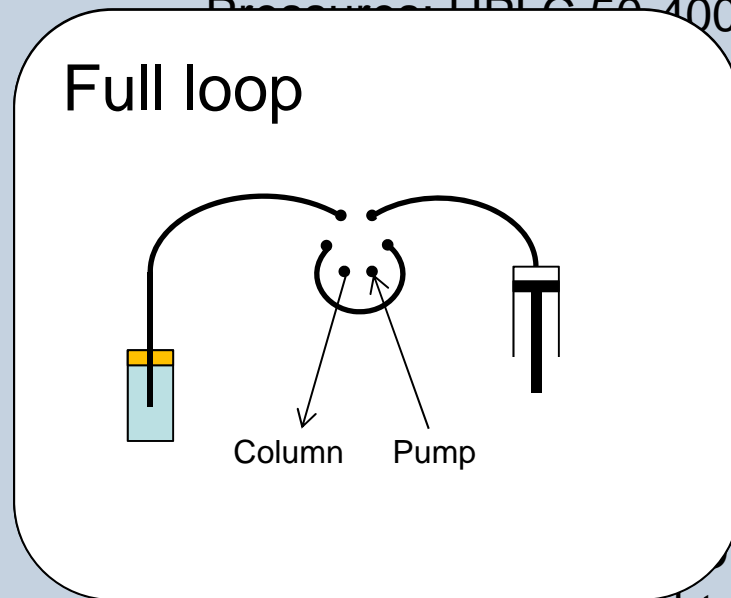
Sample Constraints:  
Needs to be soluble in the LC system

# Liquid Chromatography Scale

| Column Diameter | Description | Flow rate         |
|-----------------|-------------|-------------------|
| >100mm          | Industrial  | ~ 1 L/min         |
| 10 - 50mm       | Preparative | ~ 20 mL/min       |
| 2 - 4.6mm       | Analytical  | ~ 0.5 mL/min      |
| 0.5 – 1 mm      | Microflow   | ~ 100 $\mu$ L/min |
| 0.05 – 0.1 mm   | Nanoflow    | ~ 100 nL/min      |

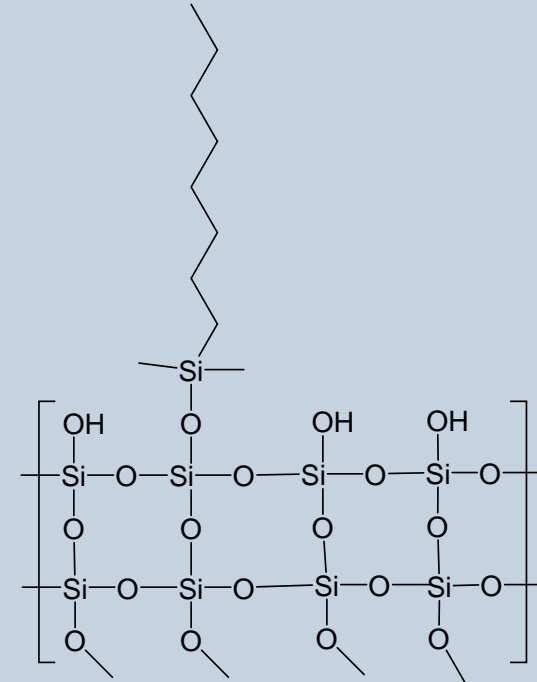
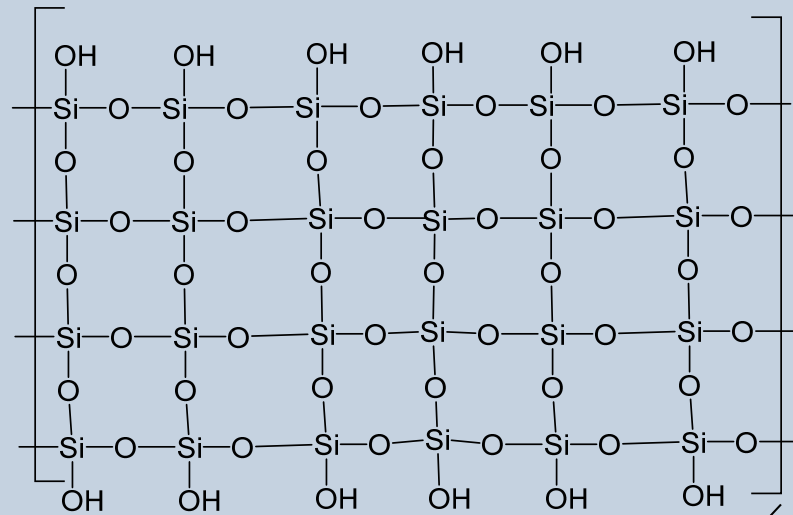
# Liquid Chromatography Hardware

- Pump



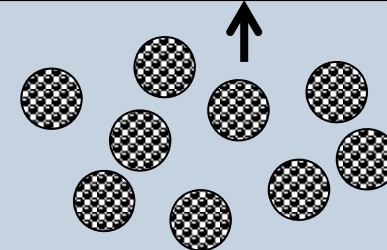
Mode Arr  
EX,  
evaporative light scattering detectors.

# Column technology



Milling

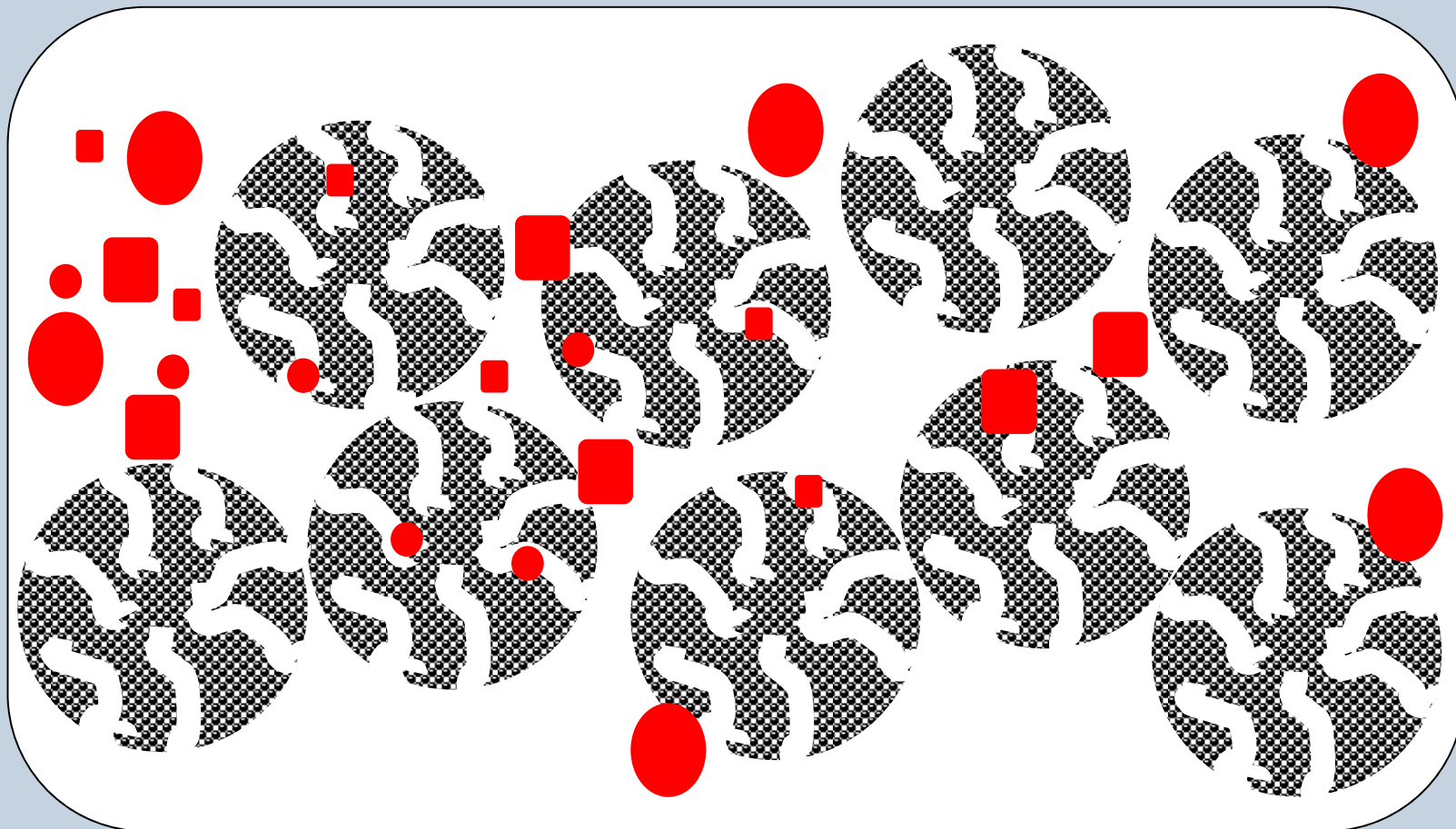
Derivatise



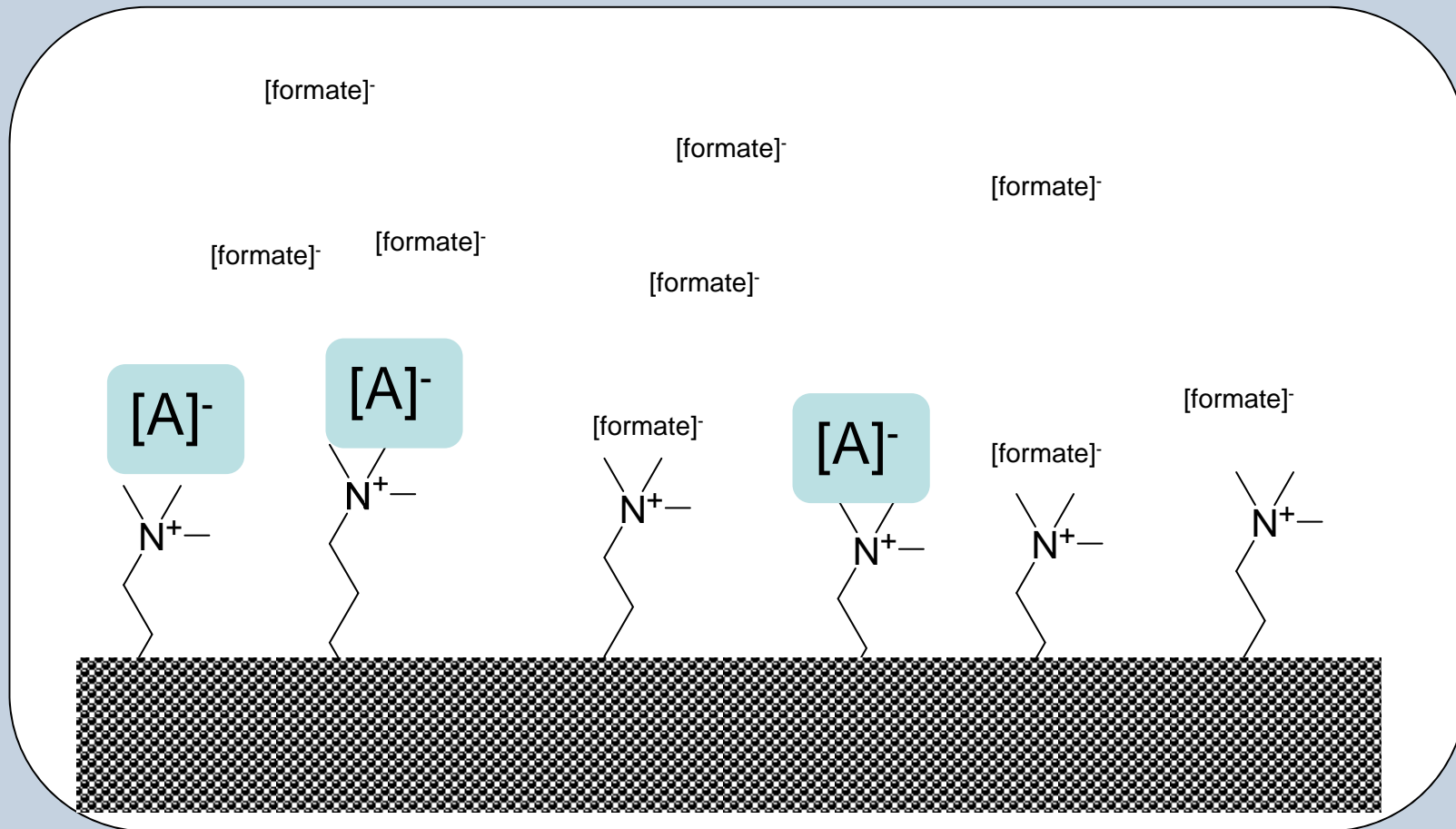
# Liquid Chromatography: Separation mechanisms:

- Size exclusion
- Ion exchange
- Normal Phase
  - Classic
  - Aqueous normal phase
  - SCF
- Reversed phase
  - Detailed discussion later

# Size exclusion chromatography



# Ion exchange chromatography





# Normal phase liquid chromatography

- Non-polar Mobile phase
  - Hexane, DCM, ethyl-acetate
- Polar stationary phase
  - Bare silica, amino, amide, and chiral phases
- Separation mechanisms
  - H-bond, Dipole interactions (stearic interactions)
- Ideal for:
  - Very hydrophobic analytes, very polar analytes, structural isomers, chiral separations
- Problems
  - Intolerant to impurities (water, protic solvents, salts)
- Variants
  - Aqueous normal phase/HILIC, super-critical fluid chromatography

# Reversed phase liquid chromatography

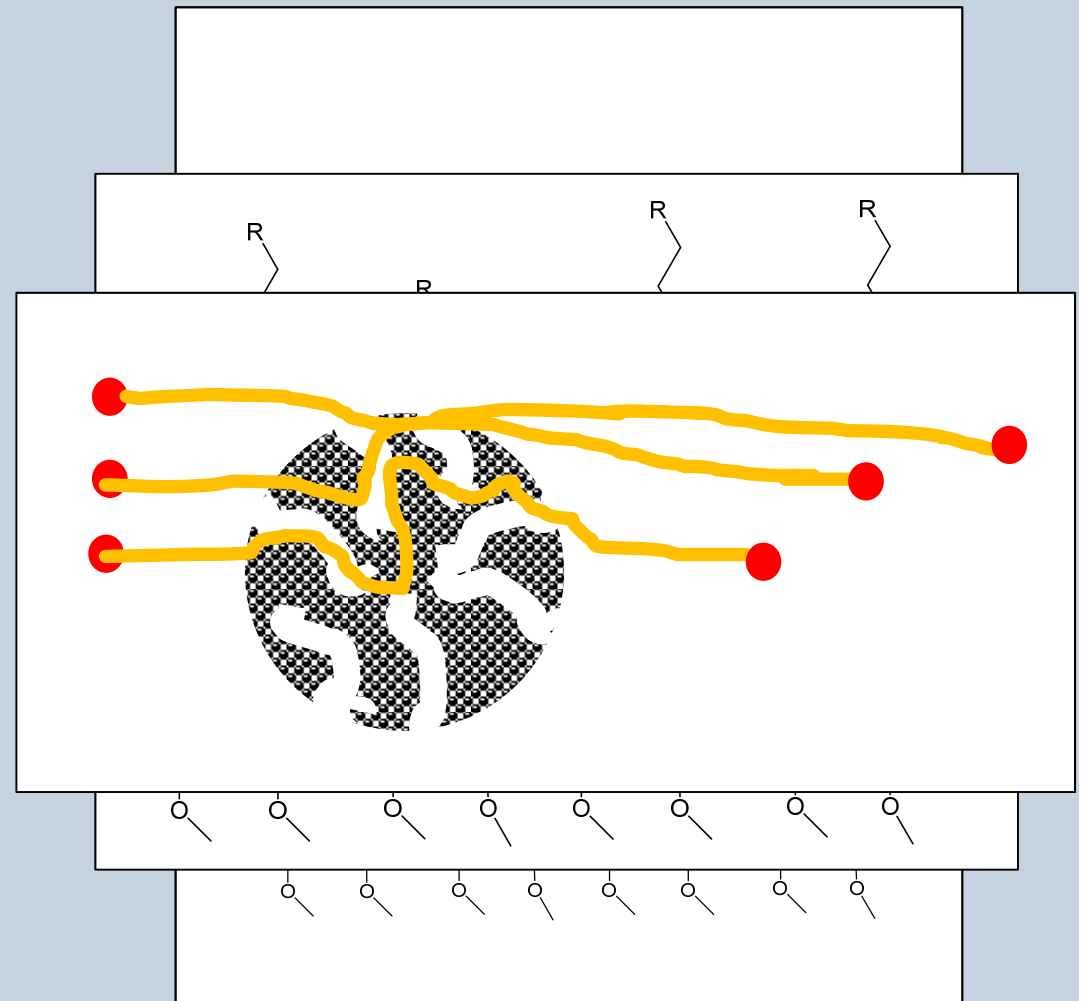
The most popular type of liquid chromatography

- Polar Mobile phase
  - Water, Methanol, Acetonitrile
- Non-polar stationary phase
  - C18, C8, C4, phenyl etc.
- Separation mechanisms
  - Hydrophobicity/lipophilicity
- Ideal for:
  - Very wide range of analytes
  - High loading capacity
- Problems
  - Next slide

# Reversed phase liquid chromatography

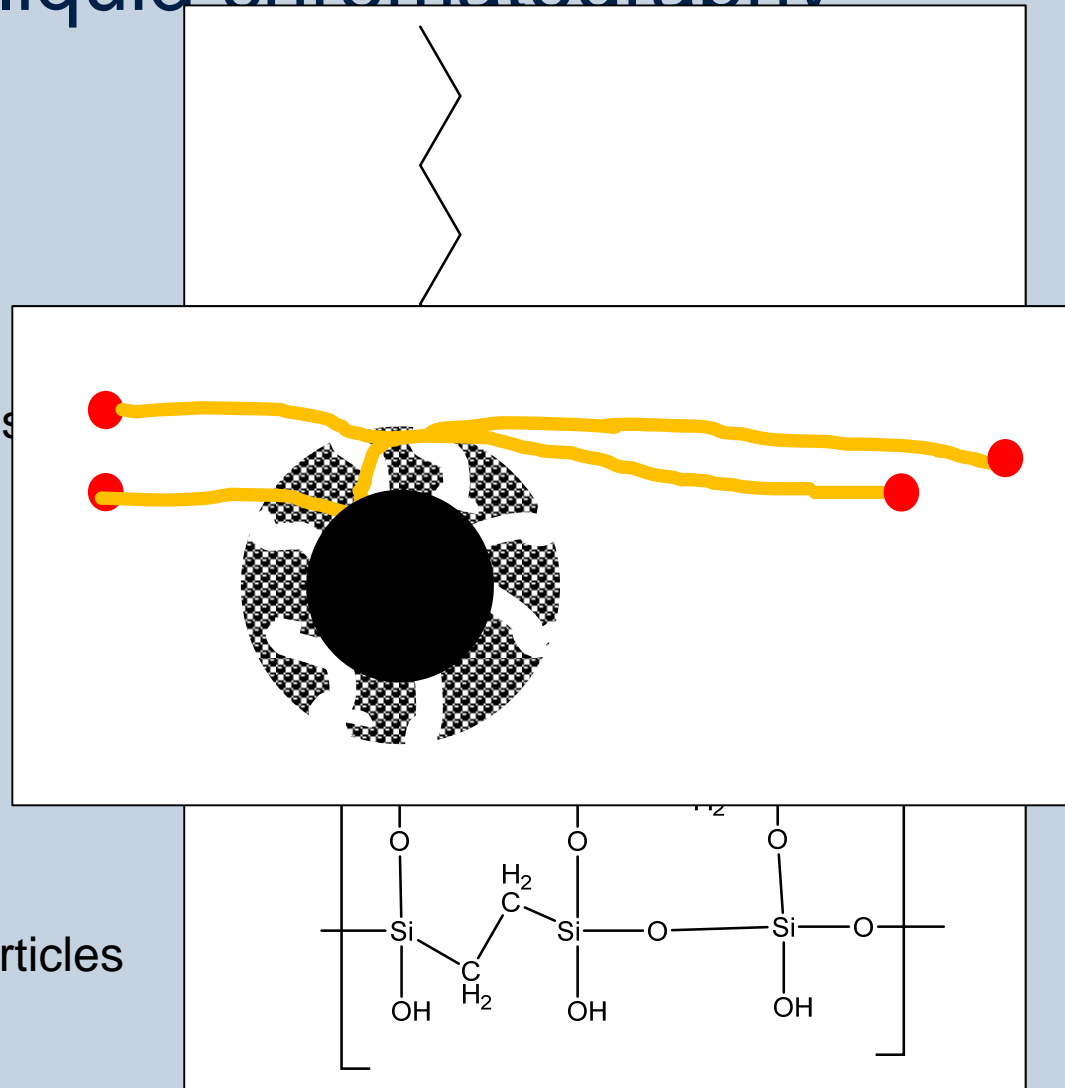
## Problems

- Very polar analytes
  - Ligand collapse
- Mixed mode interactions
  - Si-OH
- pH instability
  - 'Normal' silica pH 2-8
- Mechanical dispersion
  - Multiple paths
  - Size exclusion effects



# Reversed phase liquid chromatography solutions

- ~~Ion Pairing~~
- ligand redesign
  - Polar embedded groups
  - End-capping
  - Multi dentate
- Hybrid silica
- Silica support redesign
  - Smaller particles
  - Superficially porous particles
  - monoliths



# Connecting LC to MS

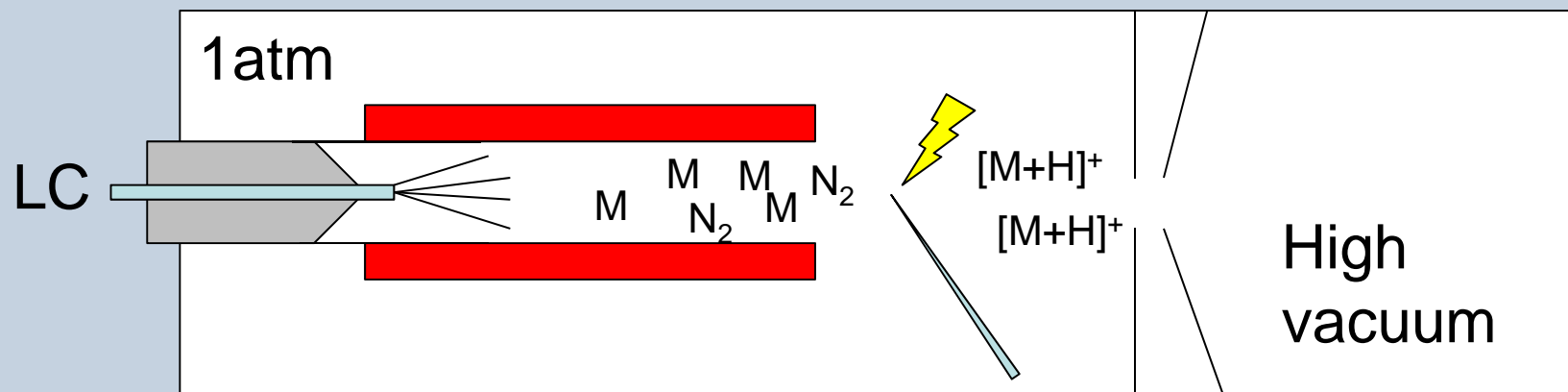
Many older and less frequently used techniques:

- APPI, Thermospray, Moving belt interface, continuous flow FAB

Two major techniques:

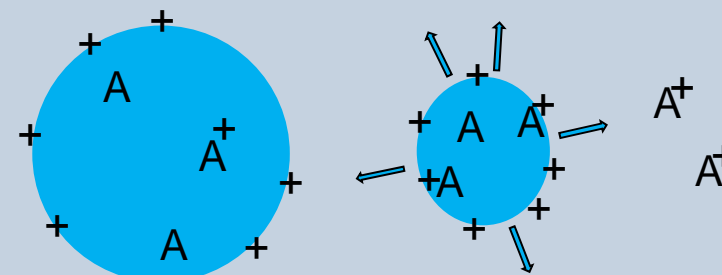
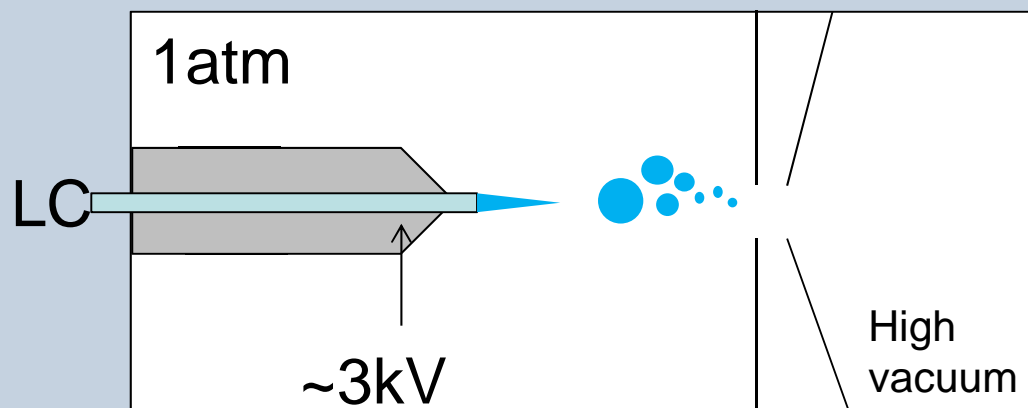
- APCI
- Electrospray

# LC-APCI



- Flow rate: ~1 mL/min design dependant
  - Sufficient to ensure nebulisation
- Solvent: compatible with non-polar solvents
  - Make sure your source gas is nitrogen... BANG!
- Tolerant to matrix introduction
- Problems
  - Soot formation, limited chemical range

# LC-Electrospray



- Flow rate: The lower the better
  - 'Standard' sources <math><0.2\text{ mL/min}</math>. → flow splitting
  - Nano flow has incredible sensitivities
- Solvent: water miscible, protic
- Problems
  - Matrix effects

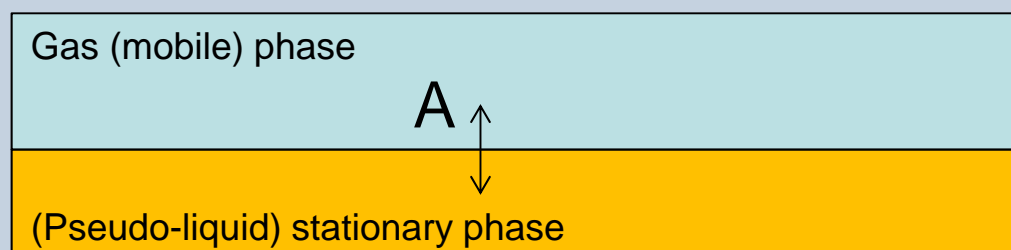
# Gas Chromatography

- Basics
- Hardware
  - GC
  - Packed column vs. capillary
  - Stationary phase chemistries
    - Non-polar
    - Polar
    - Specialist
- GC-MS interfacing



# Gas Chromatography Basics

- Principle separation mechanism is volatility



He, H<sub>2</sub>,  
N<sub>2</sub>, Ar

- Samples must be volatile or capable of being made volatile
- Can be packed column or Capillary column
- Can be very efficient (narrow peaks)
  - Fast diffusion
  - Open tubular column; no multiple path diffusion



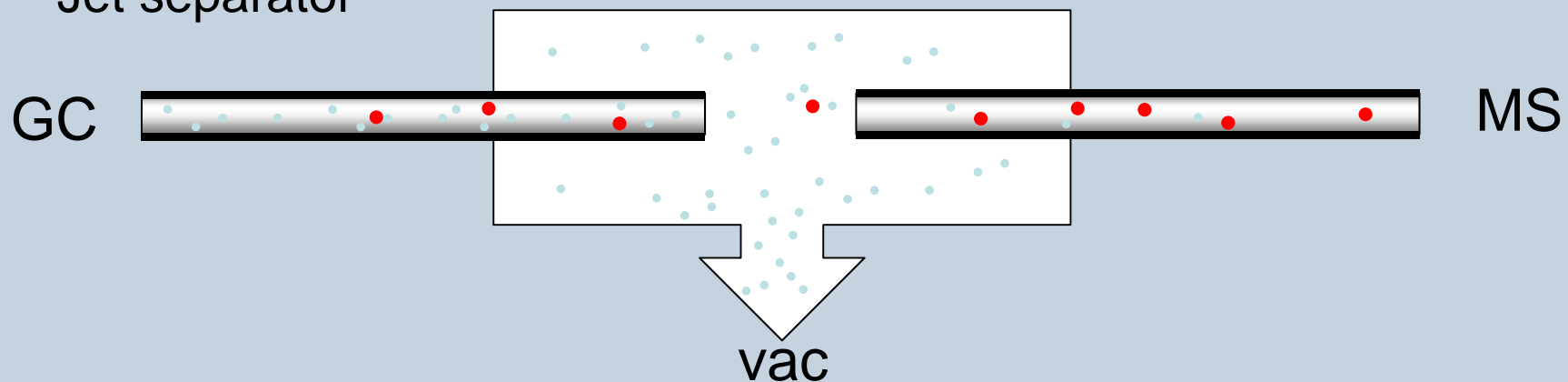
# Gas Chromatography

## Stationary phase chemistries

- NonPolar
    - “1”s (HP1, ZB1) Poly-dimethylsiloxane
    - “5”s (HP5, ZB5)
    - “50”s Poly-diphenylsiloxane
    - “1701”s (DB1701) Poly-cyanopropyl-methylsiloxane
- ↓
- Polar
    - “Wax” Polyethylene glycol

# GC-MS interface

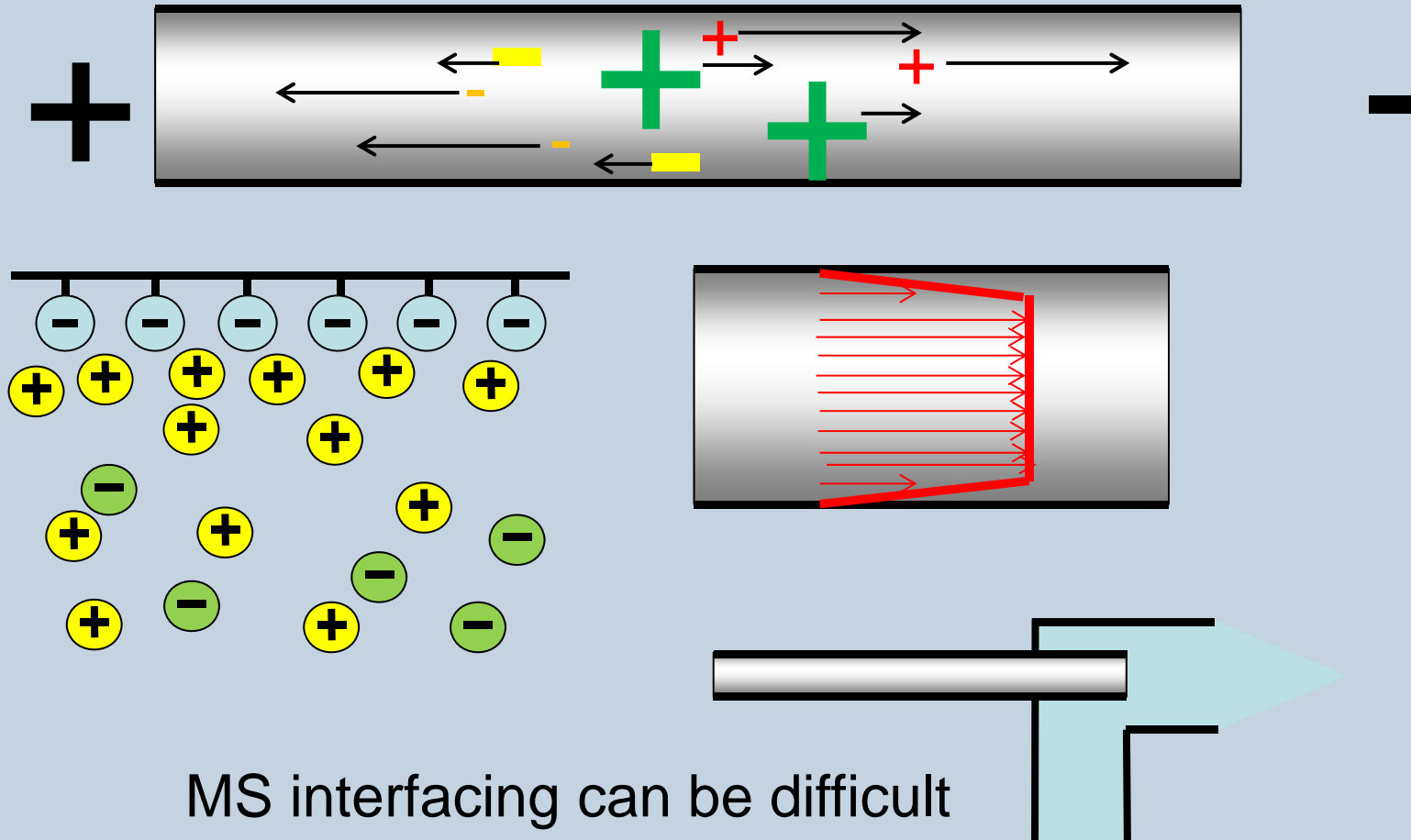
Packed column & capillaries > 0.53 mm  
Jet separator



0.32, 0.25, 1mm capillaries



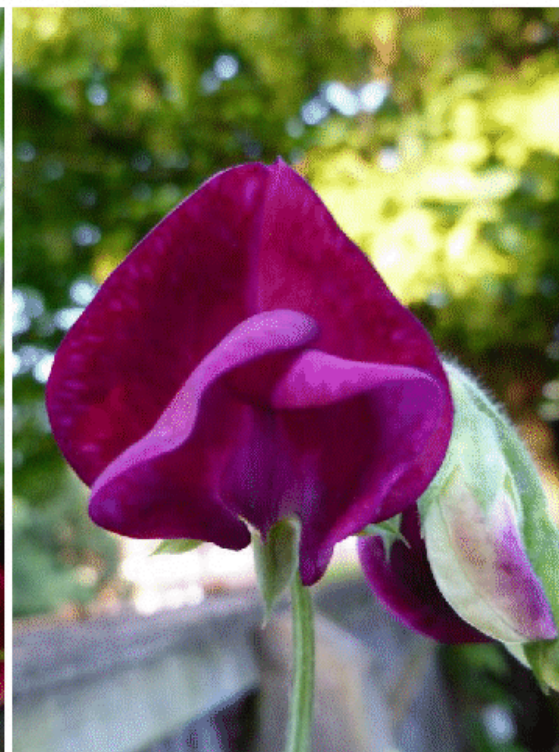
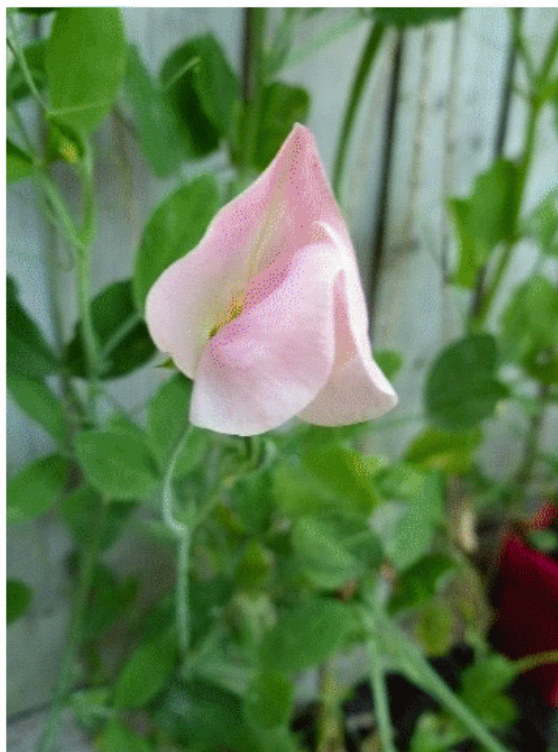
# Capillary electrophoresis



# Example 1

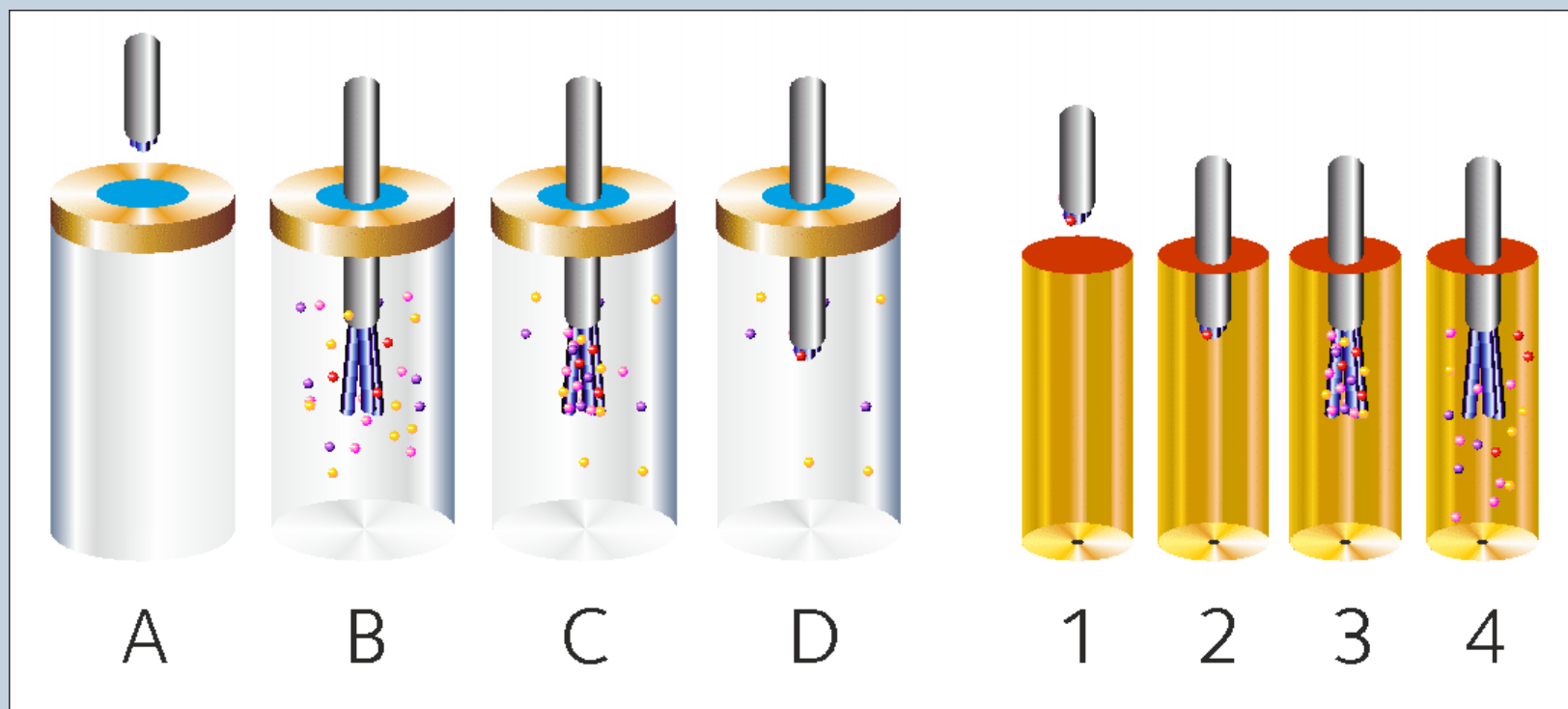
## Introduction

The new GC-MS system in the mass spectrometry research facility, department of Chemistry enables enrichment and analysis of volatile compounds. Here we present analysis of scent from individual sweetpea flowers



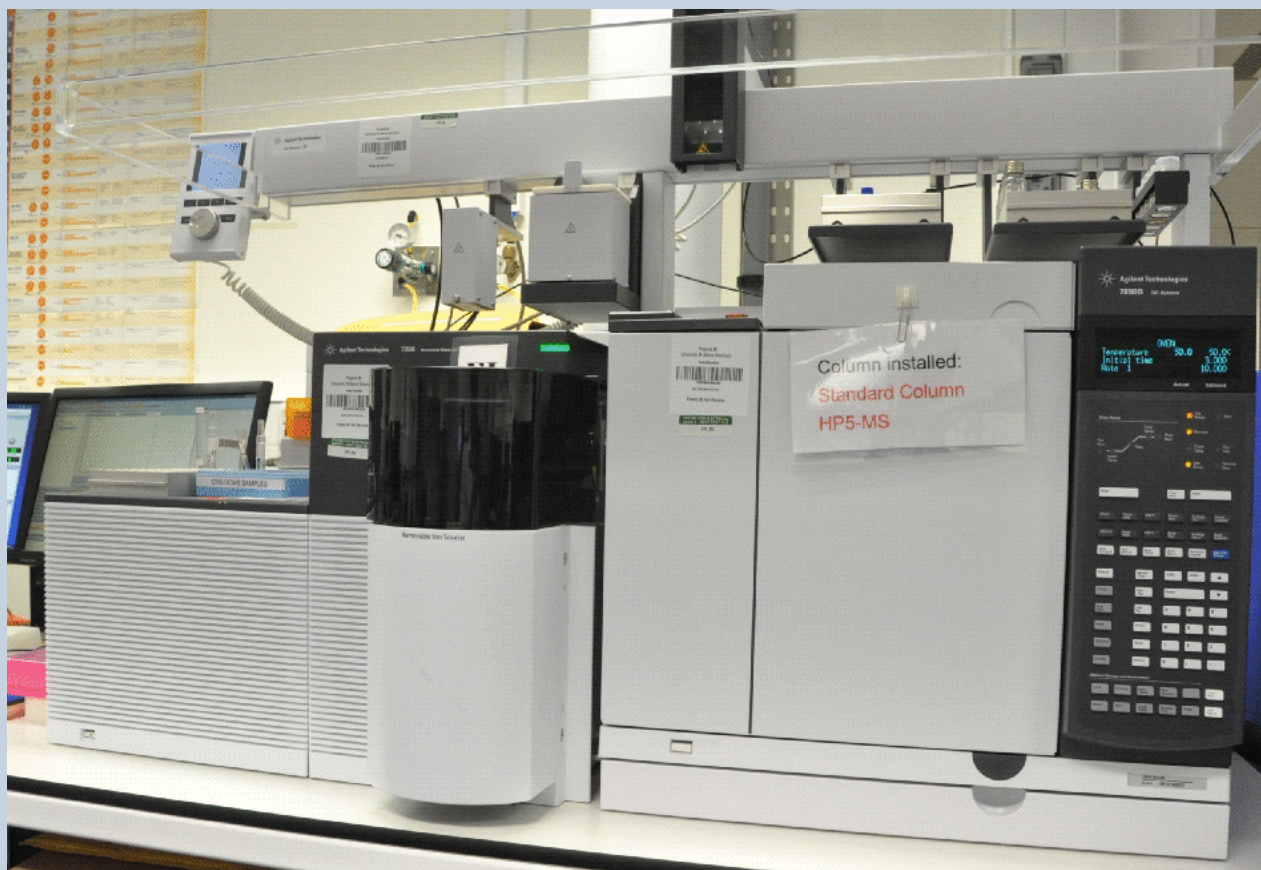
# Example 1

## SPME



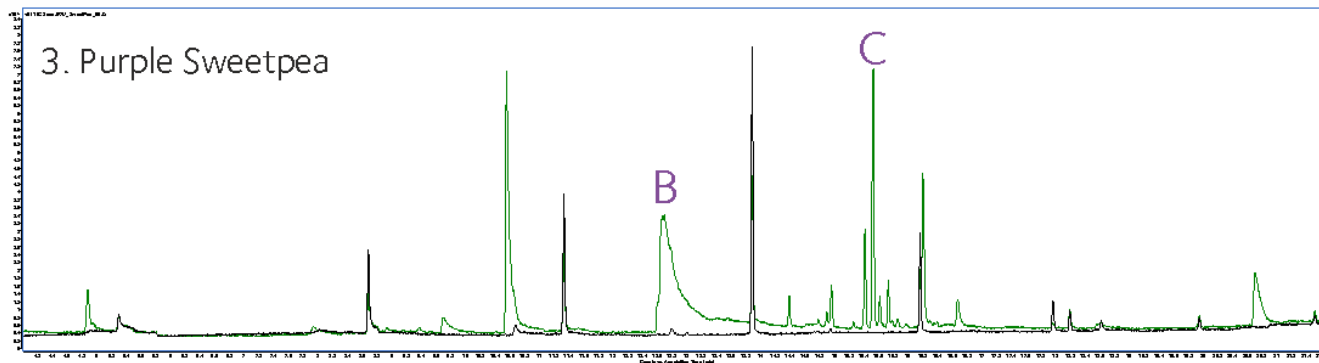
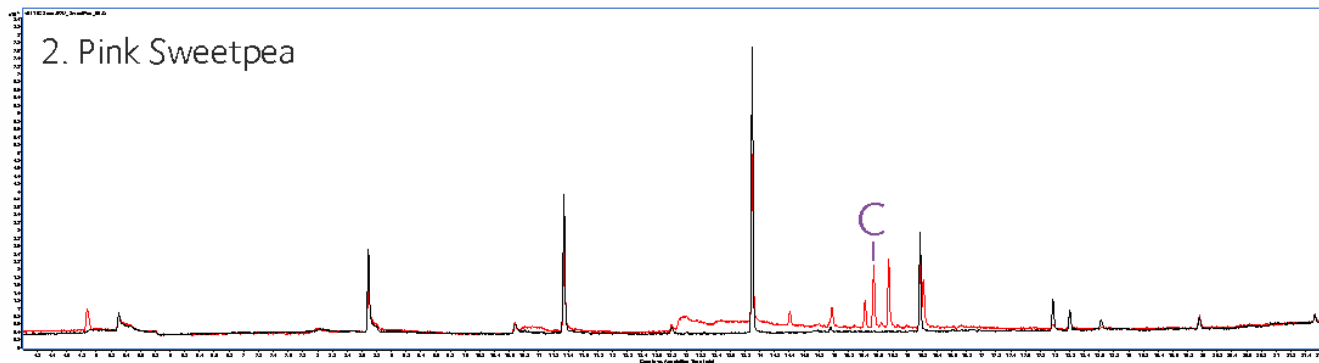
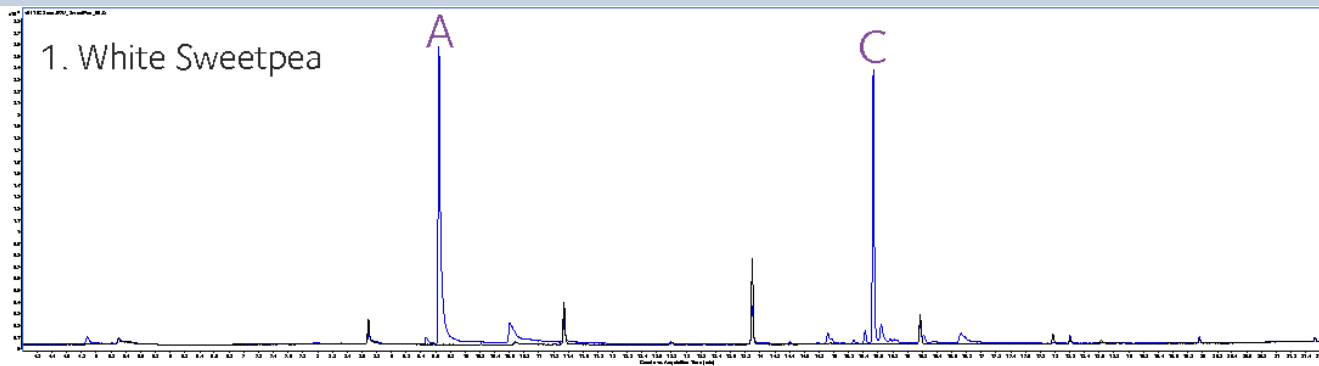


# Example 1



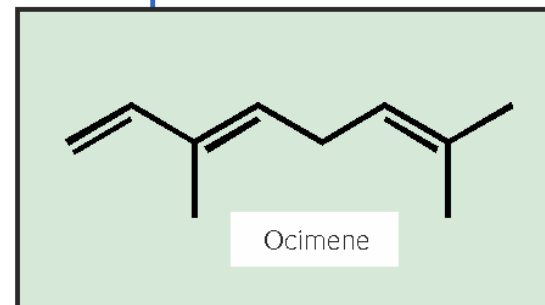
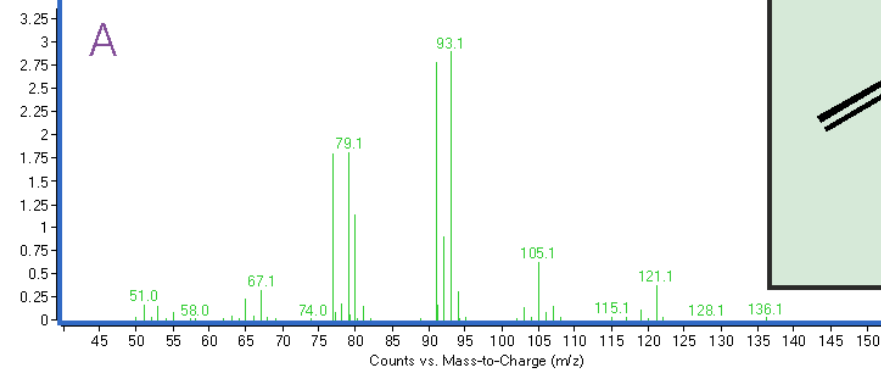


# Example 1

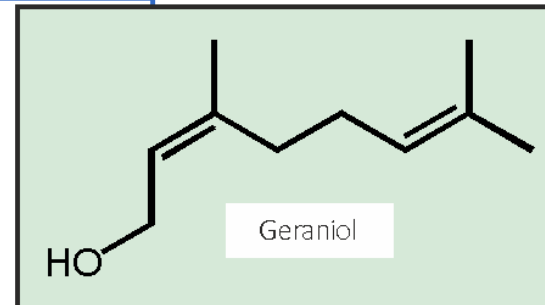
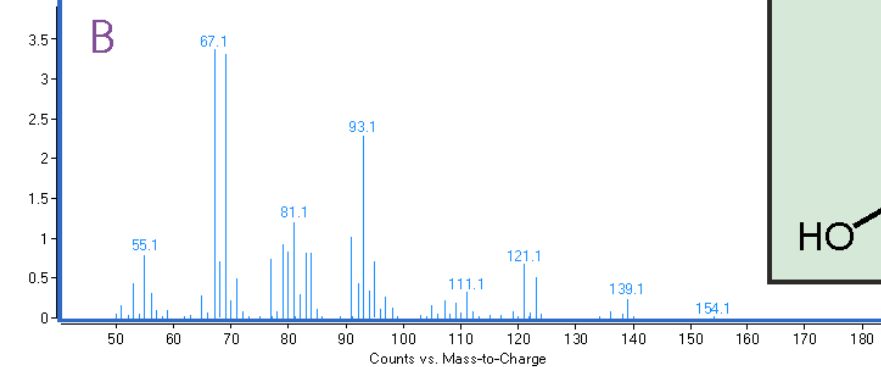


# Example 1

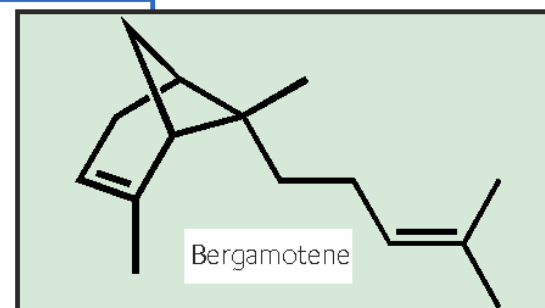
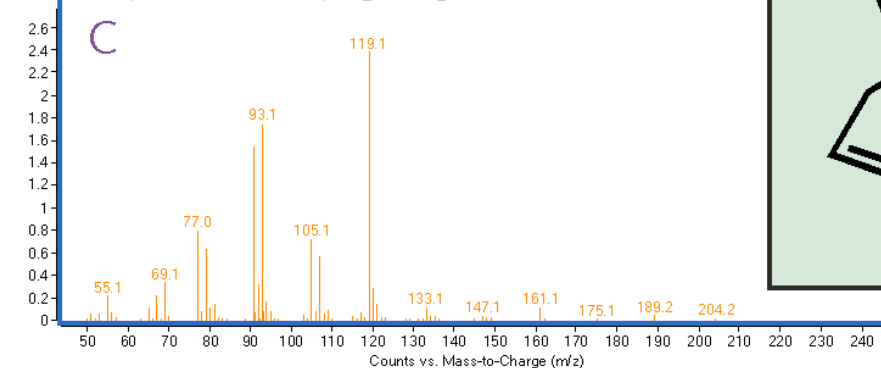
+EI Scan (9.636-9.680 min, 14 Scans) JRW\_SweetPea\_07.D Subtract



+EI Scan (12.657-12.728 min, 22 Scans) JRW\_SweetPea\_09.D

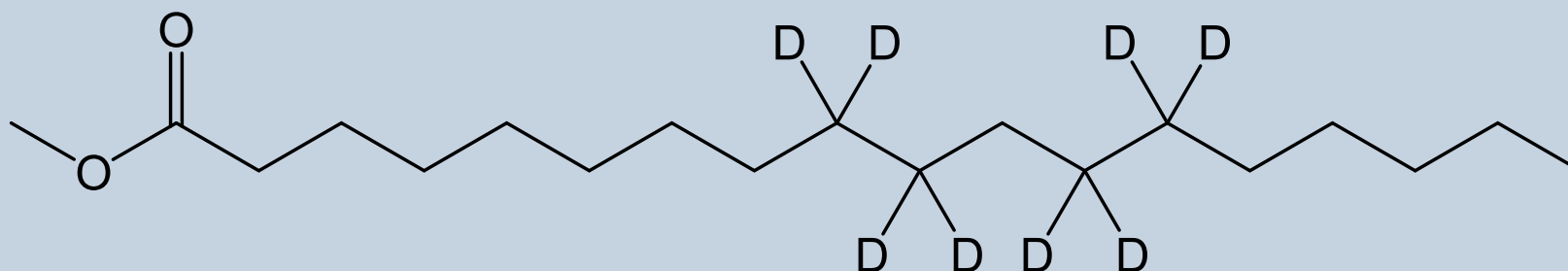


+EI Scan (15.522-15.562 min, 13 Scans) JRW\_SweetPea\_07.D Subtract



Mass spectra of volatile compounds from the peaks labelled in the chromatograms above. Chemical structures are tentative.

# Deuteration investigation



D<sub>8</sub>? - C<sub>19</sub>H<sub>30</sub>D<sub>8</sub>O<sub>2</sub> rmm=306

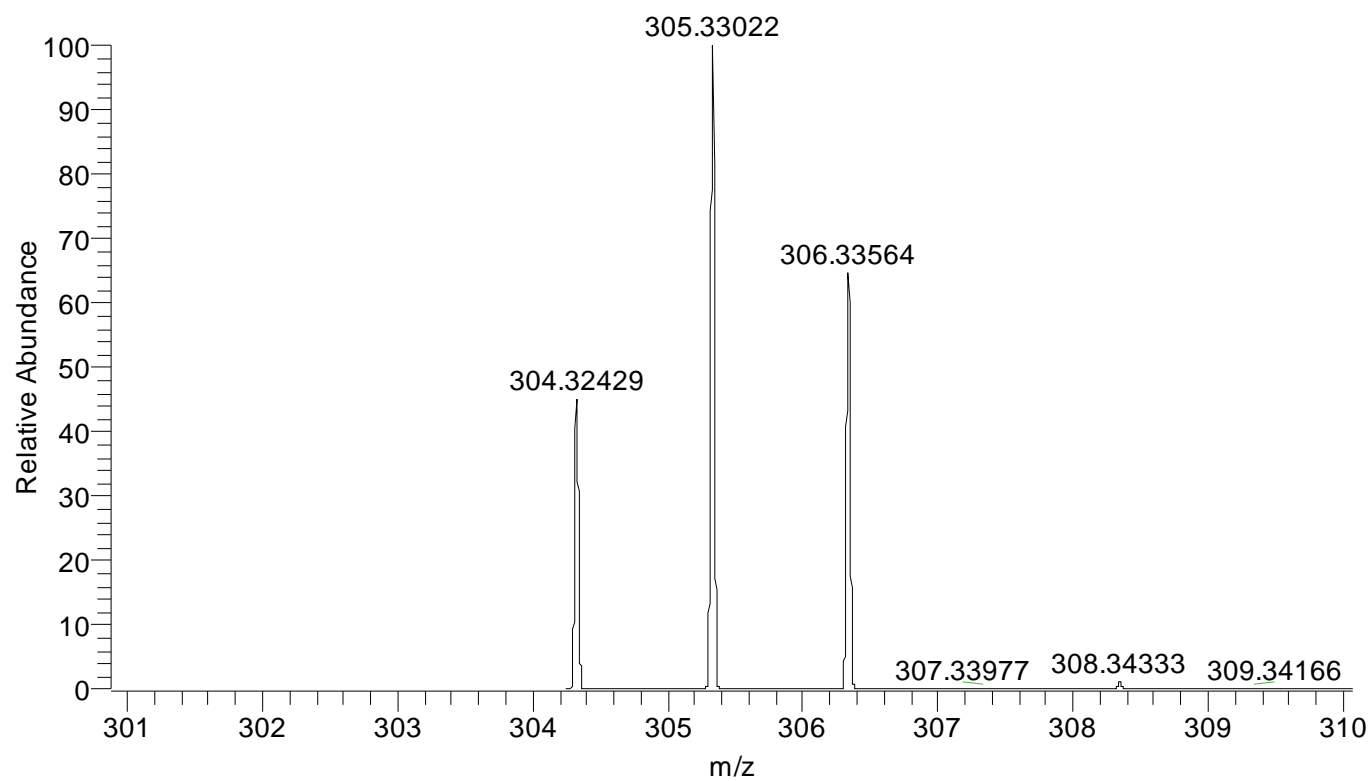
D<sub>7</sub>? - C<sub>19</sub>H<sub>31</sub>D<sub>7</sub>O<sub>2</sub> rmm=305

D<sub>6</sub>? - C<sub>19</sub>H<sub>32</sub>D<sub>6</sub>O<sub>2</sub> rmm=304

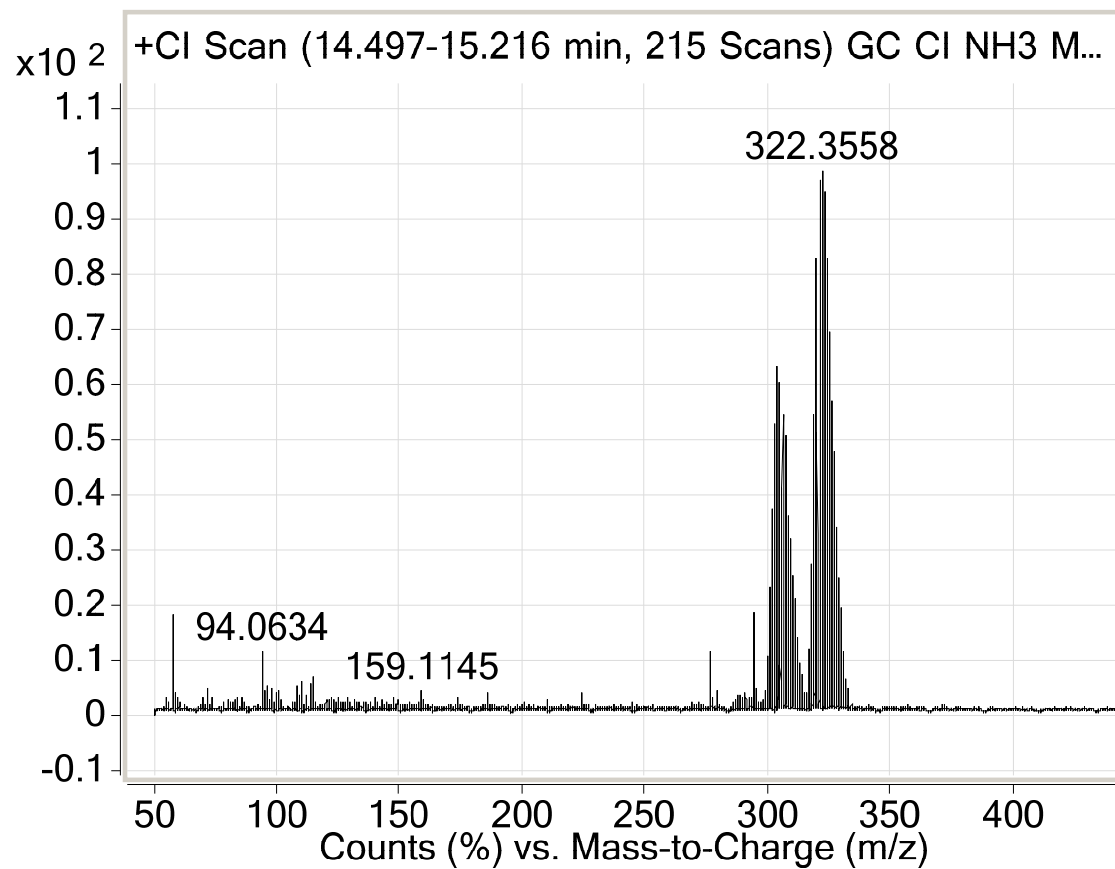
# Simulation

1-d<sub>6</sub> : 2-d<sub>7</sub> : 1d<sub>8</sub>

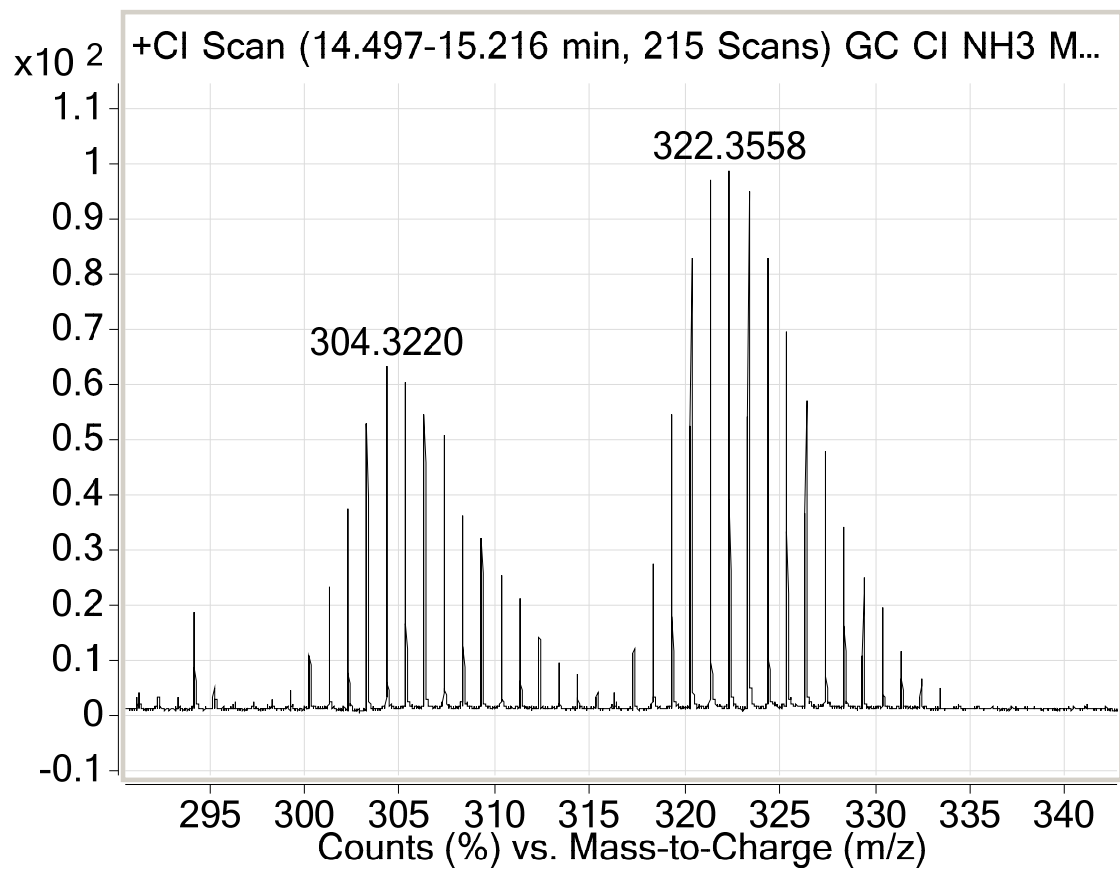
C<sub>19</sub>H<sub>30</sub>D<sub>8</sub>O<sub>2</sub>\*0.50 + C<sub>19</sub>H<sub>31</sub>D<sub>7</sub>O<sub>2</sub>\*1.00 + C<sub>19</sub>H<sub>32</sub>D<sub>6</sub>O<sub>2</sub>\*0.50: p(...



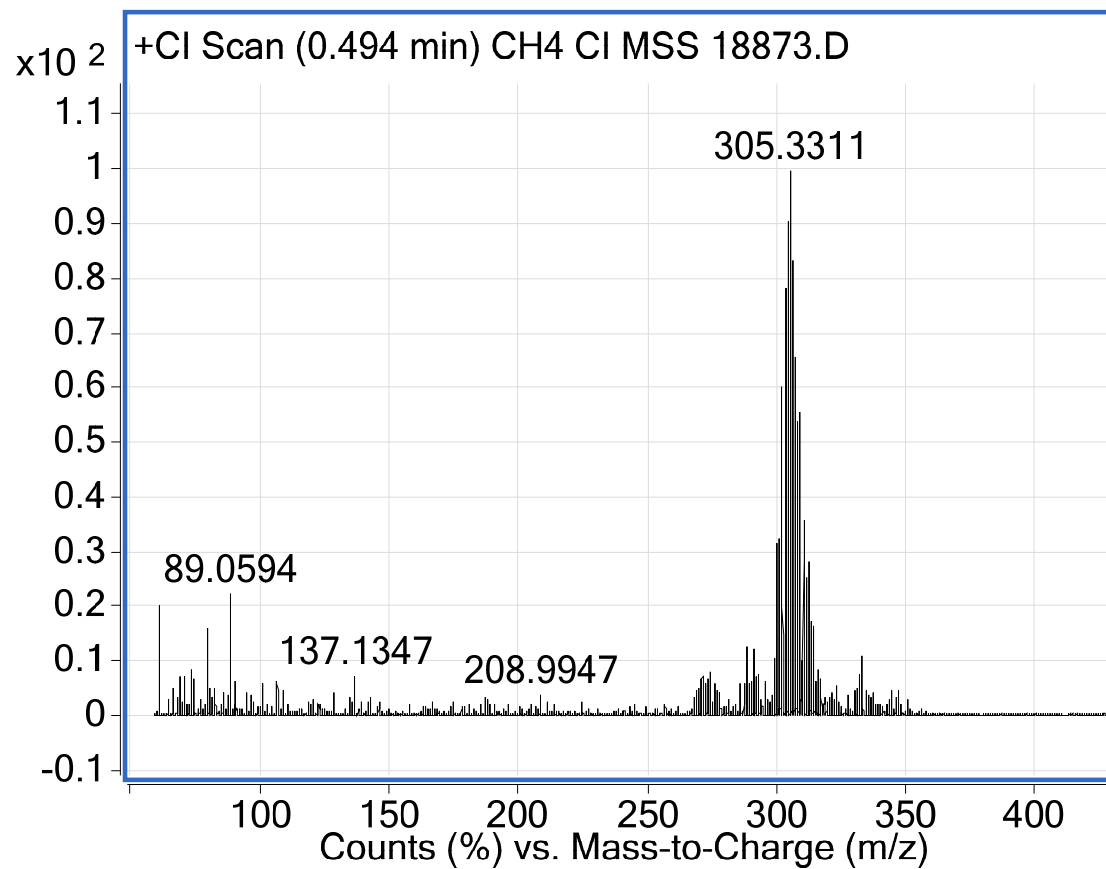
# Ammonia CI spectrum



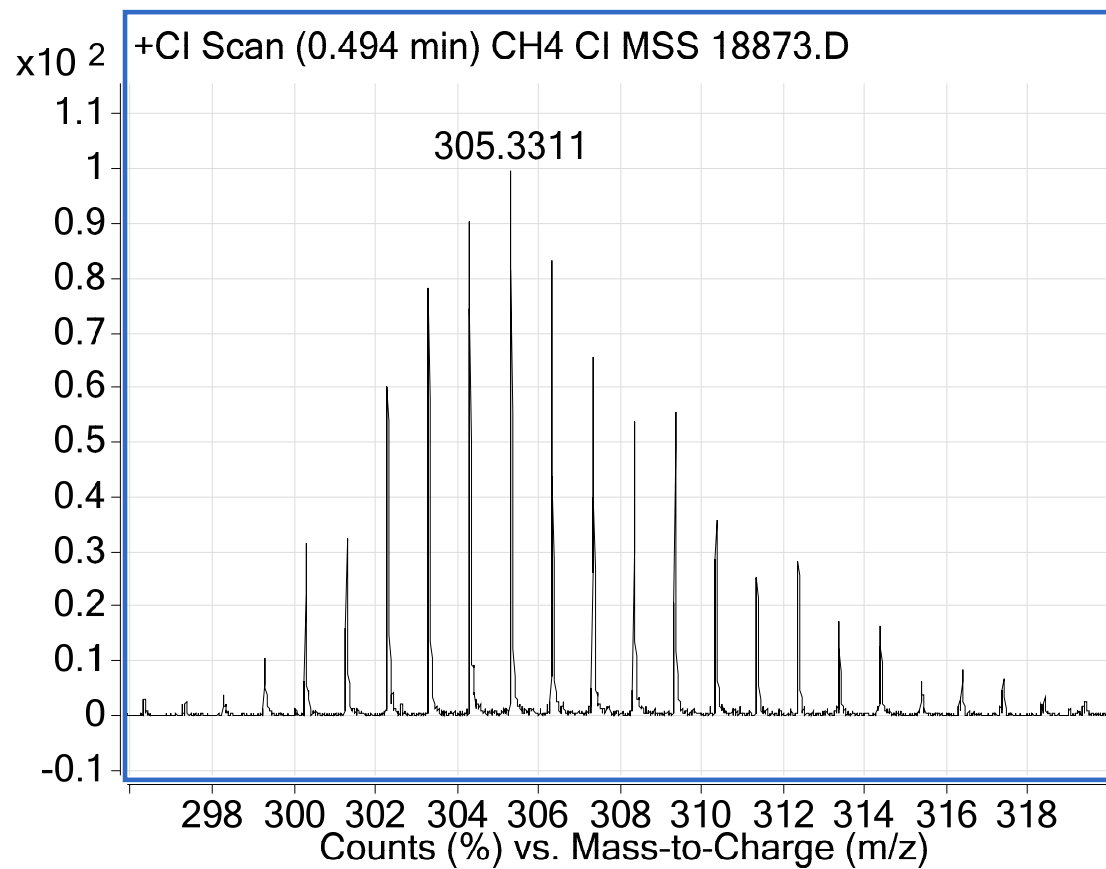
# Ammonia CI spectrum



# Methane CI spectrum

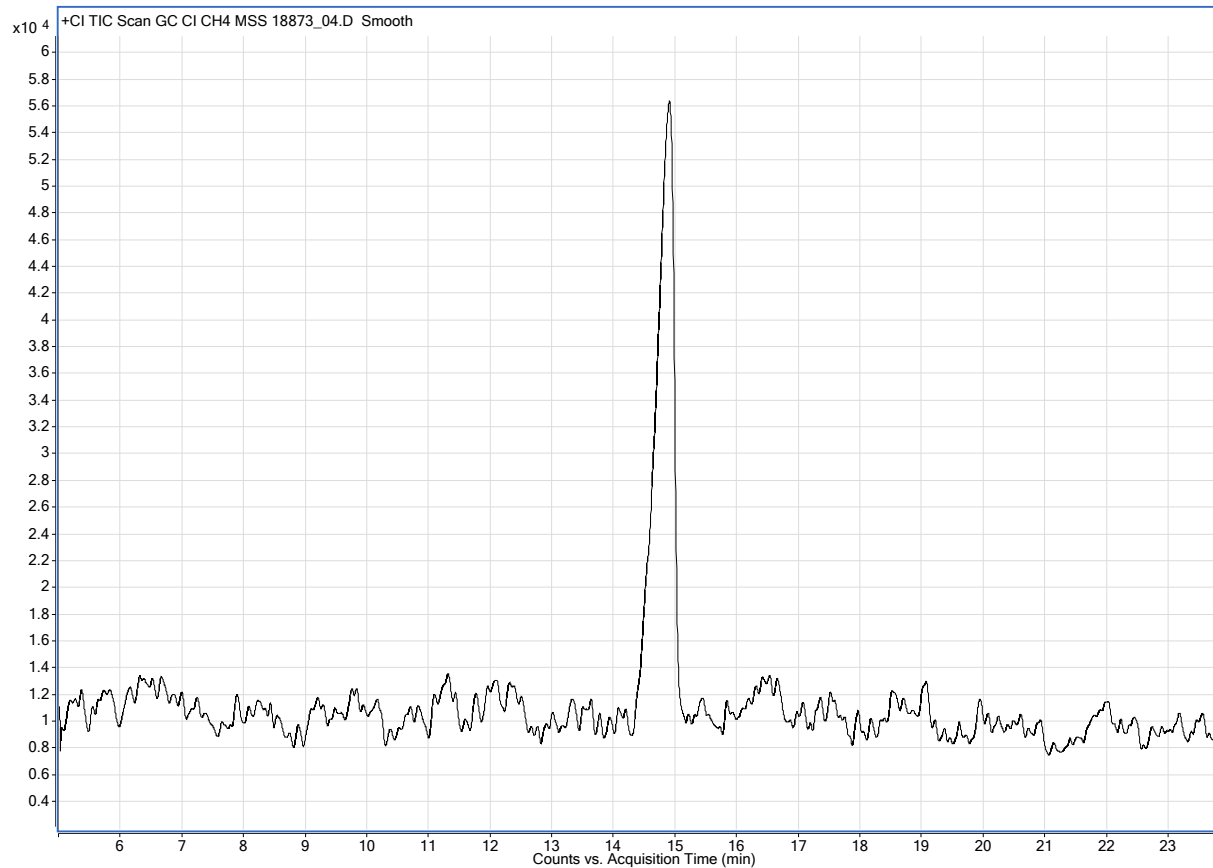


# Methane CI spectrum

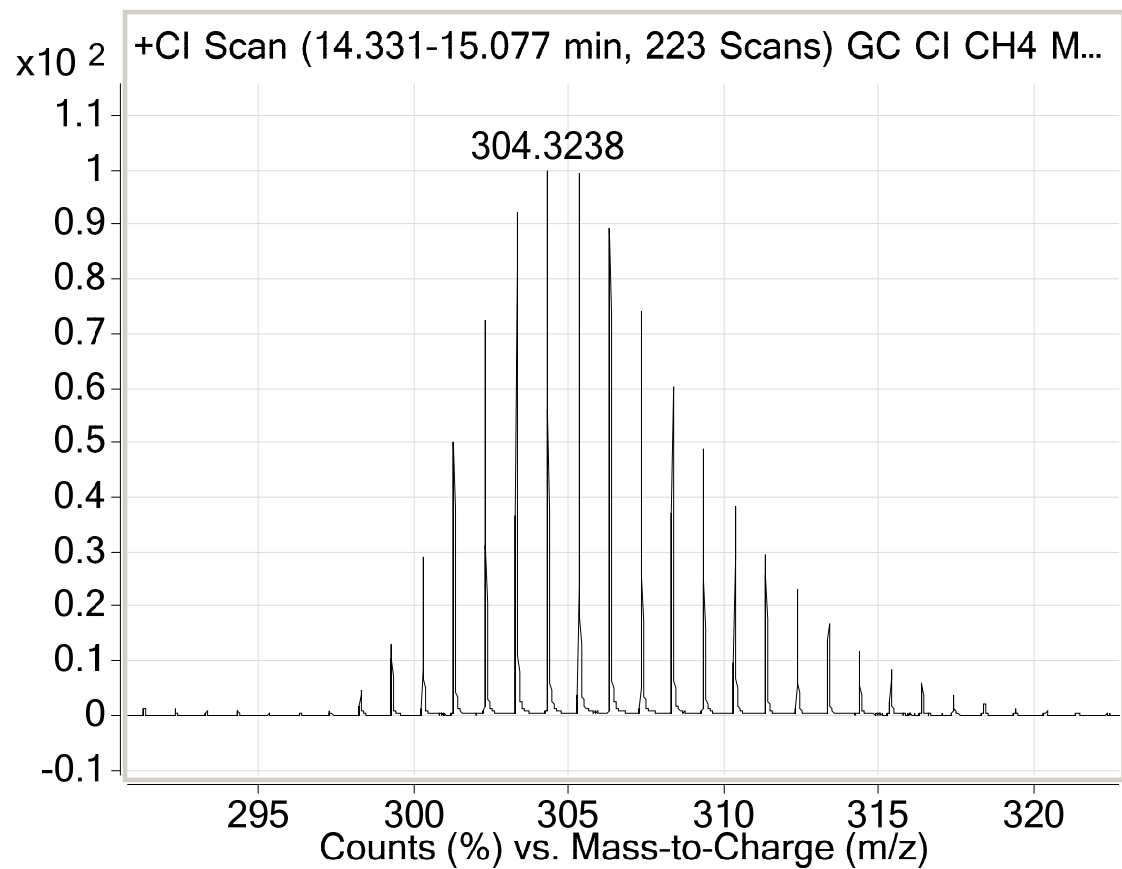




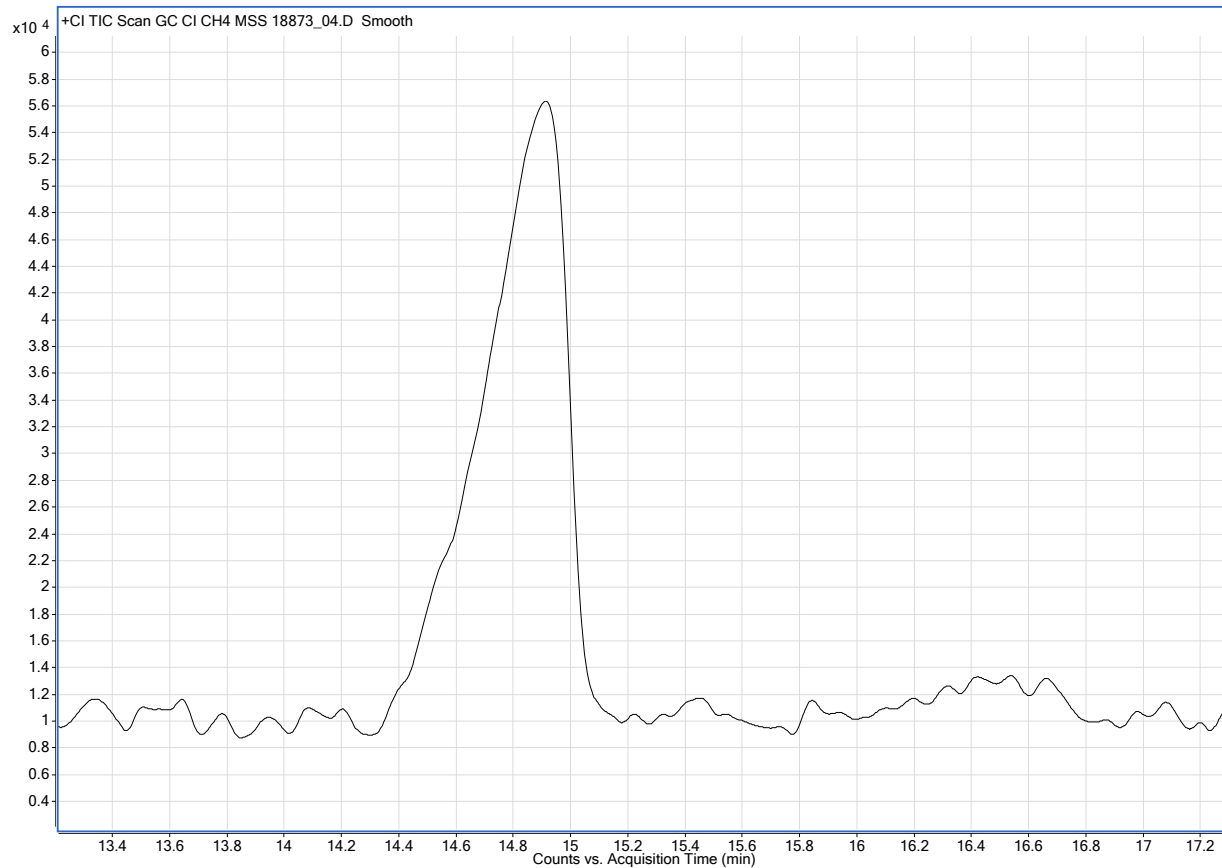
# GC Chromatogram first glance



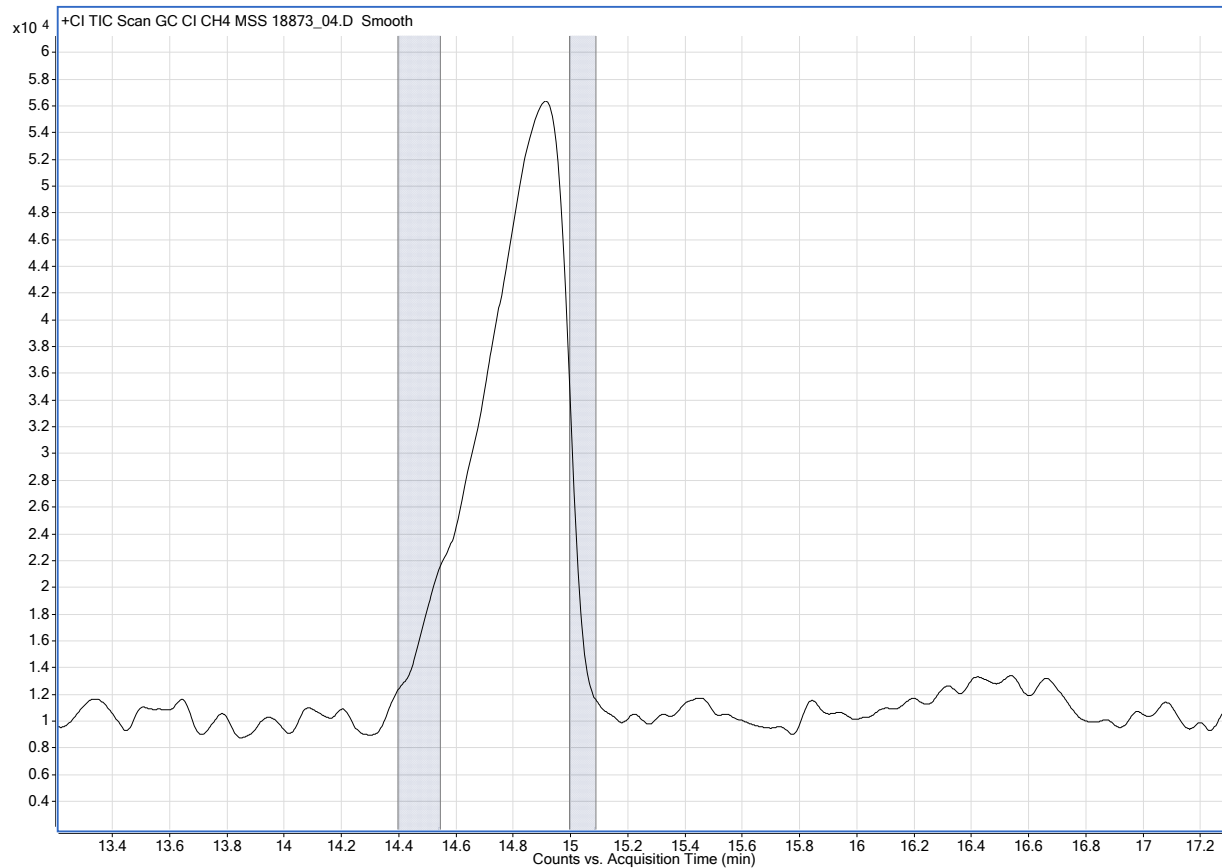
# GC Methane CI spectrum first glance



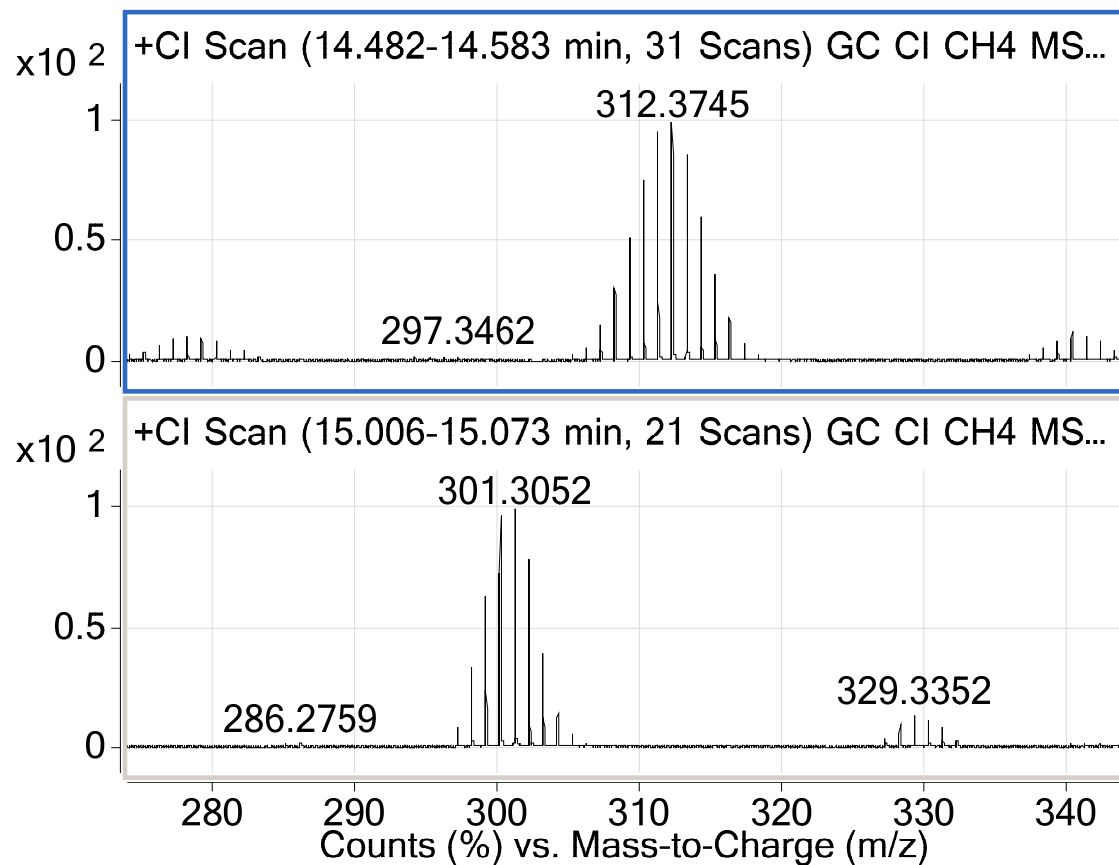
# GC Chromatogram



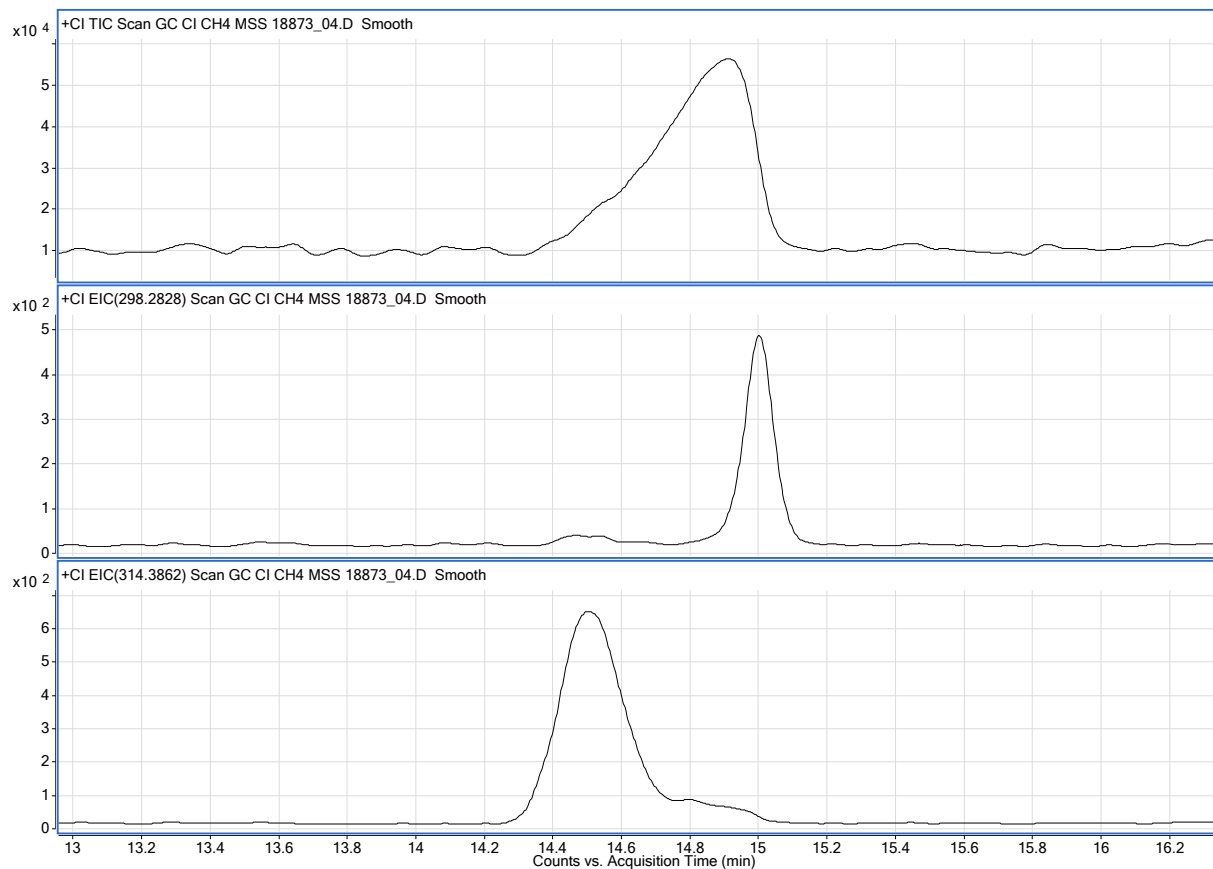
# GC Chromatogram



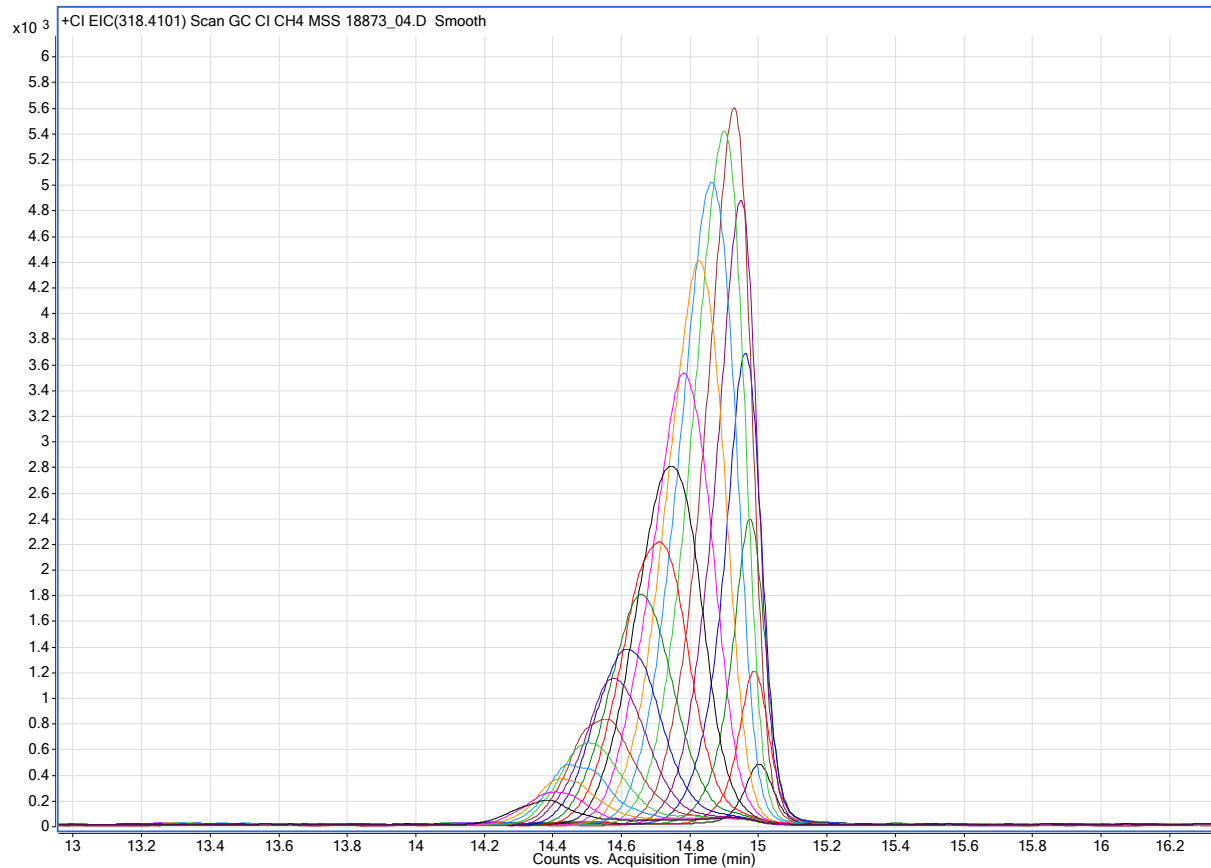
# GC Methane CI spectrum



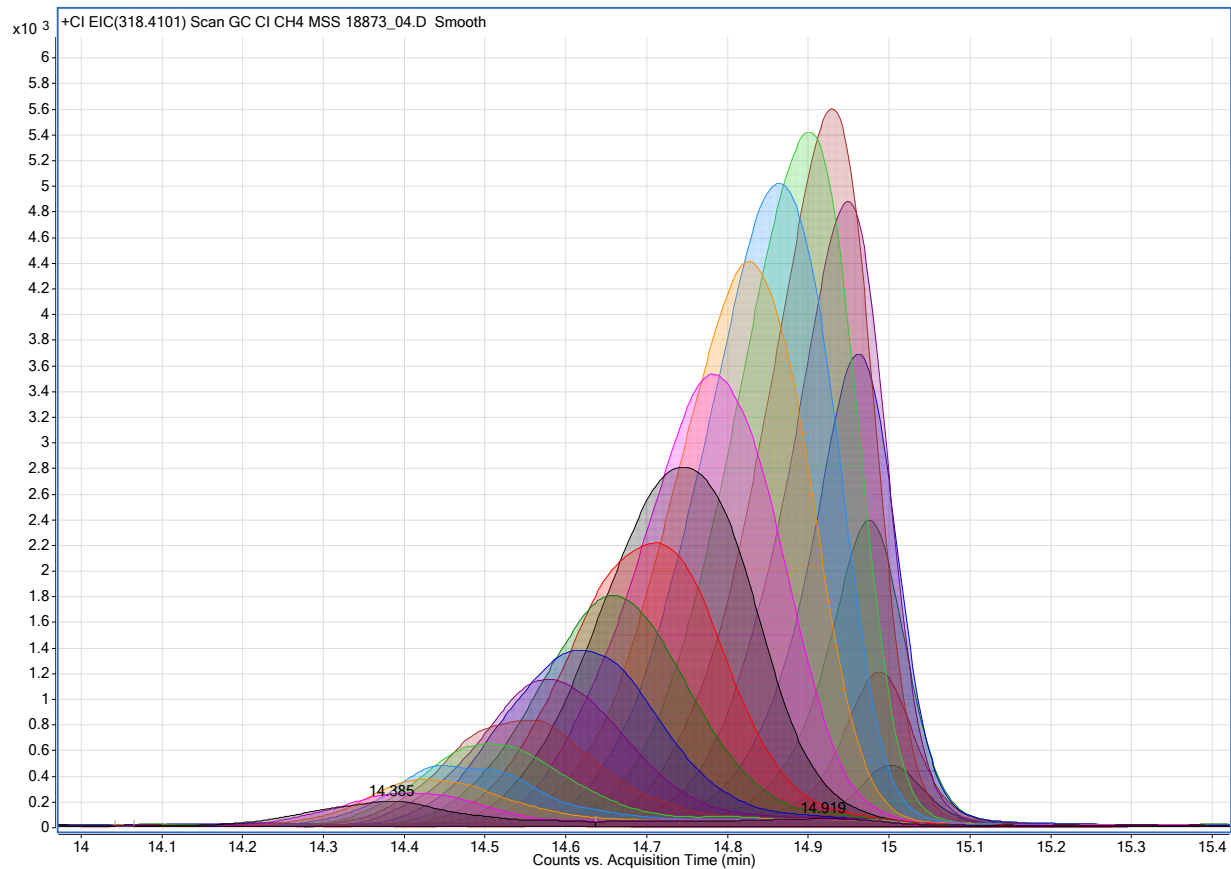
# GC Chromatogram extracted ion $m/z=298$ and $m/z=314$



# GC Chromatogram extracted ion *every isotopologue*

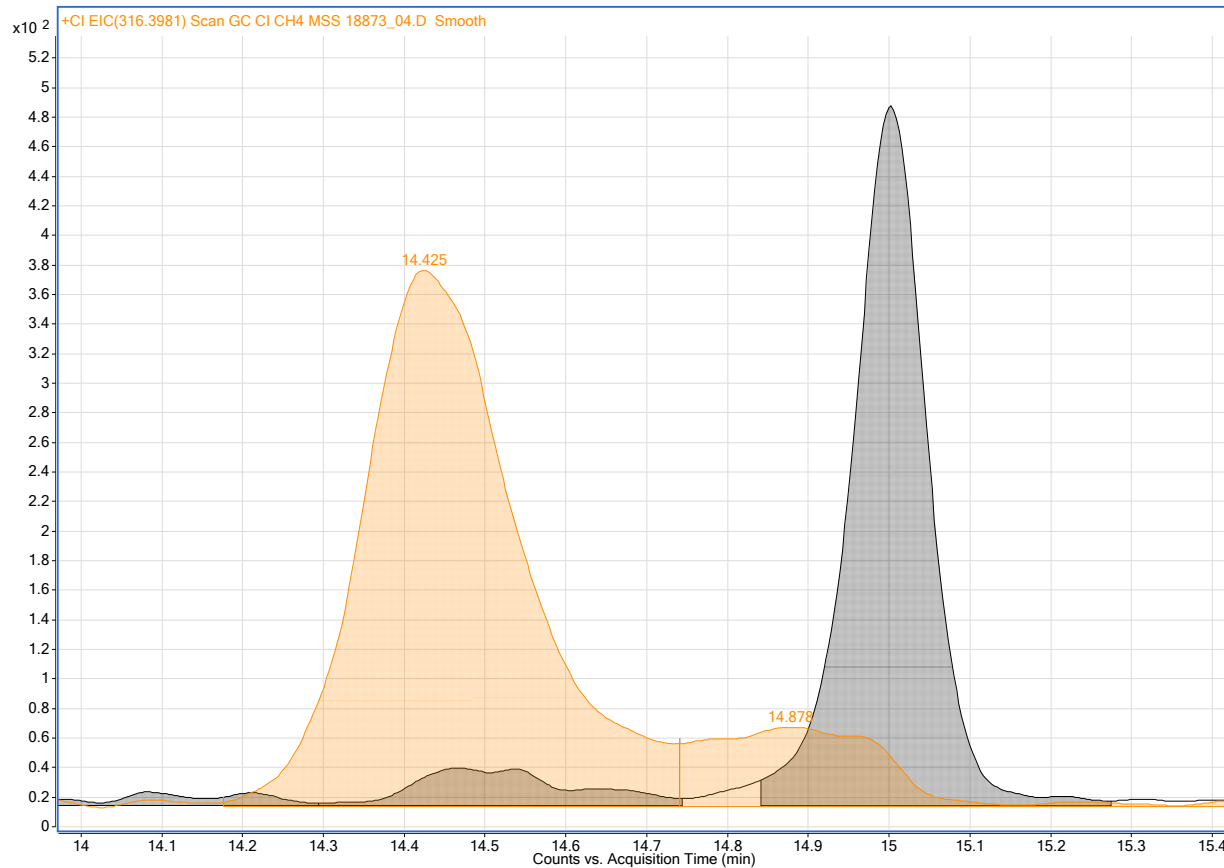


# GC Chromatogram extracted ion *every isotopologue*

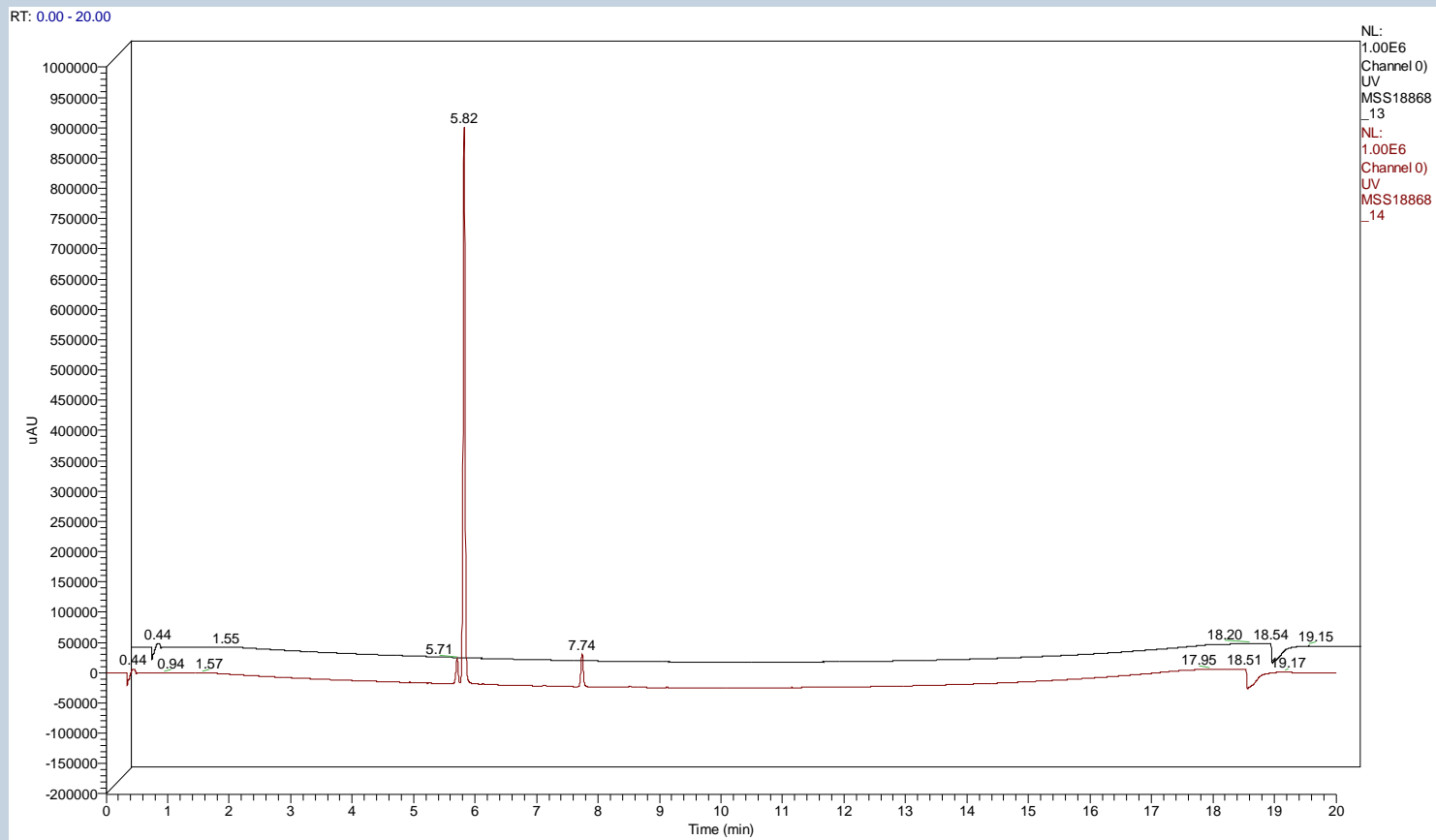




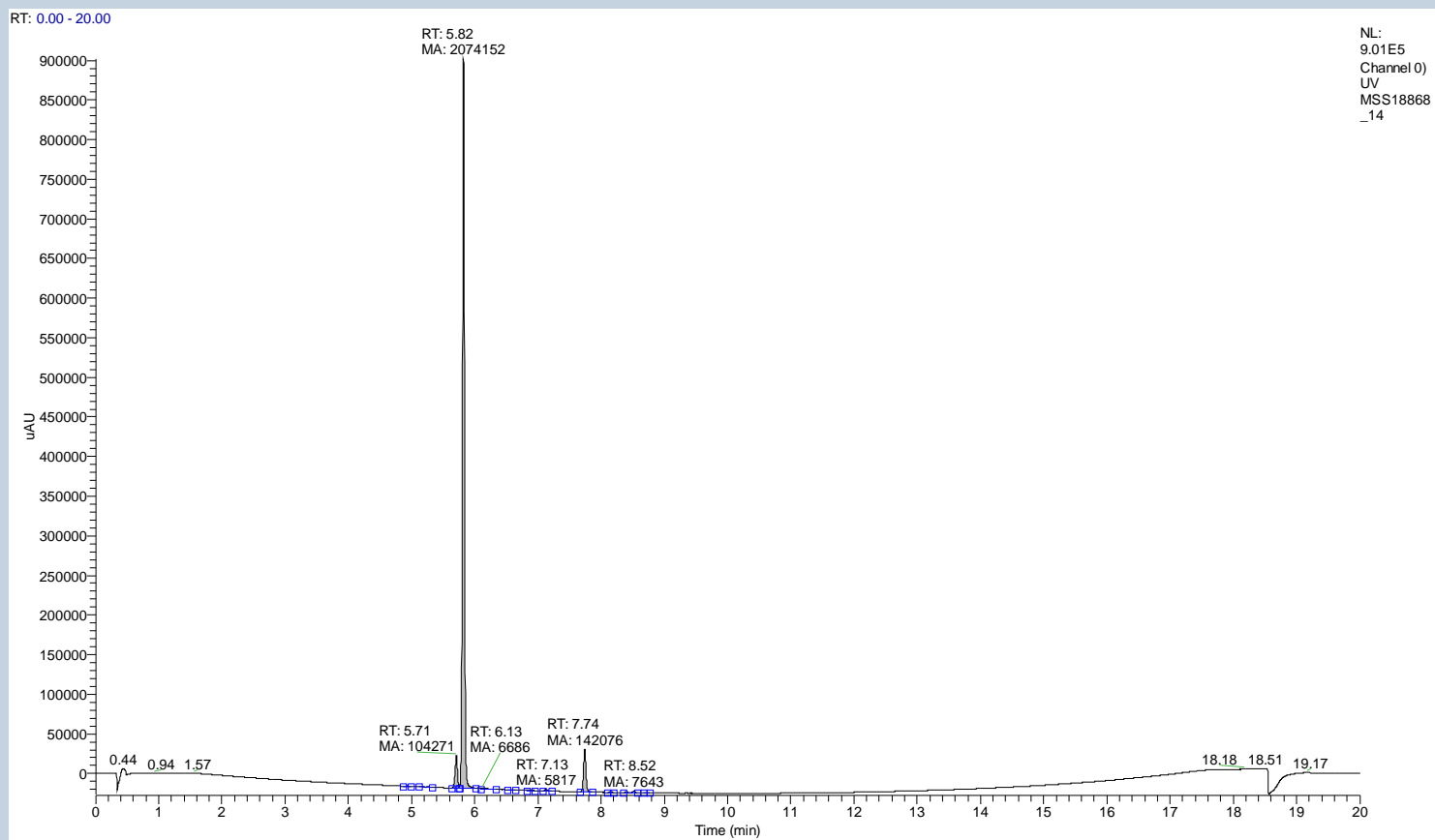
# GC Chromatogram extracted ion $m/z=298$ and $m/z=314$ peak width



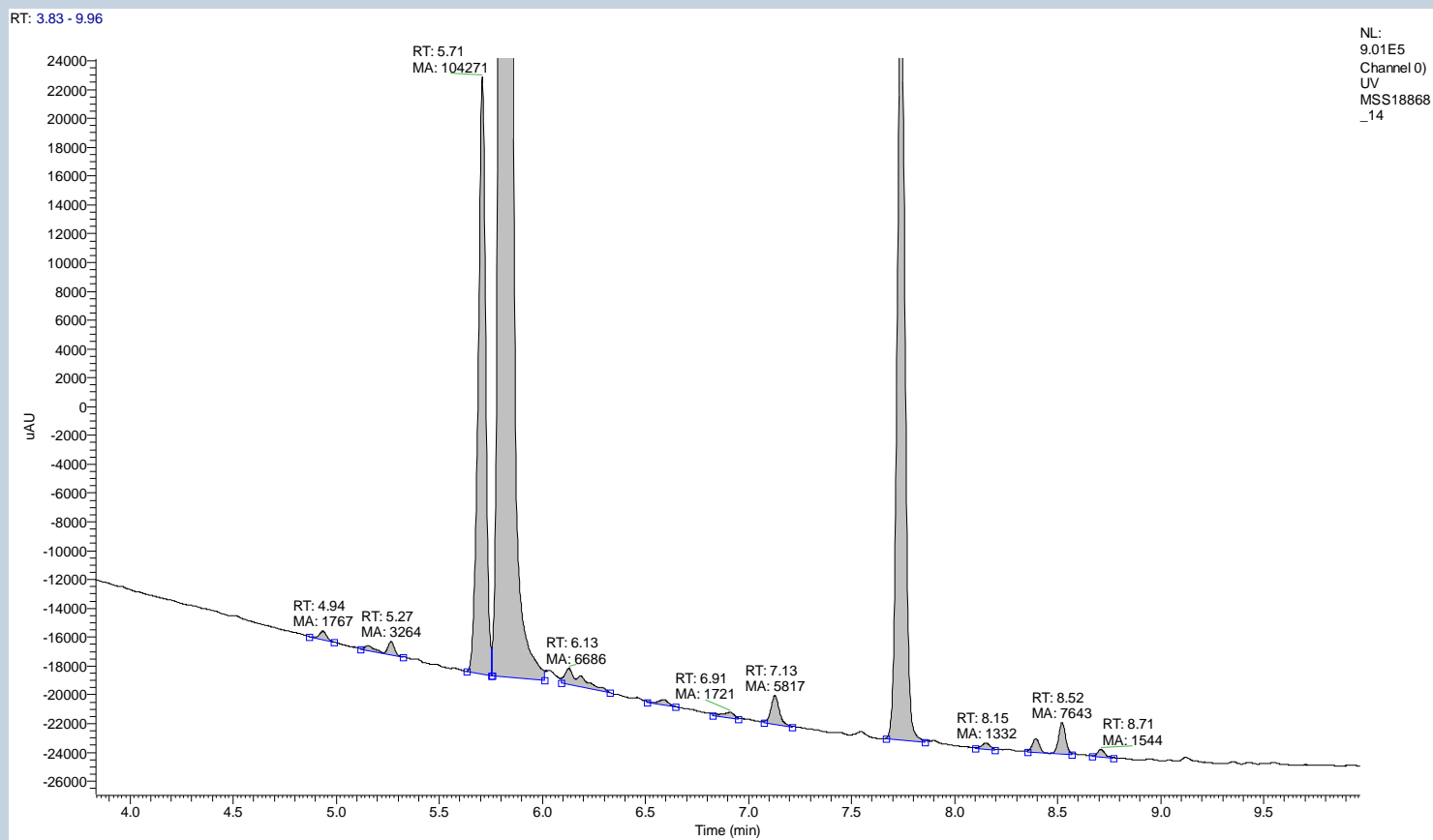
# “Purity” by LC-MS a can of worms



# UV Chromatogram at 322nm



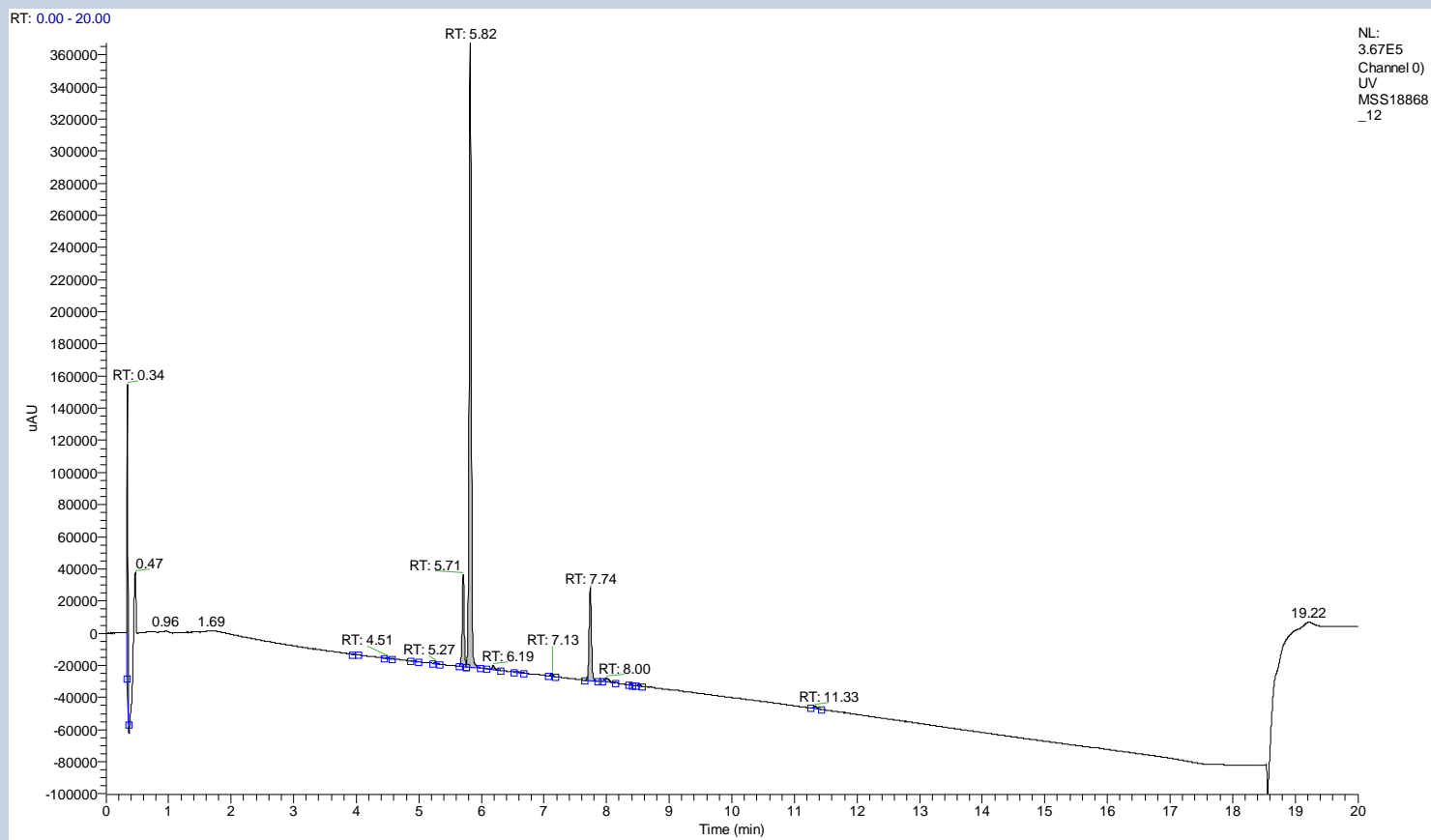
# UV Chromatogram at 322nm



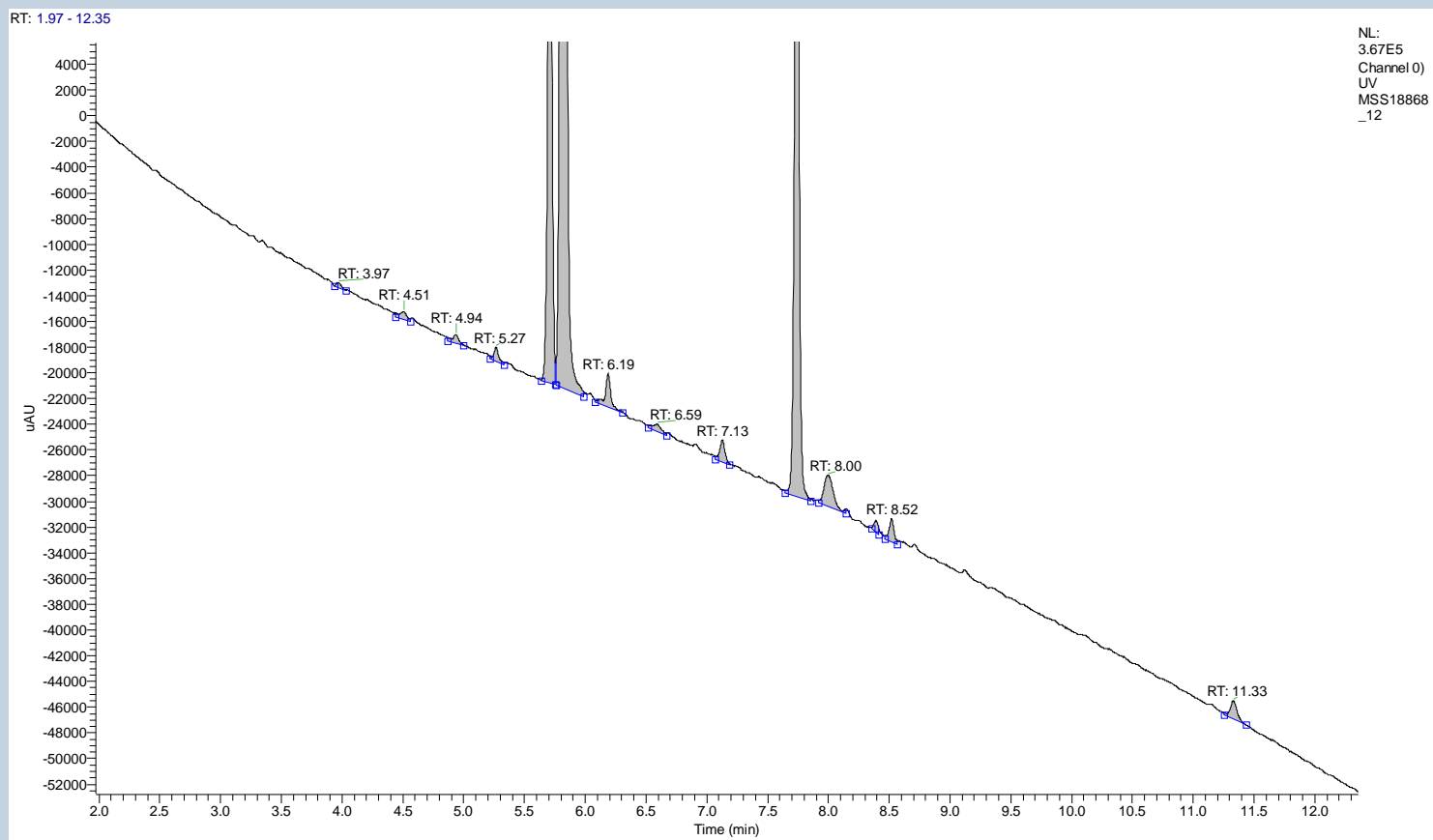
# Peak area table (322nm)

|                              |      |          |       |
|------------------------------|------|----------|-------|
| PEAK LIST                    |      |          |       |
| MSS18868_14.raw              |      |          |       |
| RT: 3.83 - 9.96              |      |          |       |
| Number of detected peaks: 12 |      |          |       |
| Apex RT                      | Area | %Area    |       |
|                              | 4.94 | 1766.772 | 0.08  |
|                              | 5.27 | 3264.045 | 0.14  |
|                              | 5.71 | 104270.7 | 4.43  |
|                              | 5.82 | 2074152  | 88.19 |
|                              | 6.13 | 6685.573 | 0.28  |
|                              | 6.59 | 1527.32  | 0.06  |
|                              | 6.91 | 1720.855 | 0.07  |
|                              | 7.13 | 5817.416 | 0.25  |
|                              | 7.74 | 142076.2 | 6.04  |
|                              | 8.15 | 1331.607 | 0.06  |
|                              | 8.52 | 7642.639 | 0.32  |
|                              | 8.71 | 1544.385 | 0.07  |

# UV Chromatogram at 240nm



# UV Chromatogram at 240nm

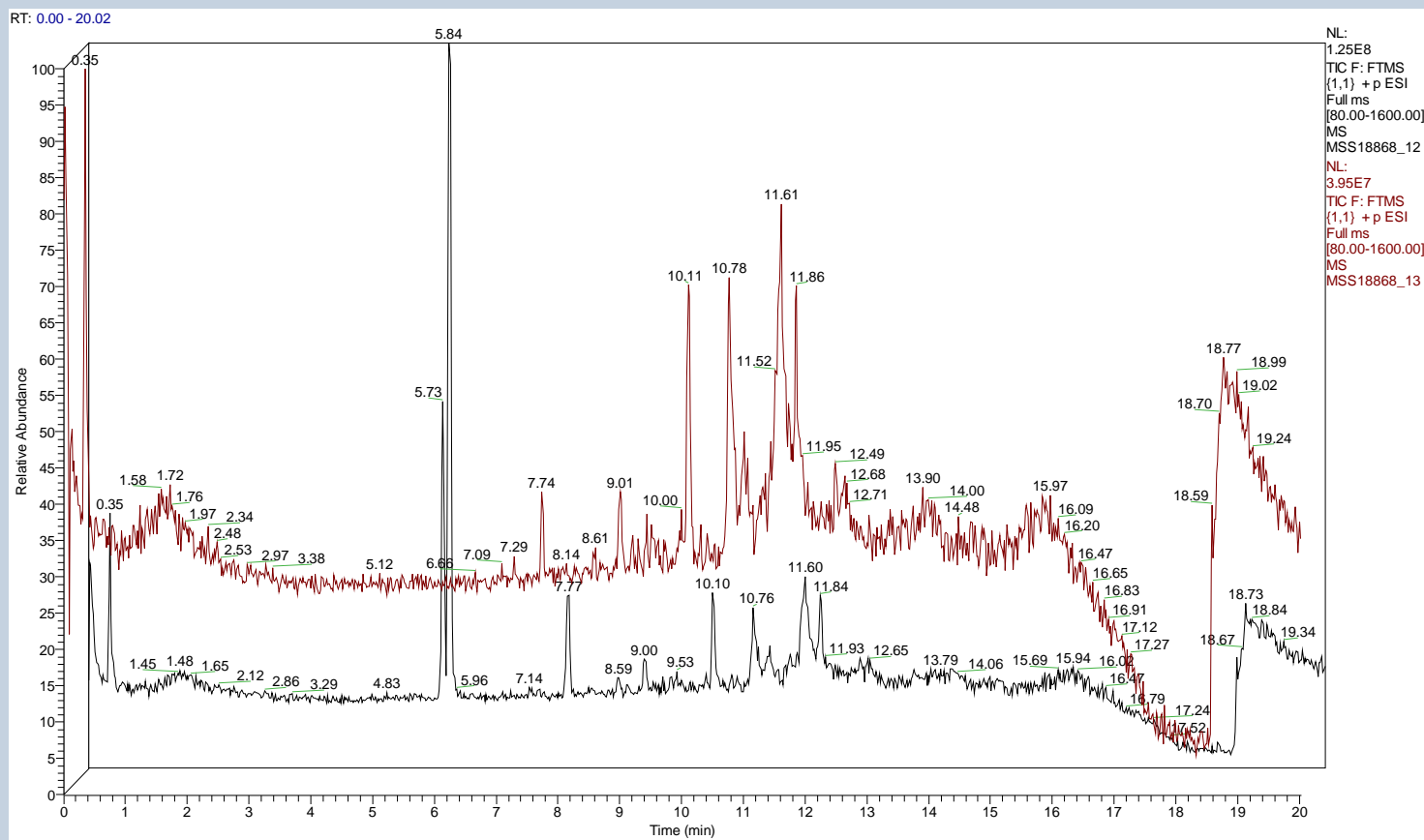


# Peak area table (240nm)

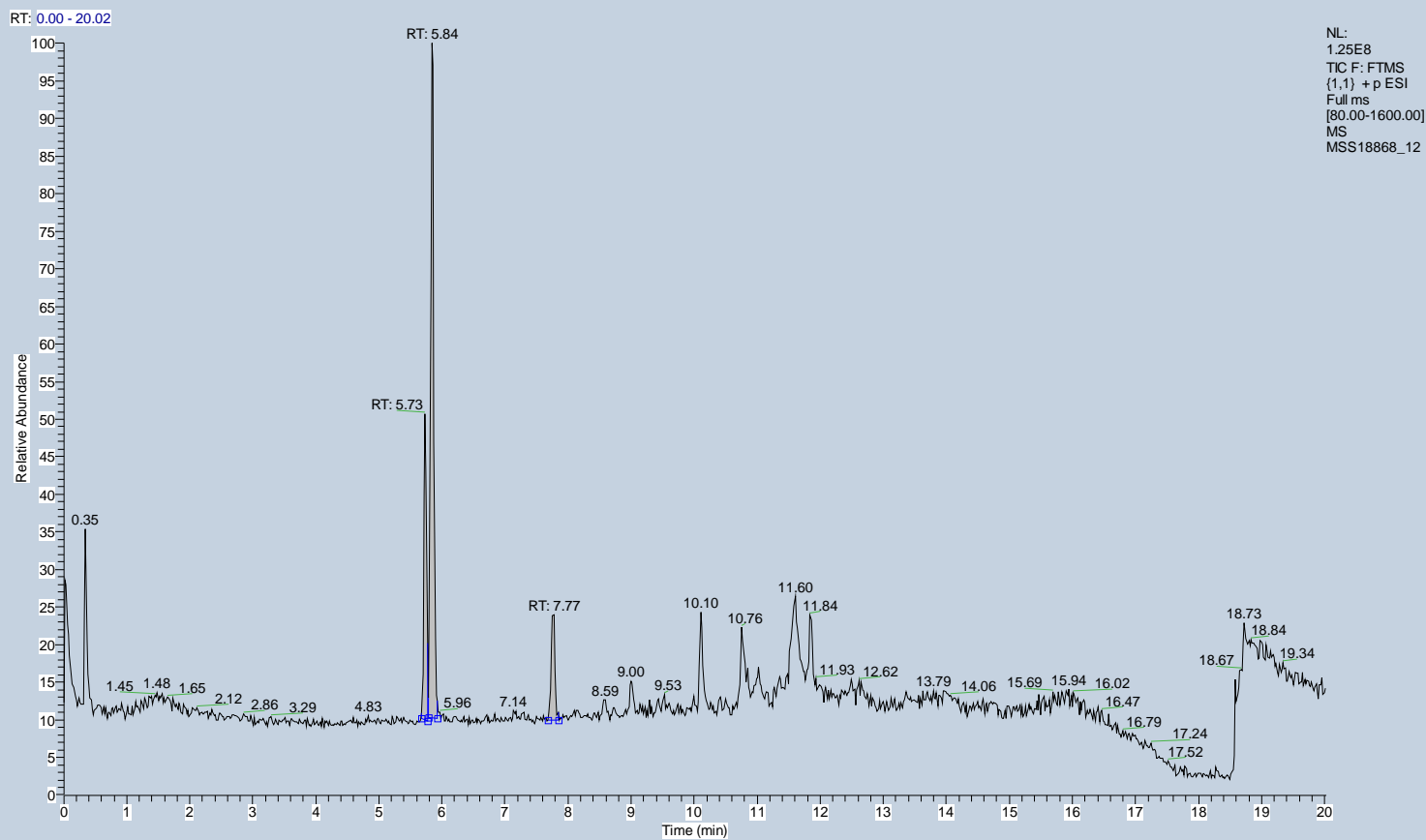
| PEAK LIST                    |       |            |       |
|------------------------------|-------|------------|-------|
| MSS18868_12.raw              |       |            |       |
| RT: 1.97 - 12.35             |       |            |       |
| Number of detected peaks: 15 |       |            |       |
| Apex RT                      | Area  | %Area      |       |
|                              | 0.34  | 190741.58  | 13.47 |
|                              | 3.97  | 1165.396   | 0.08  |
|                              | 4.51  | 2693.562   | 0.19  |
|                              | 4.94  | 2527.462   | 0.18  |
|                              | 5.27  | 3057.909   | 0.22  |
|                              | 5.71  | 129118.483 | 9.12  |
|                              | 5.82  | 892036.507 | 63.01 |
|                              | 6.19  | 8201.617   | 0.58  |
|                              | 6.59  | 2892.024   | 0.2   |
|                              | 7.13  | 5260.836   | 0.37  |
|                              | 7.74  | 151938.723 | 10.73 |
|                              | 8     | 13224.811  | 0.93  |
|                              | 8.39  | 1976.459   | 0.14  |
|                              | 8.52  | 5189.208   | 0.37  |
|                              | 11.33 | 5682.965   | 0.4   |



# Mass chromatogram TIC pos



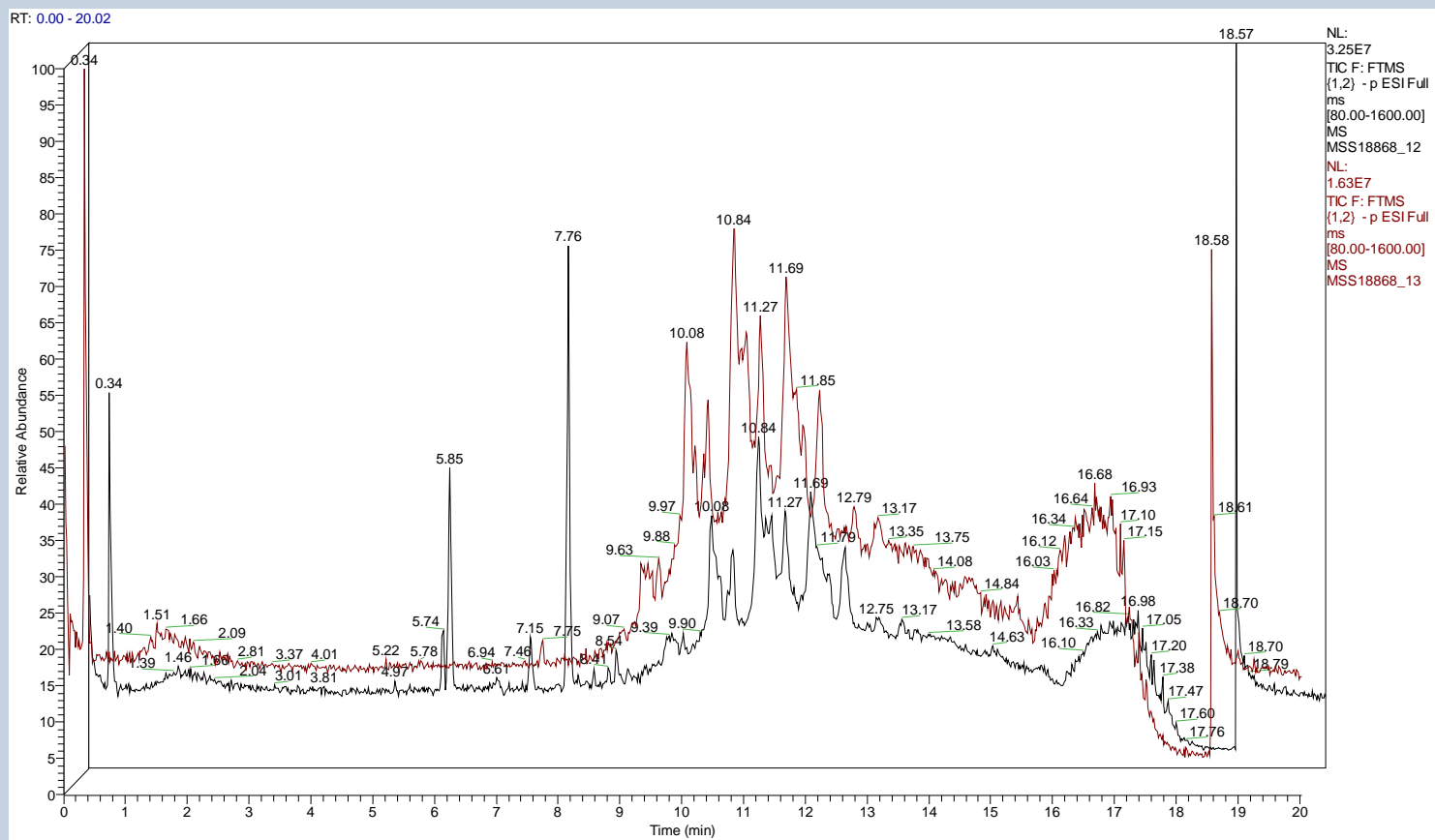
# Mass chromatogram TIC pos



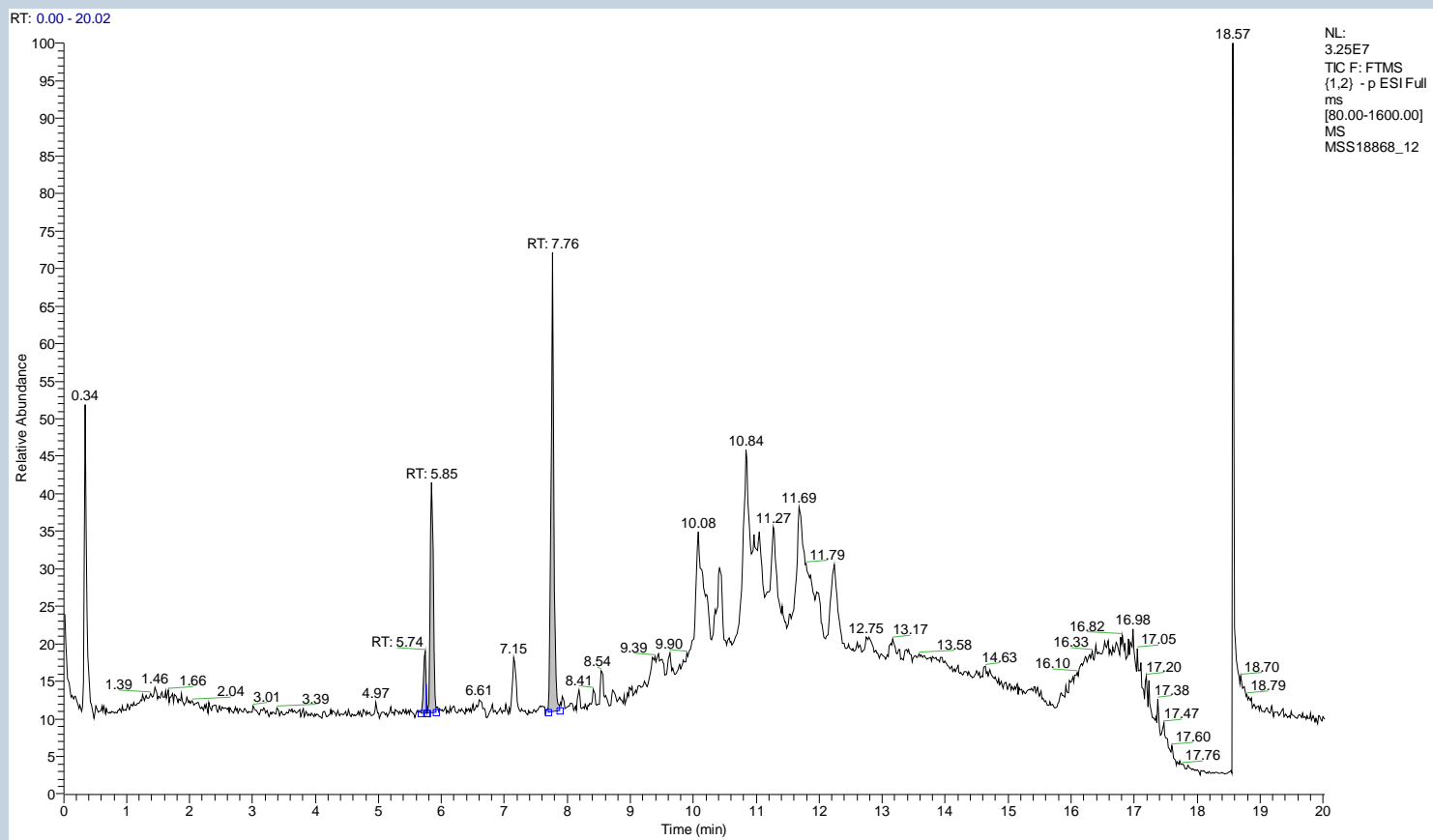
# Peak area table (positive TIC)

|                             |             |       |  |
|-----------------------------|-------------|-------|--|
| PEAK LIST                   |             |       |  |
| MSS18868_12.raw             |             |       |  |
| RT: 0.00 - 20.02            |             |       |  |
| Number of detected peaks: 3 |             |       |  |
| Apex RT                     | Area        | %Area |  |
| 5.73                        | 153838800.8 | 24.35 |  |
| 5.84                        | 404375386.6 | 64.02 |  |
| 7.77                        | 73471112.5  | 11.63 |  |

# Mass chromatogram TIC neg



# Mass chromatogram TIC neg



# Peak area table (negative TIC)

|                             |             |       |  |
|-----------------------------|-------------|-------|--|
| PEAK LIST                   |             |       |  |
| MSS18868_12.raw             |             |       |  |
| RT: 0.00 - 20.02            |             |       |  |
| Number of detected peaks: 3 |             |       |  |
| Apex RT                     | Area        | %Area |  |
| 5.74                        | 6983375.211 | 6.69  |  |
| 5.85                        | 34258905.82 | 32.8  |  |
| 7.76                        | 63216928.44 | 60.52 |  |

# Round up

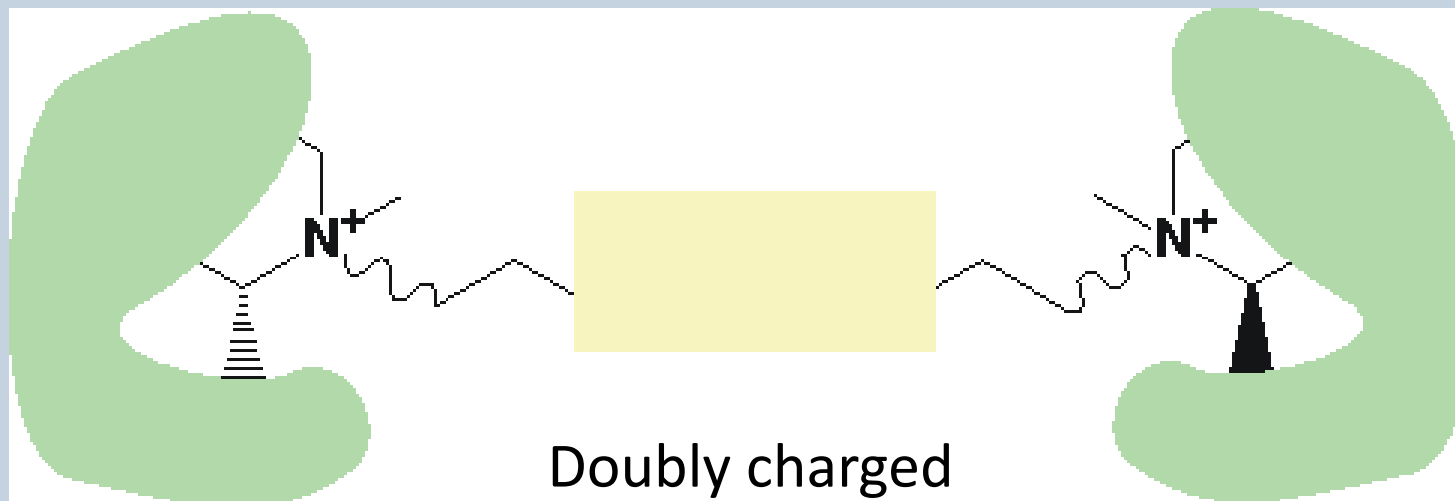
- Mass spectrometry and separation science is a great combo
- Many problems have be faced and solved
  - Check the literature and commercial application notes!
- Many more problems to be investigated

# Extra examples



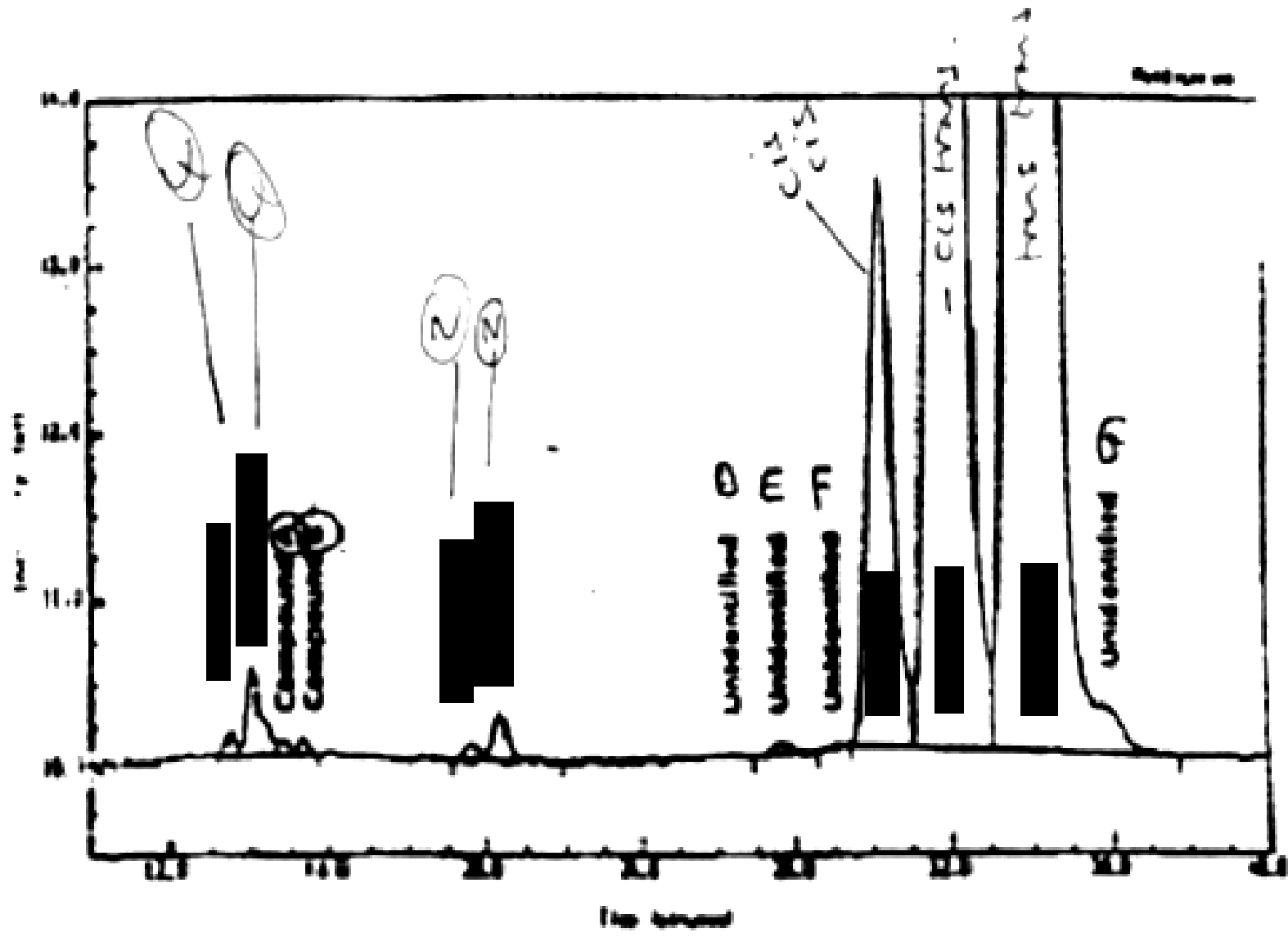
## Example 2

- Legacy compound
- Wanted to reformulate/new therapeutic area



Doubly charged  
4 chiral centres  
RMM 1014  
*m/z* 514

# Example Sample Chromatogram

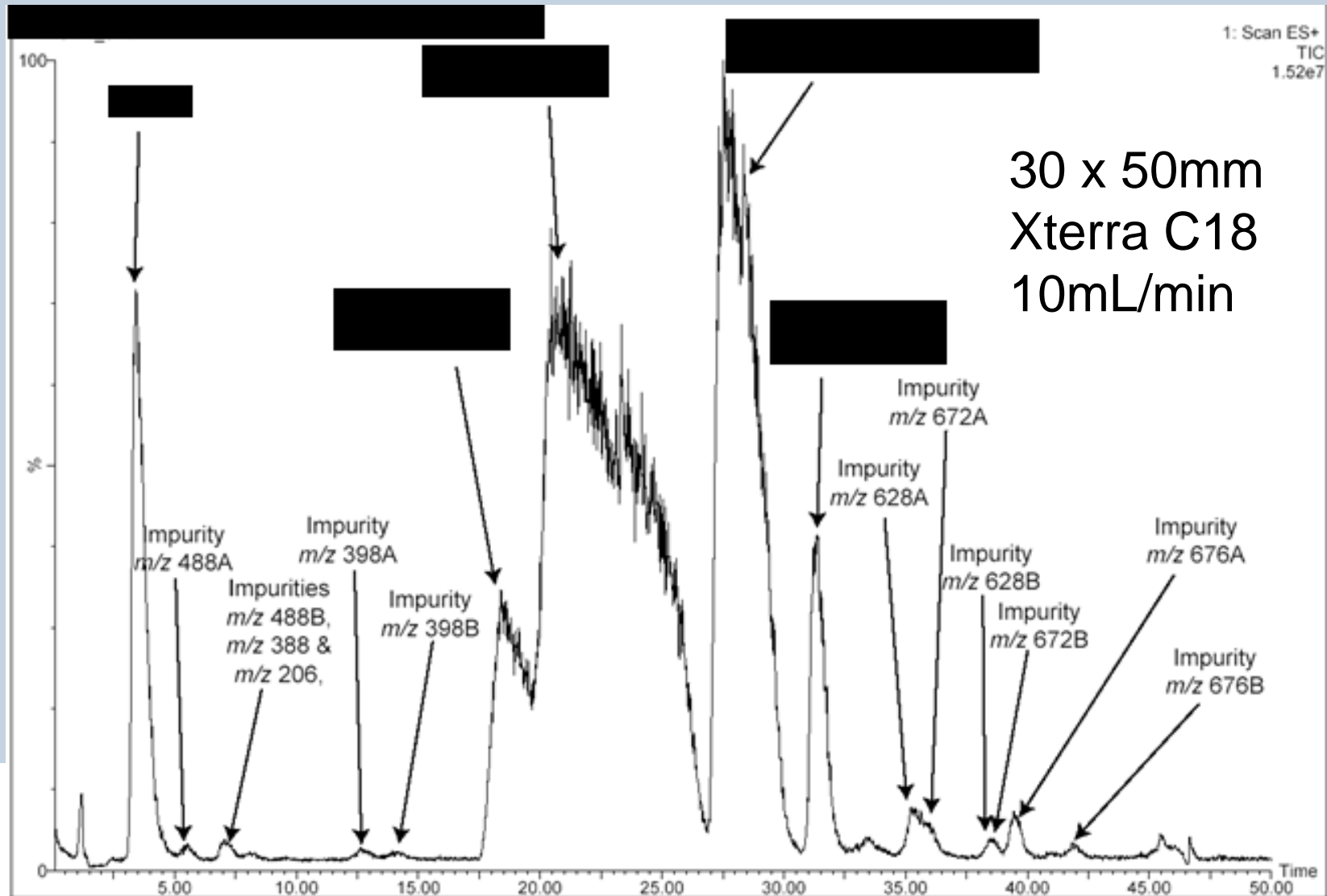


## Example 2

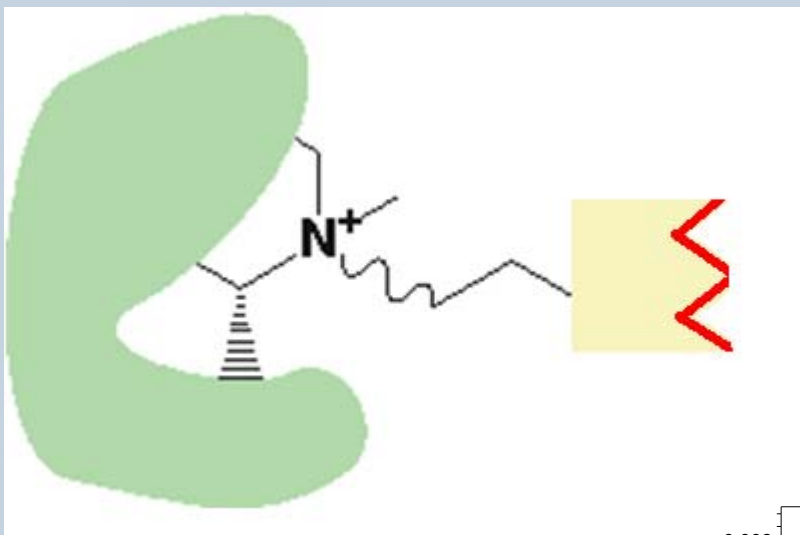
- The original LC method
- 0.4 mL.min<sup>-1</sup>
- 84 acetonitrile to 16 water
  - + 1% conc Phosphoric acid
- Whatman Partisil-5, 25cm x 4.6mm
- Bare silica

# Example 2

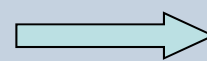
## Prep Reversed phase LC



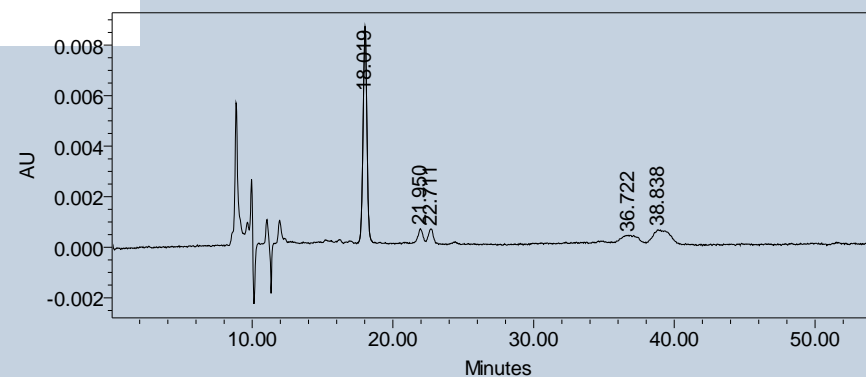
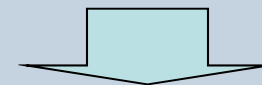
# Example 2



NMR



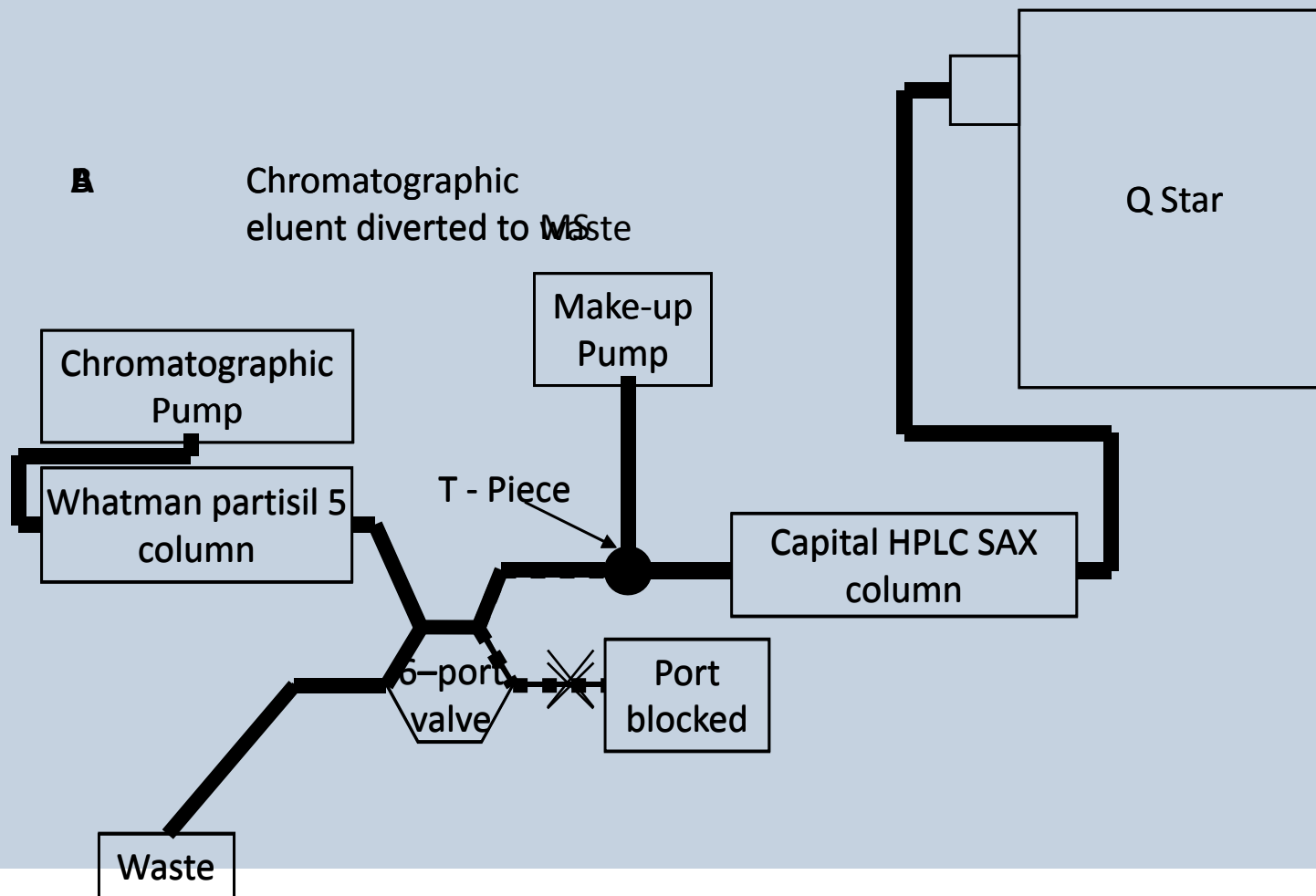
Original LC



One of early eluting imp



# Example 2



# Example 2

