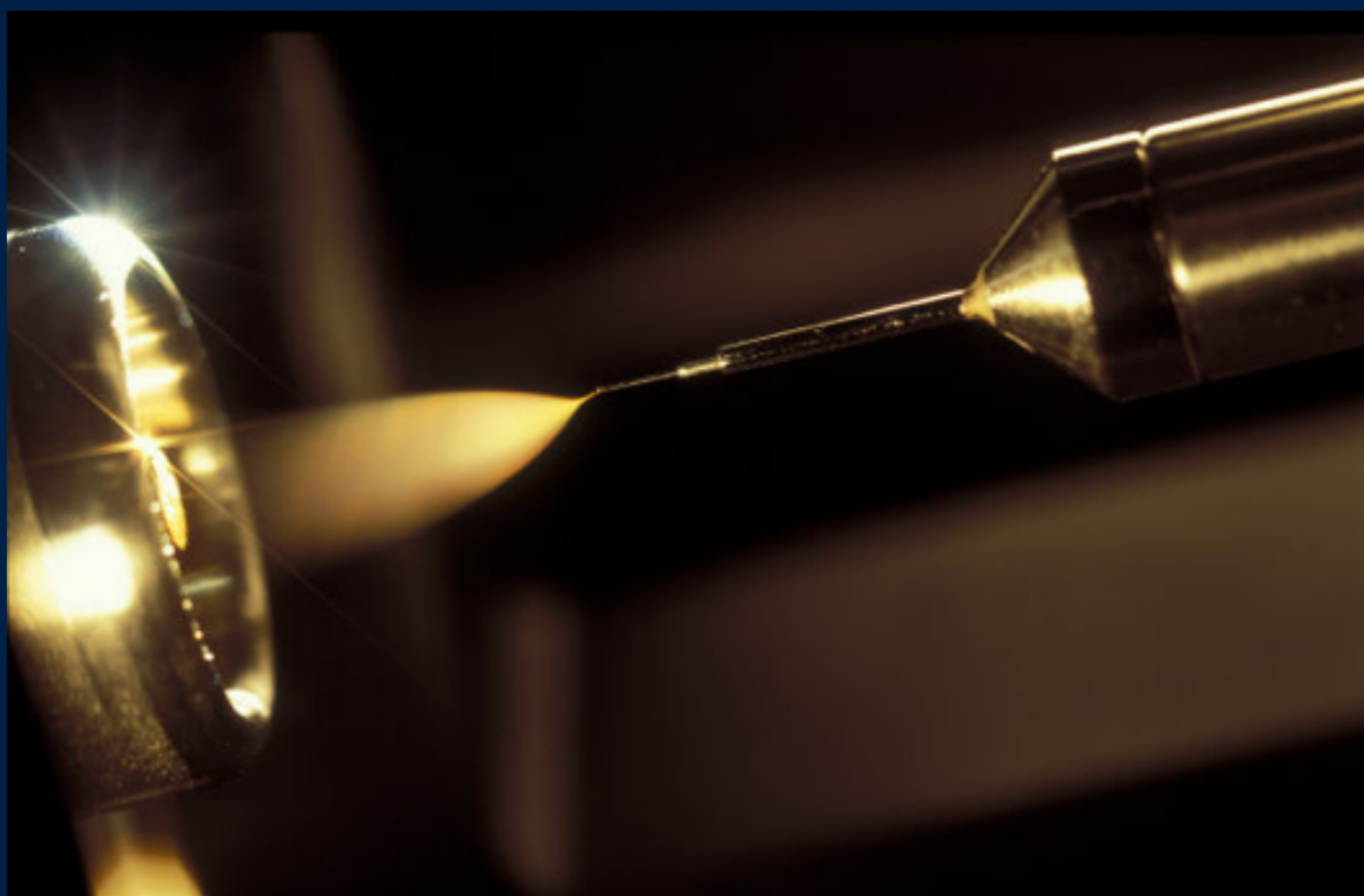


DEPARTMENT OF CHEMISTRY

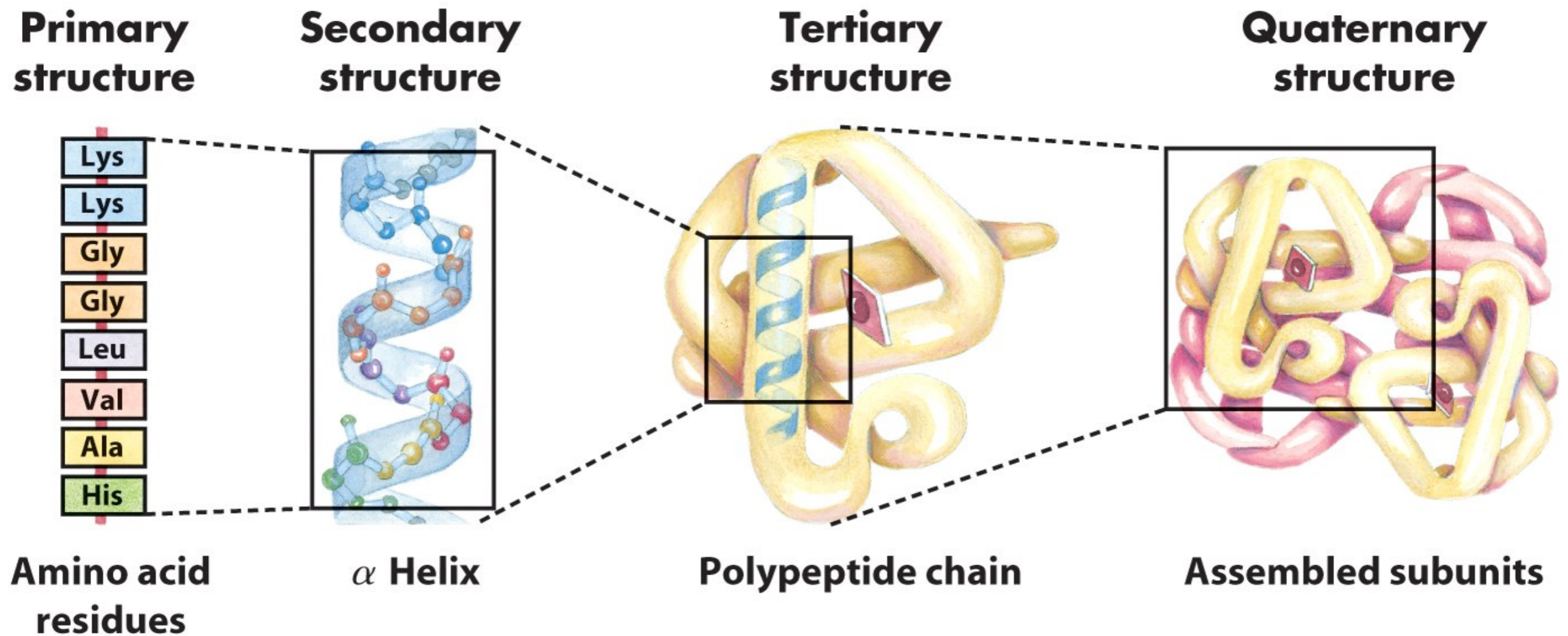
GRADUATE COURSE IN MASS SPECTROMETRY: LECTURE 8

Mass spectrometry for biophysics and structural biology



Professor Justin Benesch, 23rd November 2016

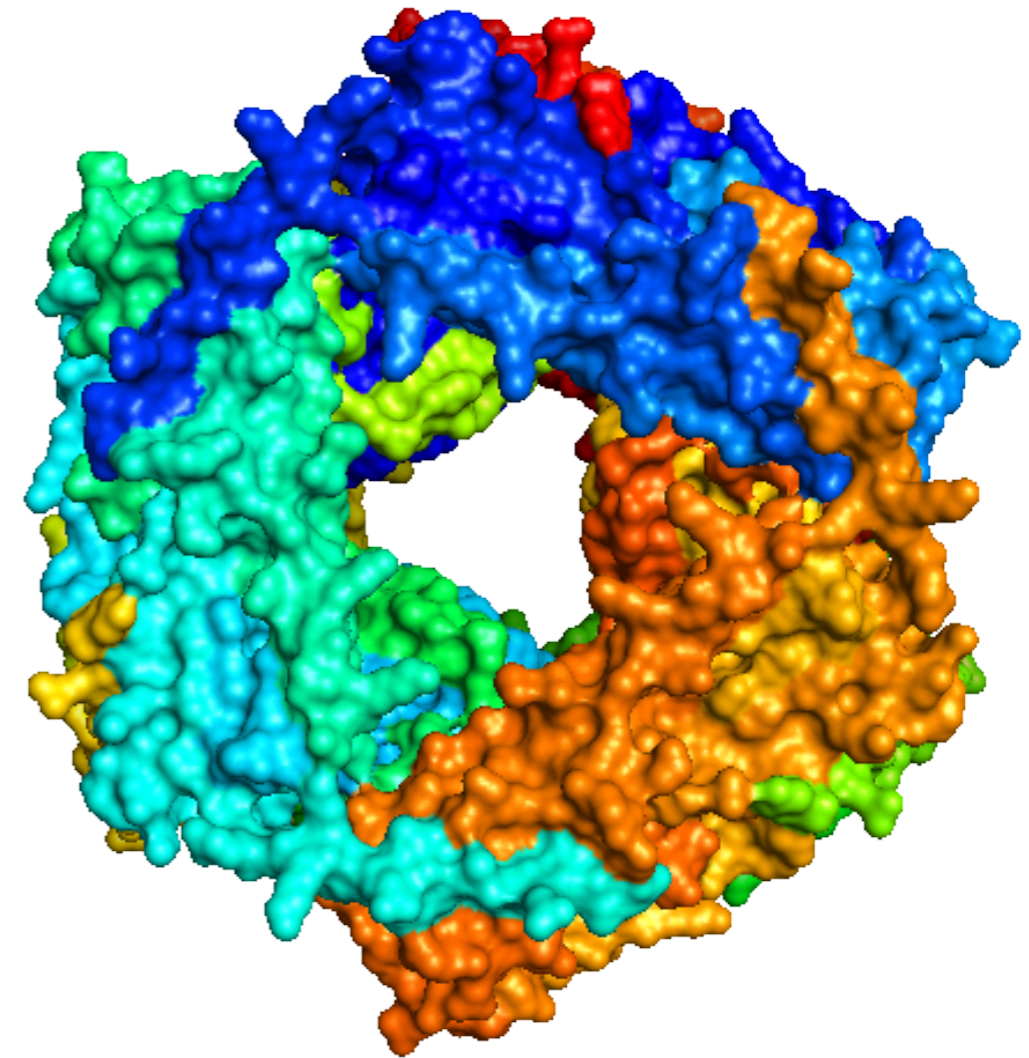
Levels of protein structure



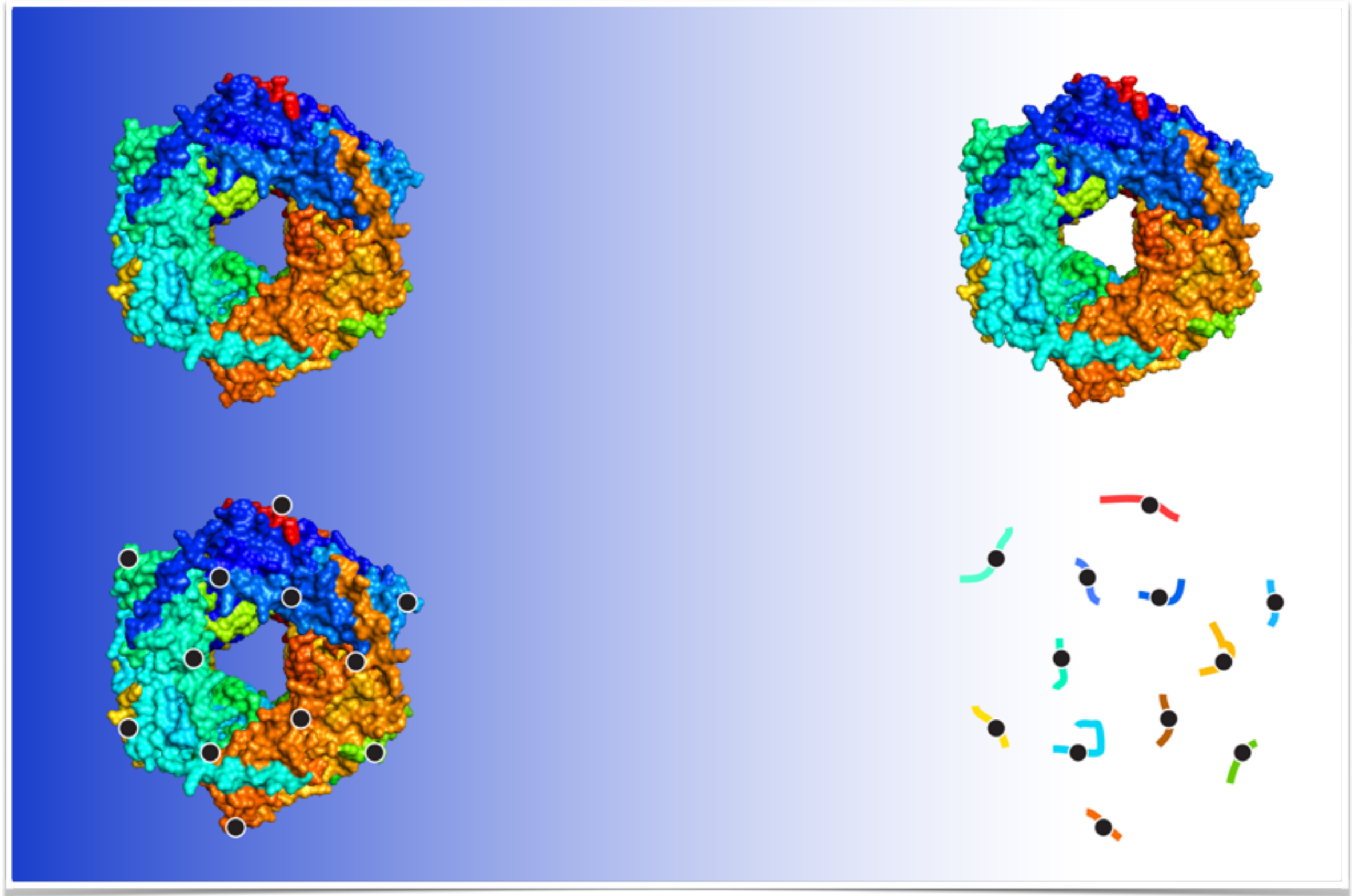
- Mass spectrometry can inform on all levels of protein structure

What do we want to know?

- Structure
 - Stoichiometry, connectivity, 3D topology, conformations, atomic coordinates
- Thermodynamics
 - Interface and interaction strengths, stability
- Kinetics
 - Rate constants, activation energies

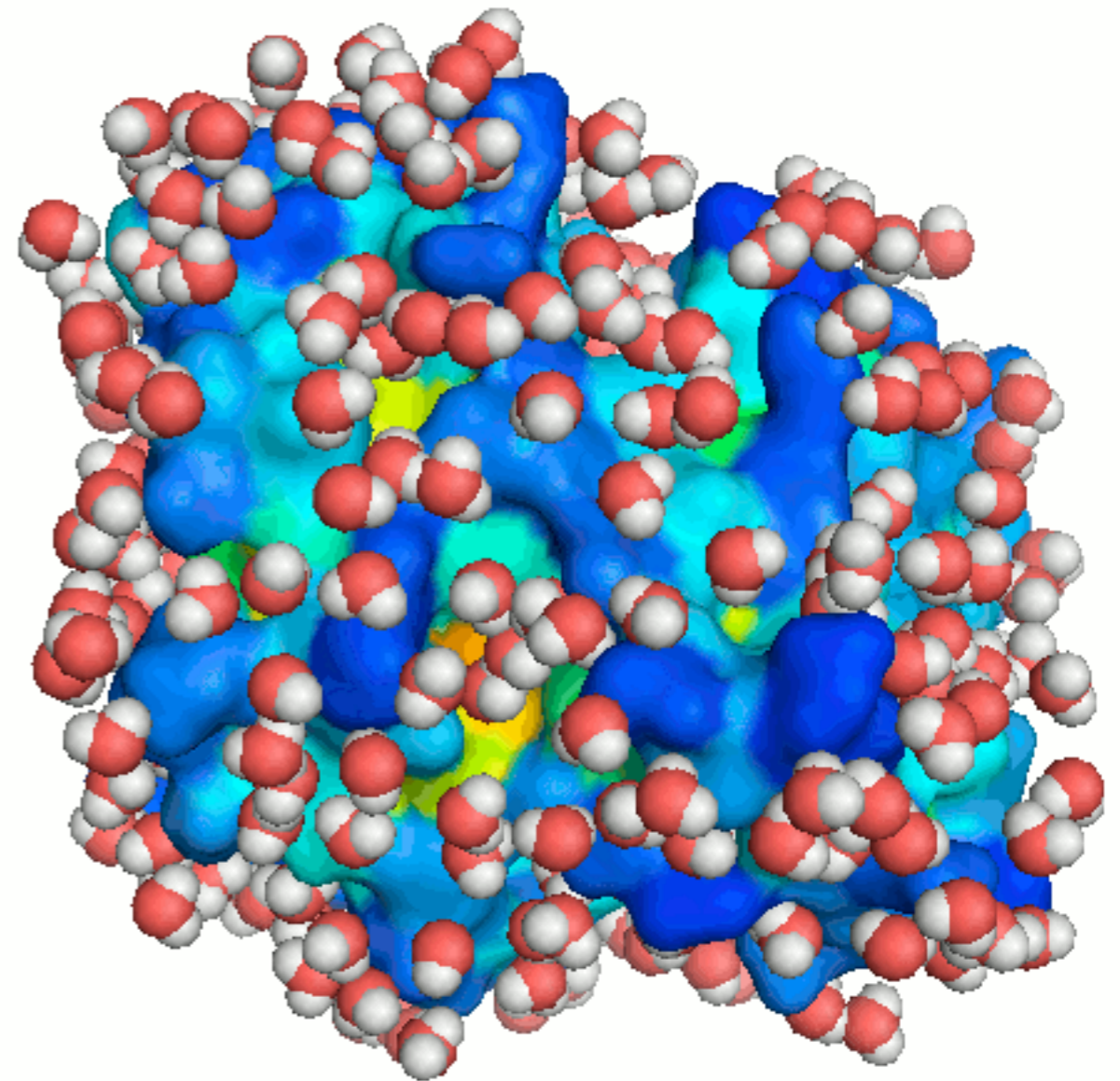


Two strategies - native or labelling

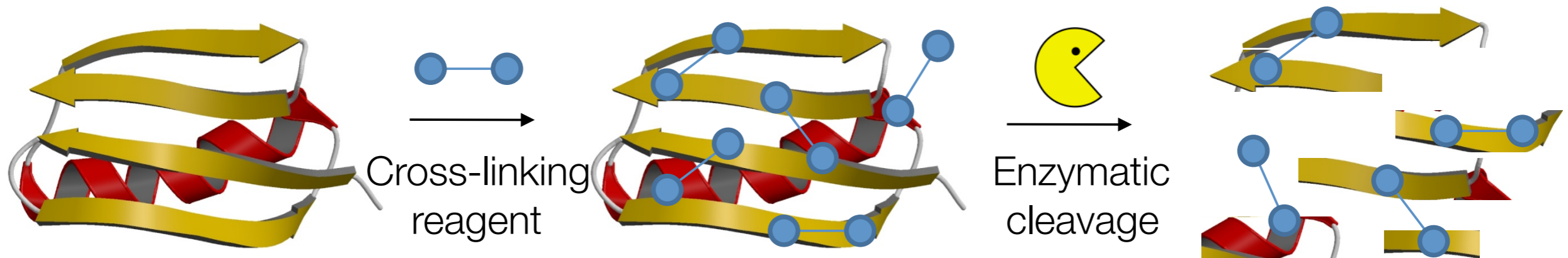


Labelling technologies with MS

- Label protein, and use MS to localise the individual labels
- Can interrogate the ‘global’ and ‘local’ levels: i.e. protein or peptide level
- Peptide level accessed by enzymolysis of protein, and further interrogation with tandem MS
- Labels reveal solvent accessibility and connectivity
- Provide means of probing protein structure and dynamic fluctuations

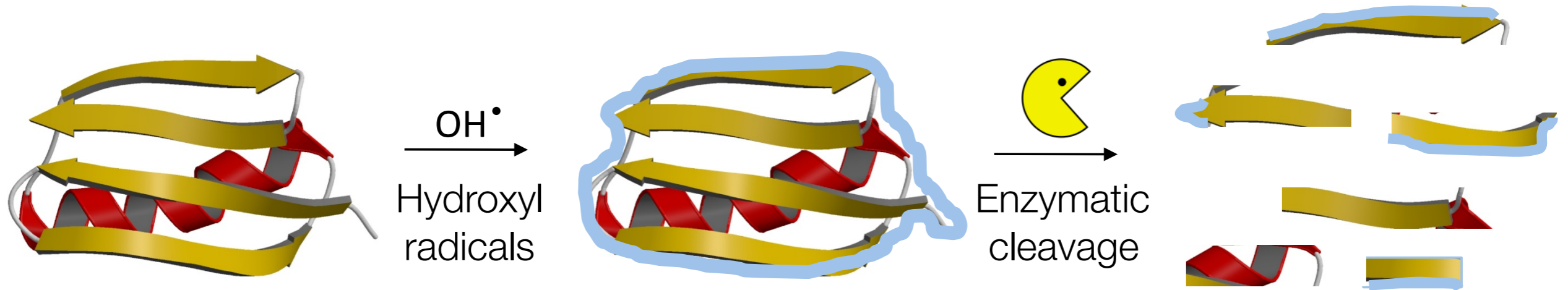


Cross-linking mass spectrometry



- Cross-linker forms covalent bonds between amino acids with appropriate functionality
- Peptide masses (and fragments) are interrogated to localise cross-linker
- Intra- and inter-protein cross-links can be formed
- Cross-links can be used to determine connectivity, and as a spatial restraint for modelling protein structures
- Problems centre on decreased detection efficiency of cross-linked peptides, kinetic effects, and interpretation of observed links

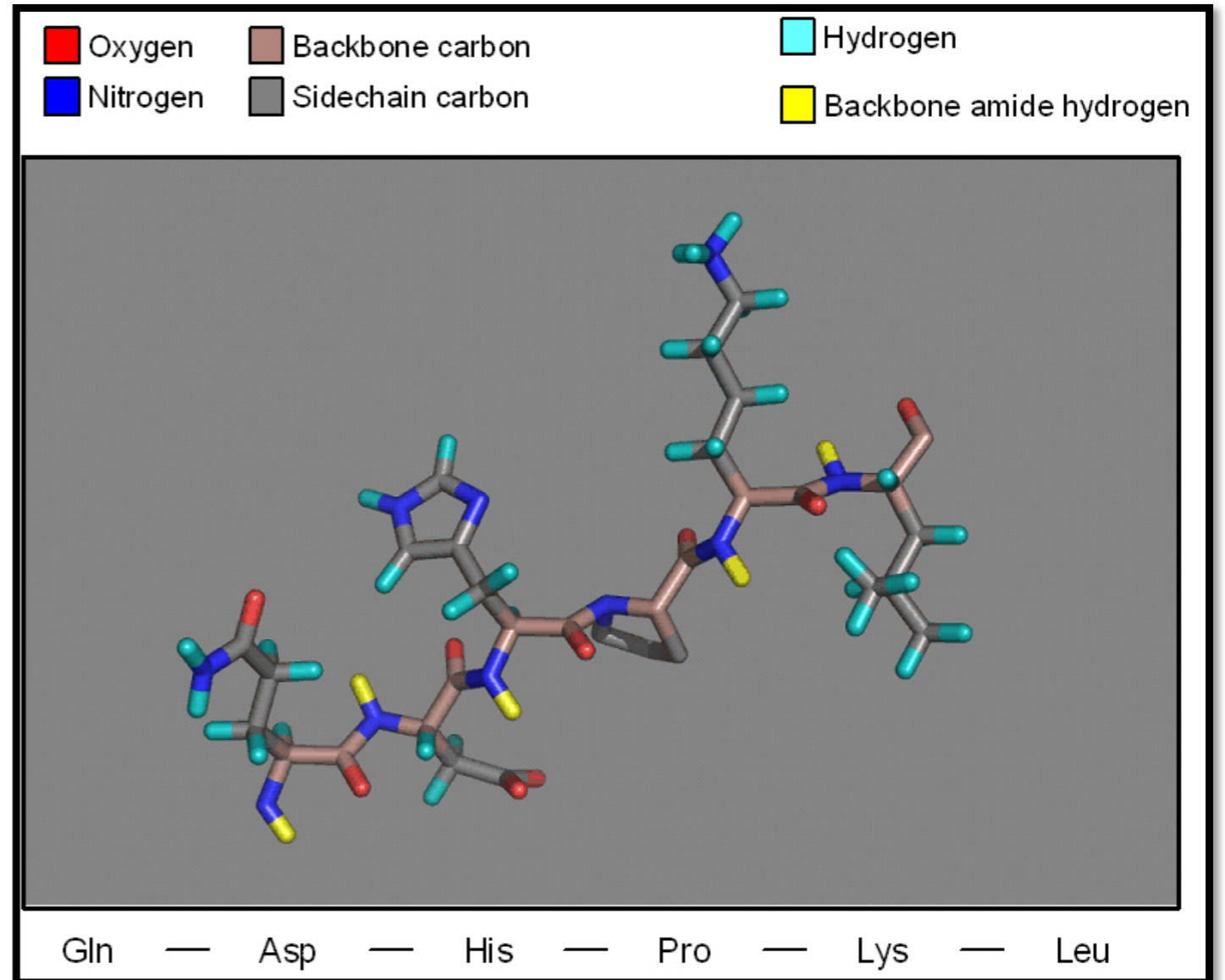
Oxidative foot-printing



- Solvent accessible amino acid side chains of the protein are oxidised
- Peptide masses (and fragments) are interrogated to localise oxidation sites
- Comparing data from proteins in complex and in isolation allows the determination of interface sites
- Problems centre on data analysis and differential intrinsic reactivity

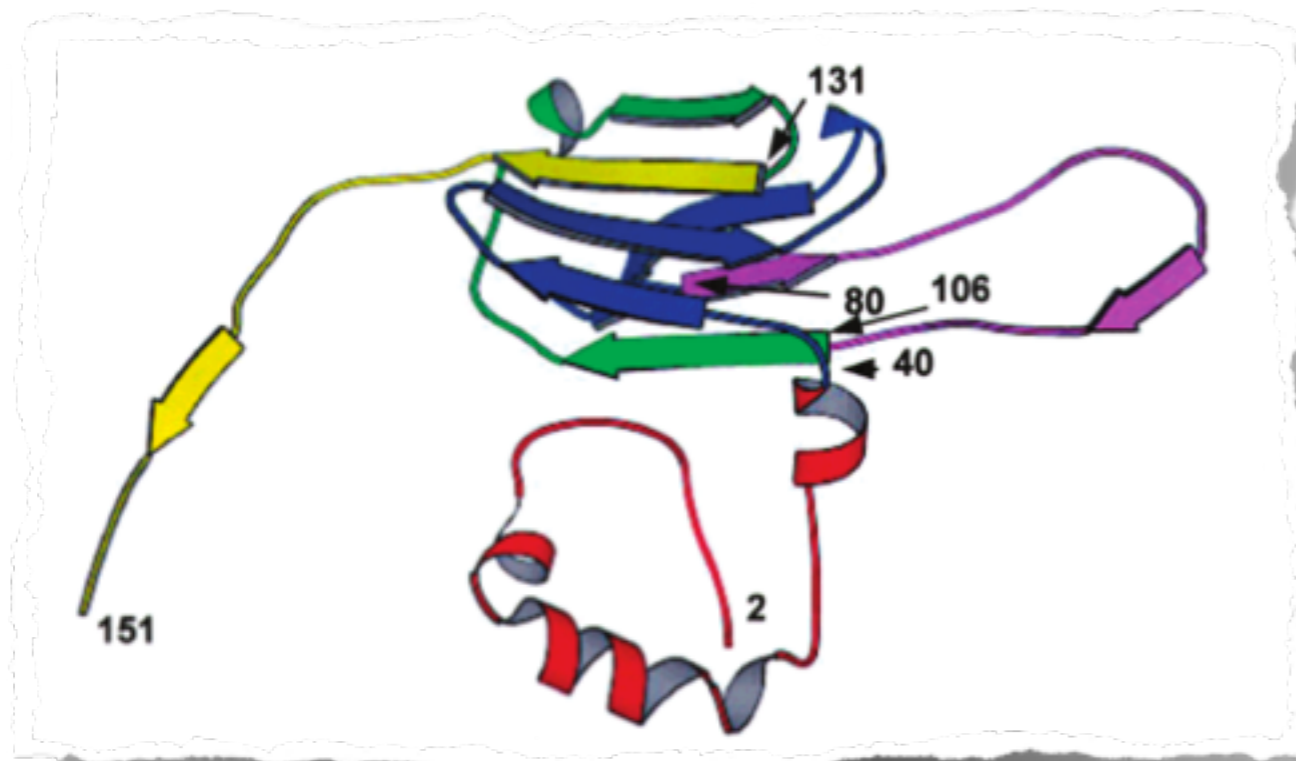
Hydrogen/deuterium exchange

- Monitor the rate at which protein hydrogens are replaced by deuteriums (or vice versa)
- Three types of hydrogen in proteins, only backbone amide hydrogens exchange at measurable rate
- Exchange can be (effectively) quenched by dropping pH to ~2.5, and temperature to 0°C
- Exchange rates reveal solvent accessibility

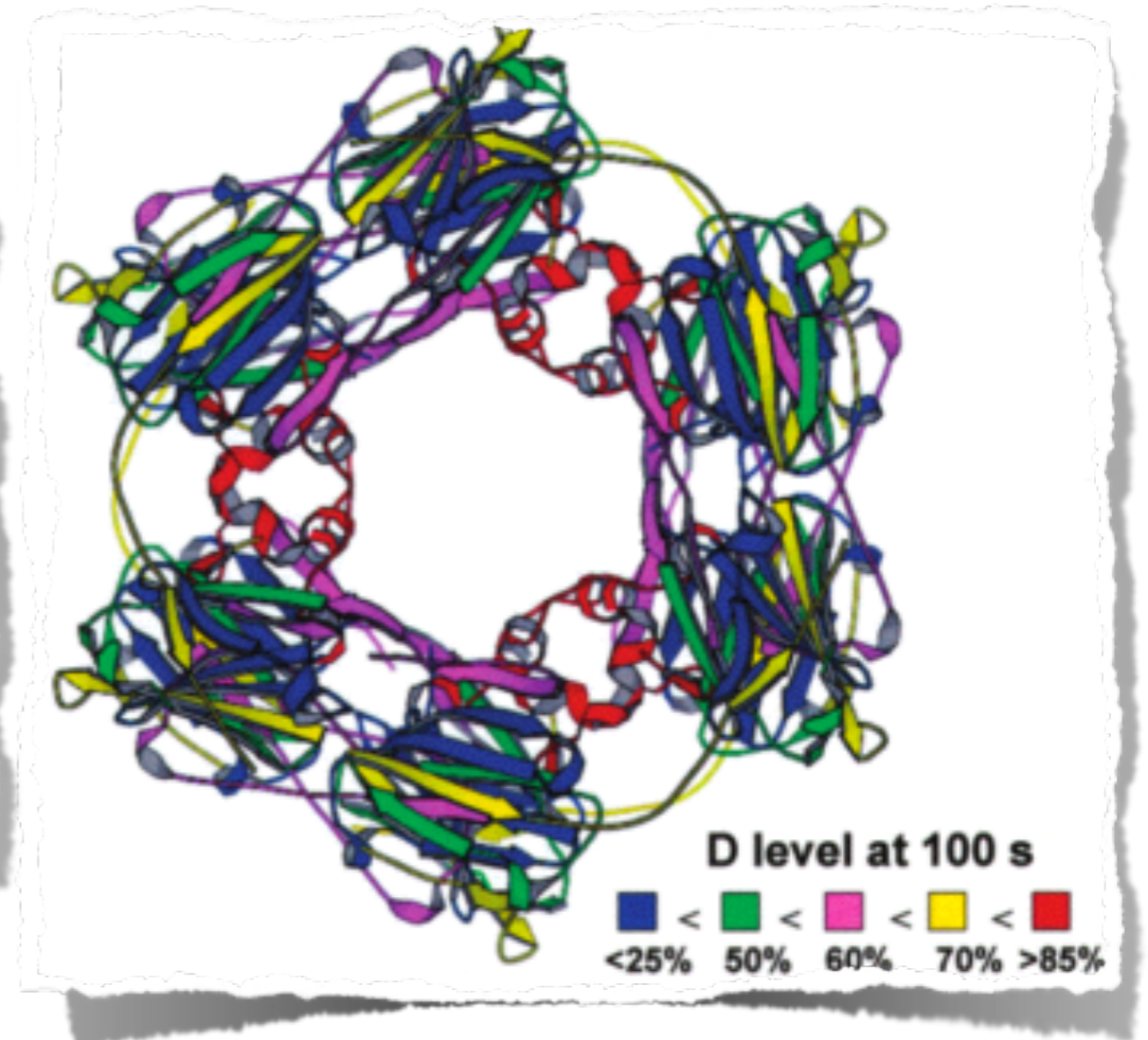


- Problems centre on localisation, and interpretation of solvent accessibility

Labelling technology - Example



Wintrode ... Smith, Biochemistry (2003) 42, 10667-73



- Hydrogen/deuterium exchange of oligomeric 'molecular chaperone' protein
- Side-chains at interfaces exchange relatively rapidly, suggesting a labile oligomer

'Soft' Ionisation

- From molecules in solution to ions in vacuum
- Established ionisation techniques resulted in covalent fragmentation of molecules
- Soft ionisation techniques allow ionisation of large ions without their fragmentation
- Current 'mass record' is >100 MDa!

Nobel Prize in Chemistry 2002

"for their development of soft desorption ionisation methods for mass spectrometric analyses of biological macromolecules"



John B. Fenn

🕒 1/4 of the prize

USA

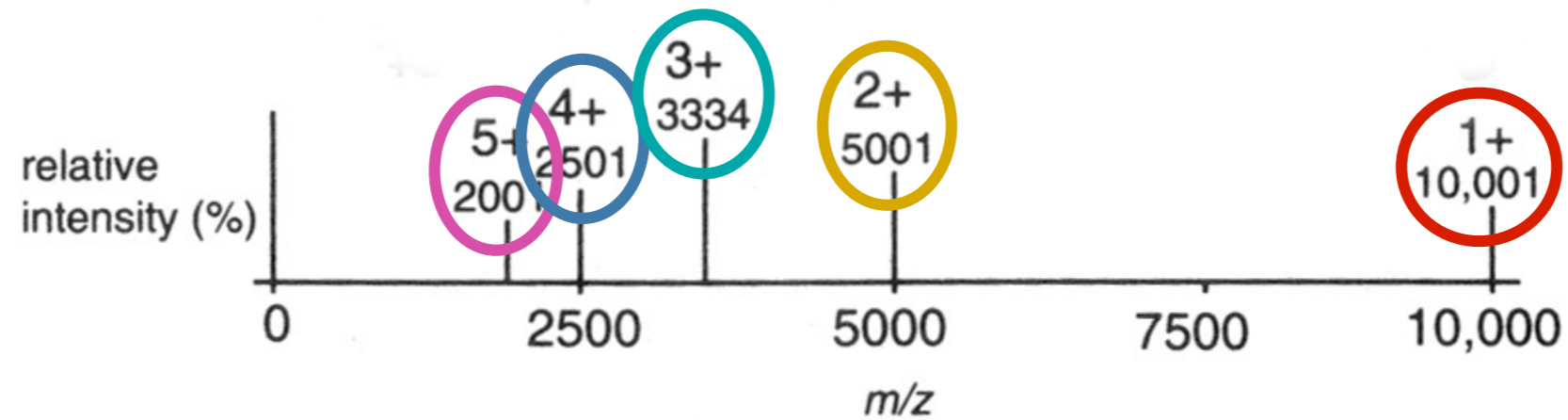


Koichi Tanaka

🕒 1/4 of the prize

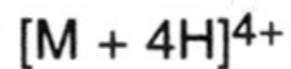
Japan

ESI mass spectrum of single protein



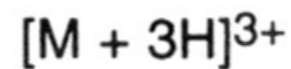
$$m/z = 10,005 \div 5$$

$$m/z = 2001$$



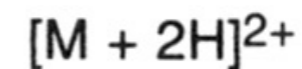
$$m/z = 10,004 \div 4$$

$$m/z = 2501$$



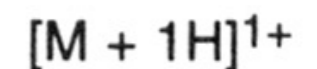
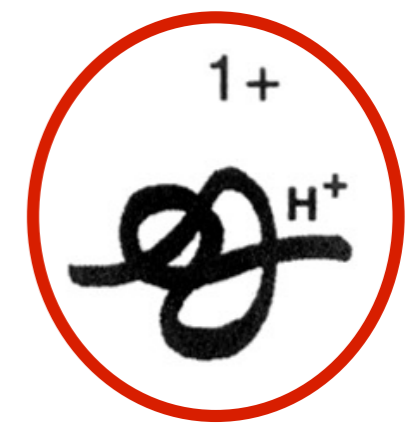
$$m/z = 10,003 \div 3$$

$$m/z = 3334$$



$$m/z = 10,002 \div 2$$

$$m/z = 5,001$$

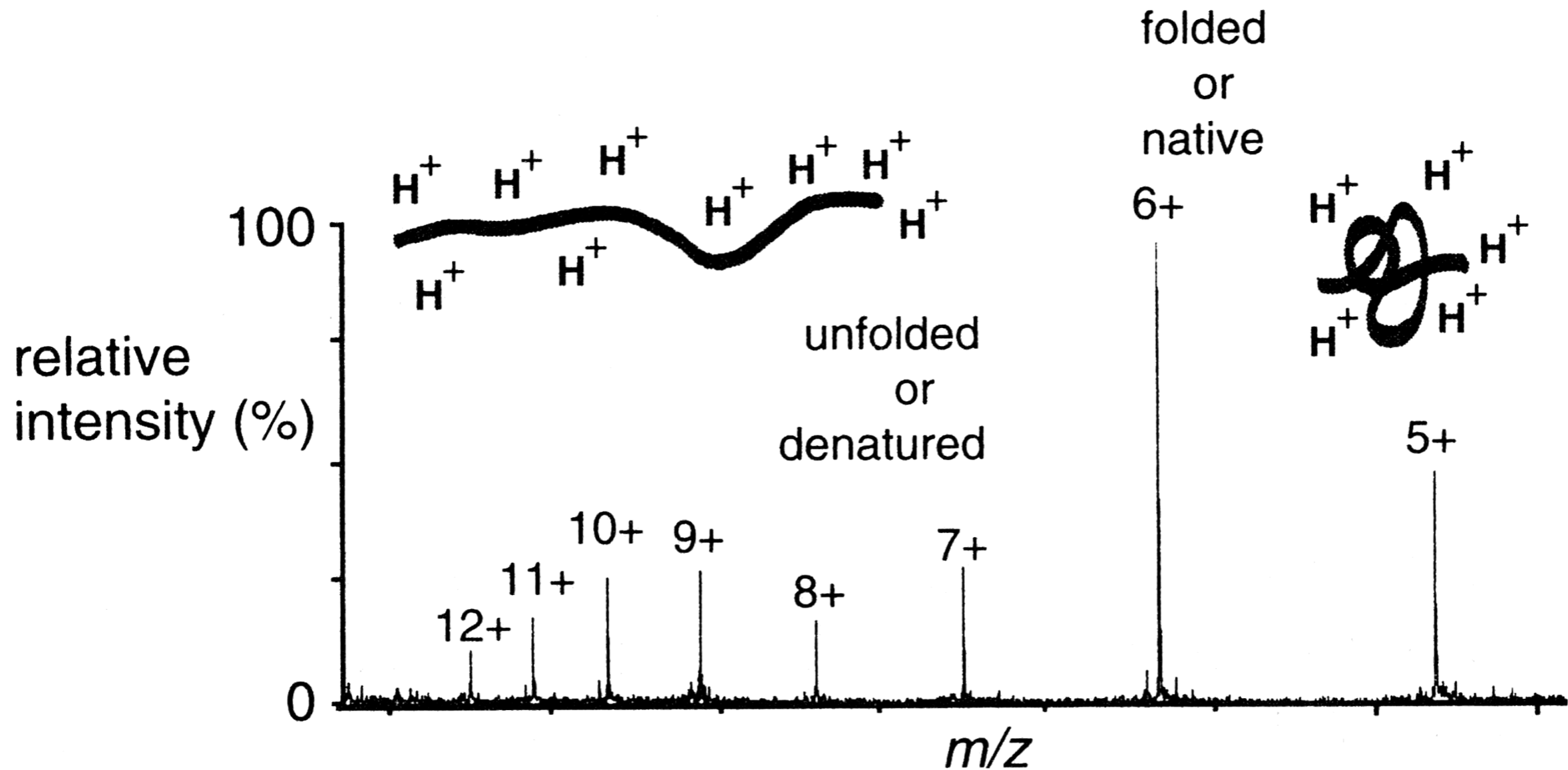


$$m/z = 10,001 \div 1$$

$$m/z = 10,001$$

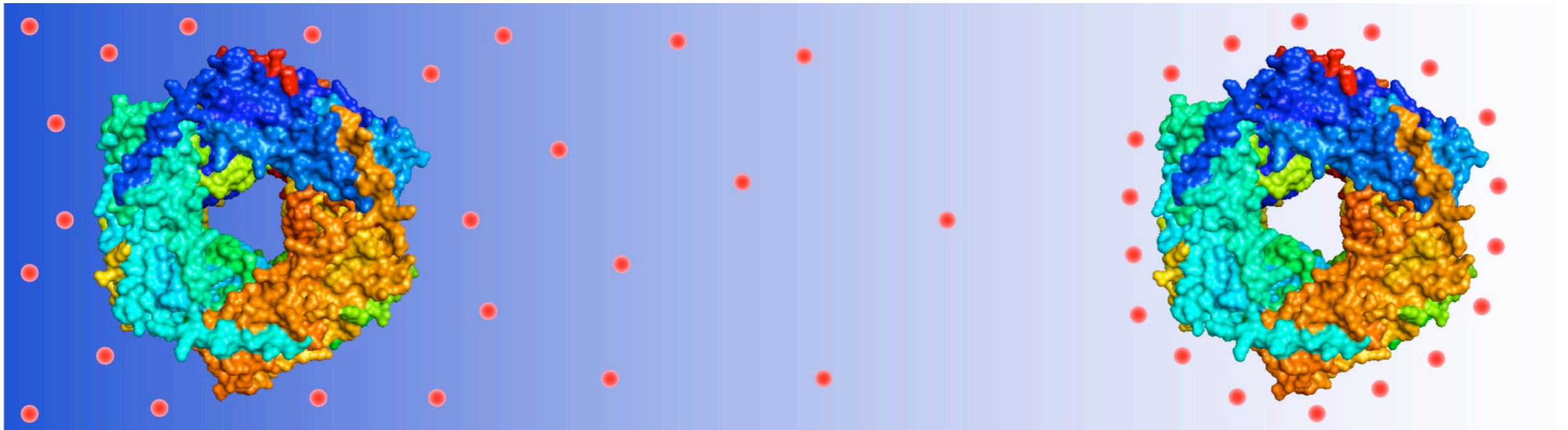
- Electrospray mass spectra show multiple 'charge states' for a 10 kDa protein

Conformational effects on ESI spectra



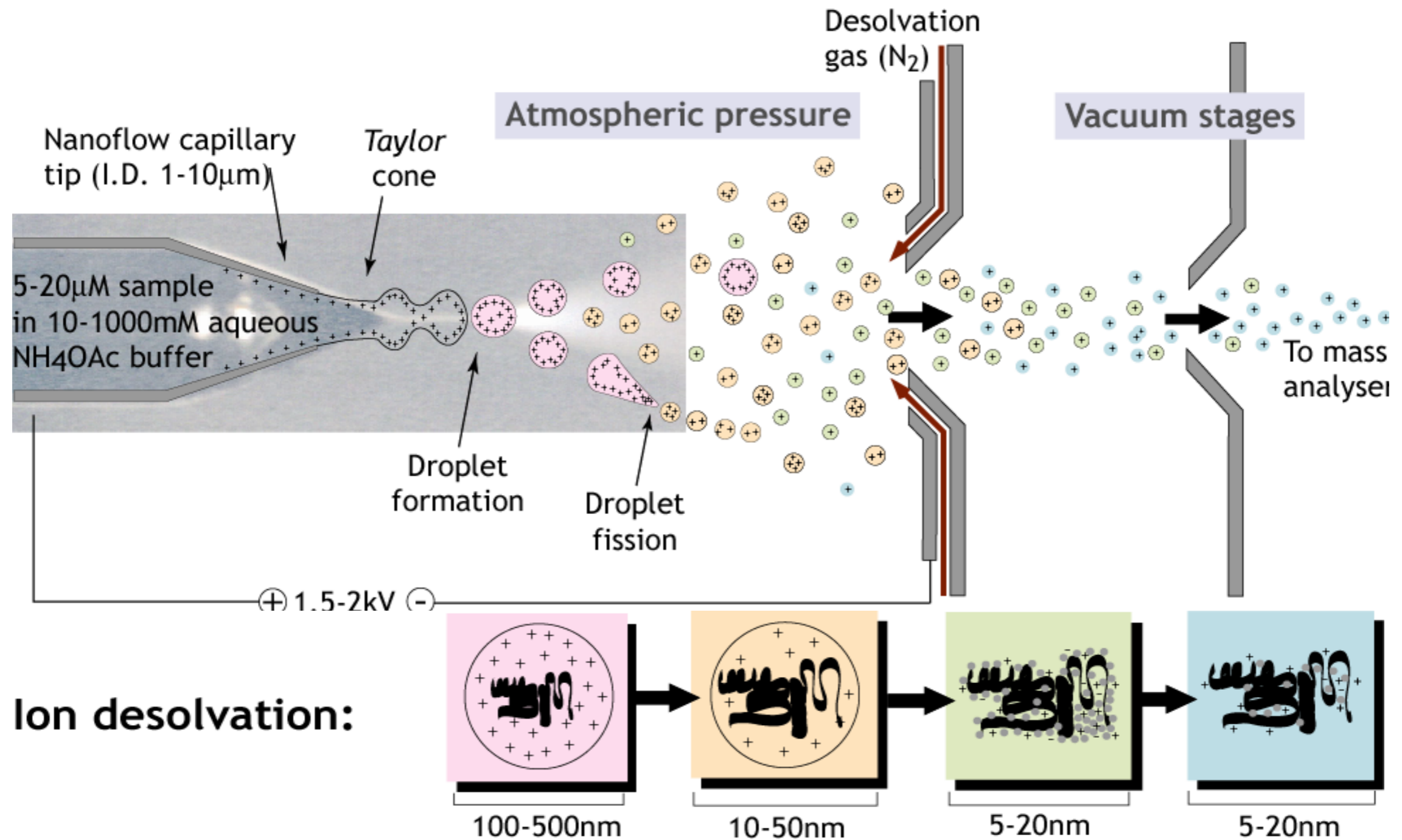
- Folded state of protein governs its surface area, and number of sites available for protonation

Maintaining noncovalent interactions



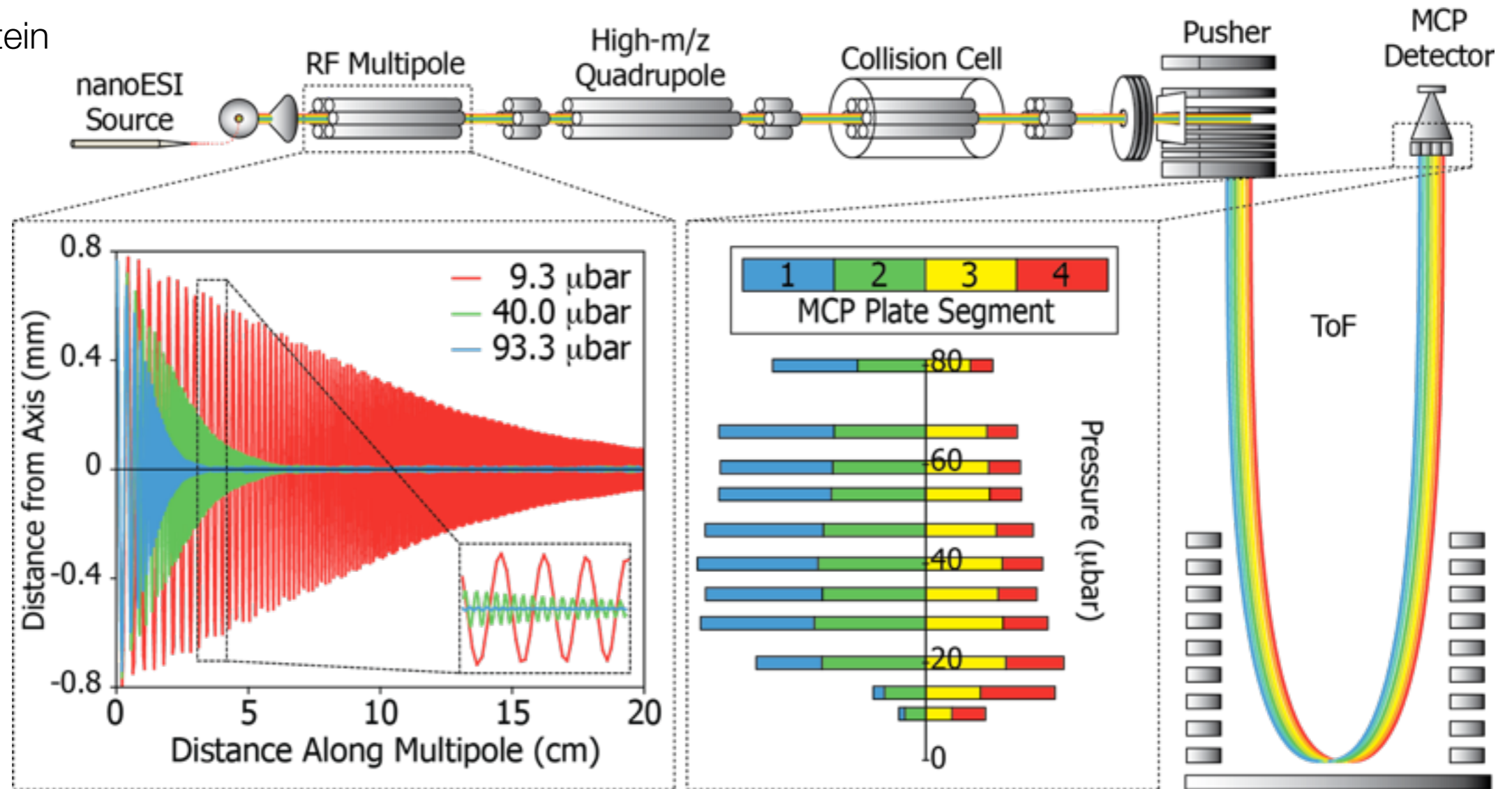
- Transfer multi-subunit protein assembly from solution into gas phase
- Requires control of ionisation conditions, and ion transmission

Nano-electrospray ionisation (nESI)



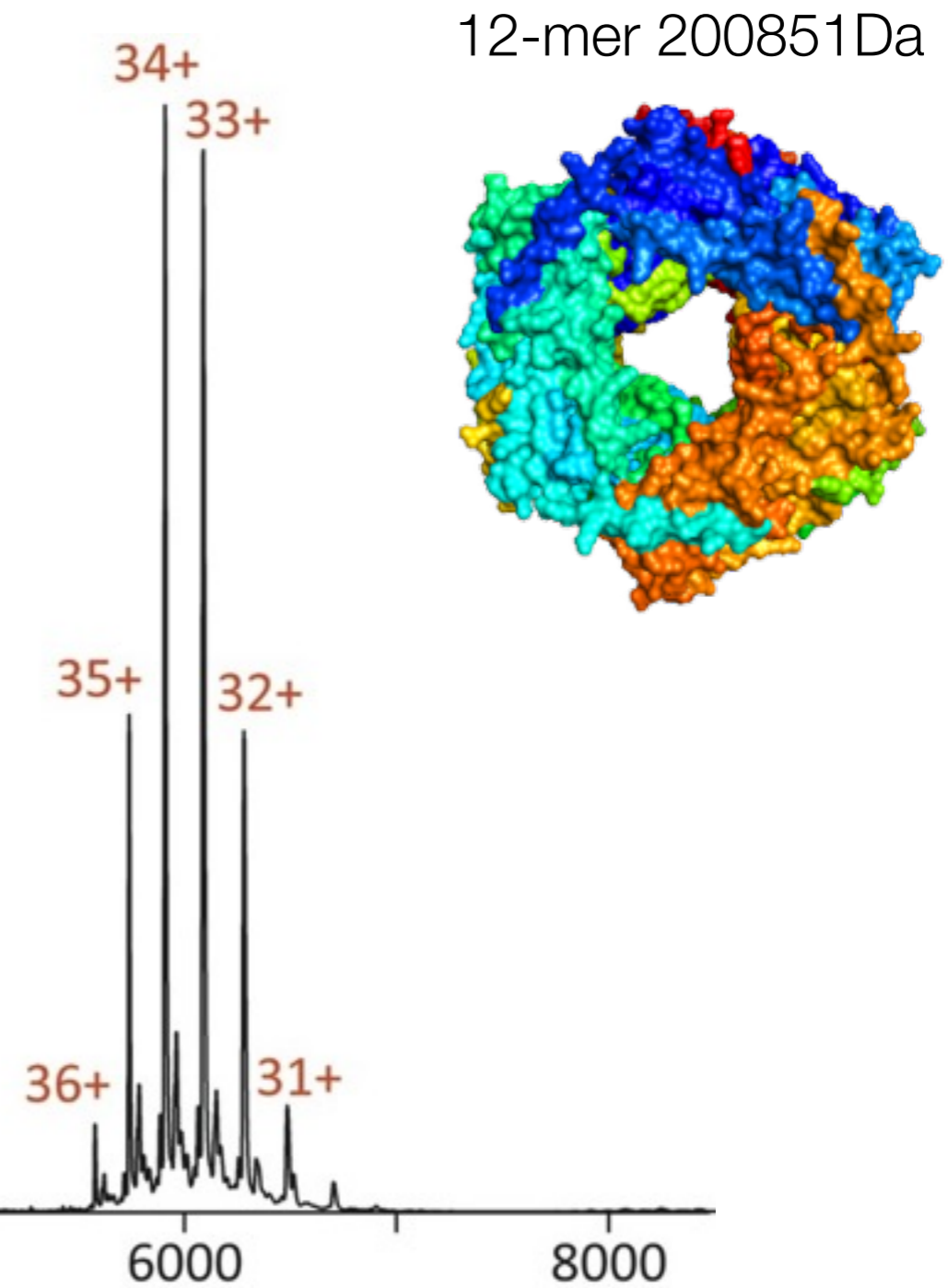
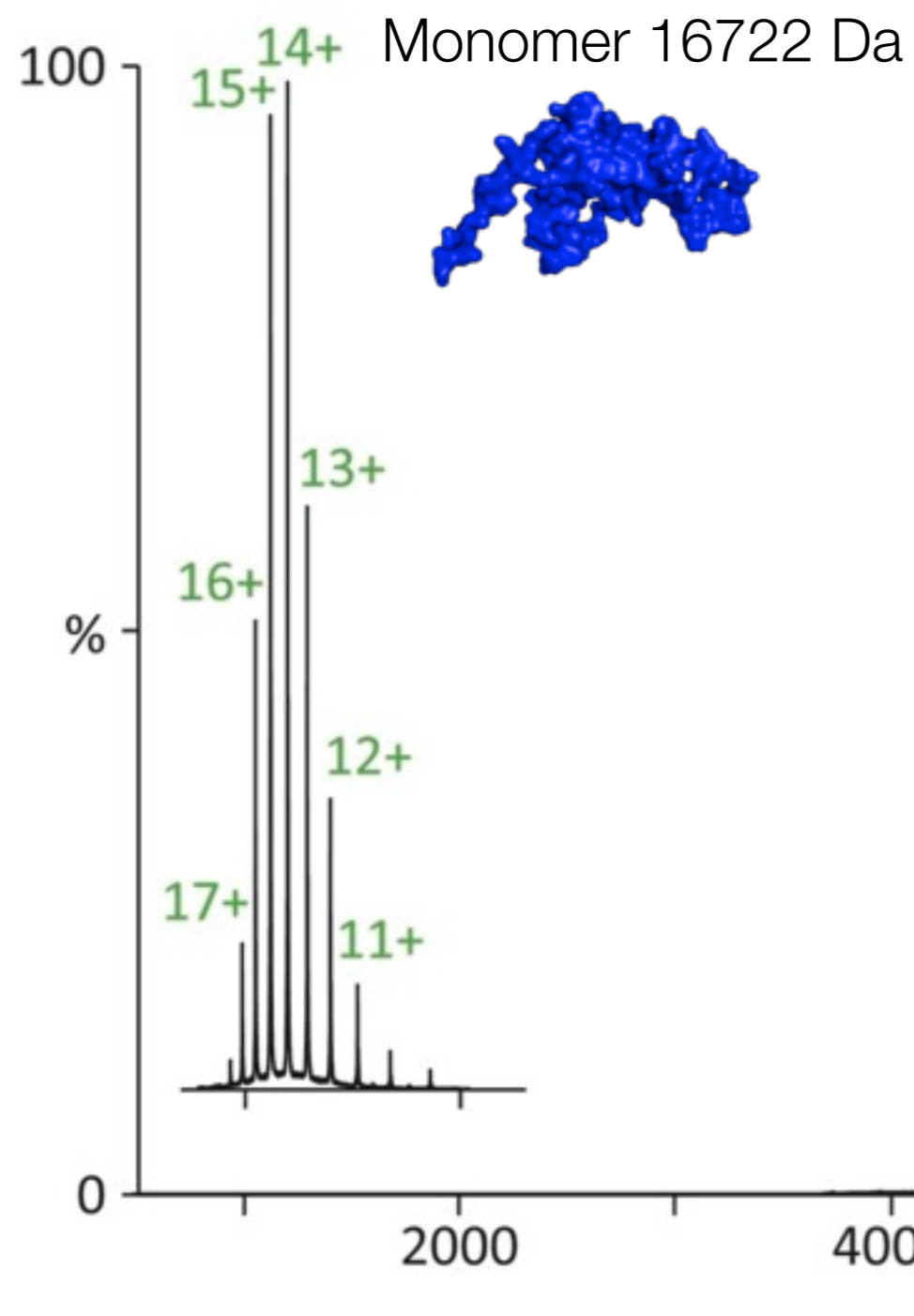
Collisional focussing

147 kDa Protein
Assembly

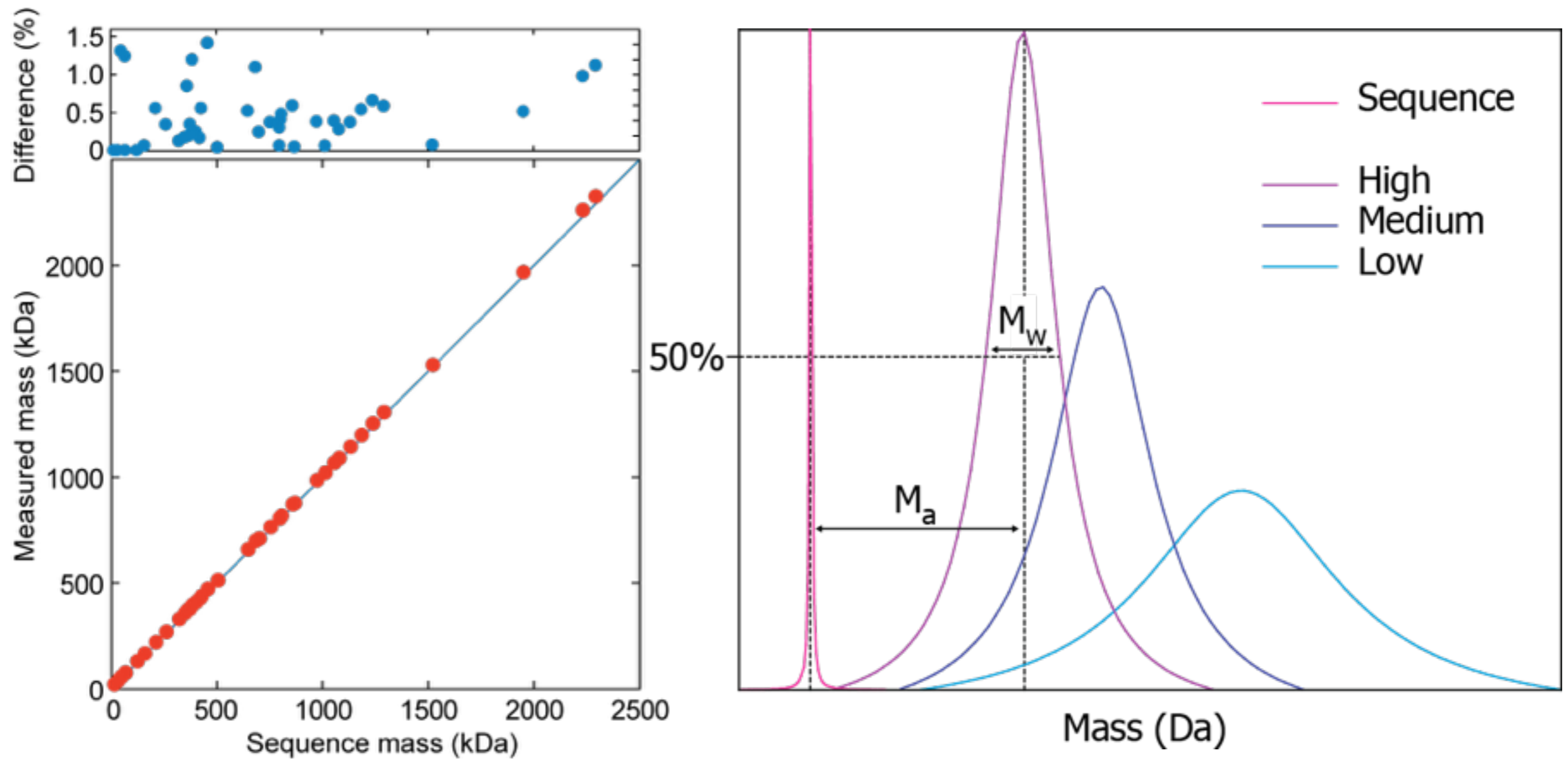


- Both axial and radial components of the ions' velocity can be dampened by collisions with background gas

nESI mass spectrum

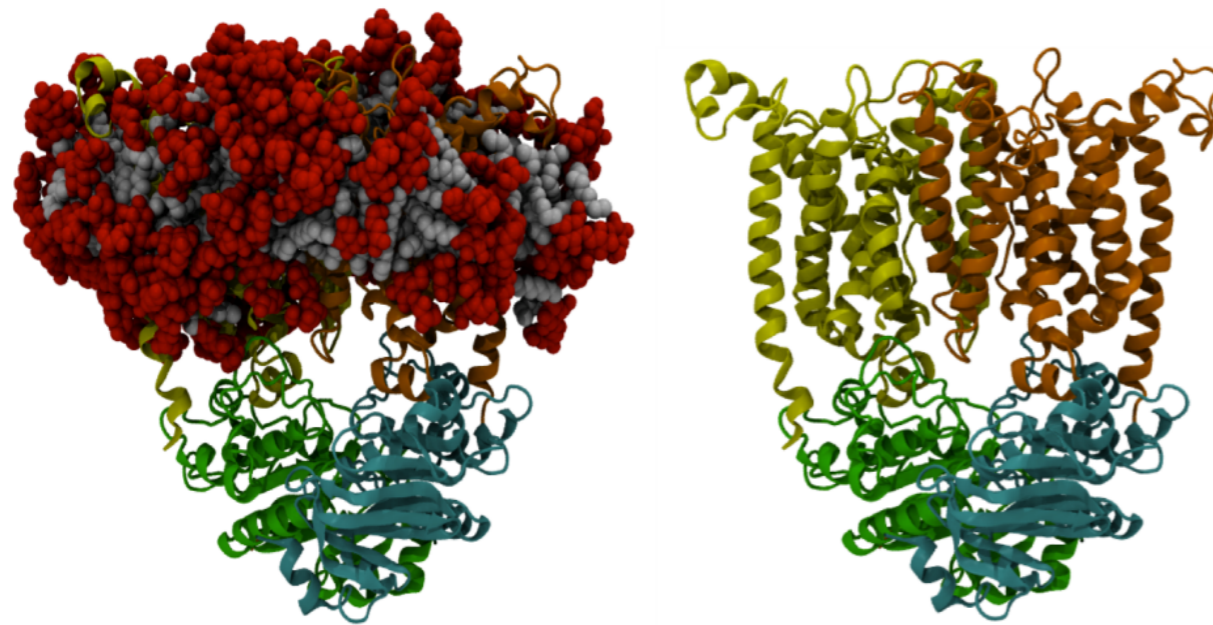


Mass accuracy

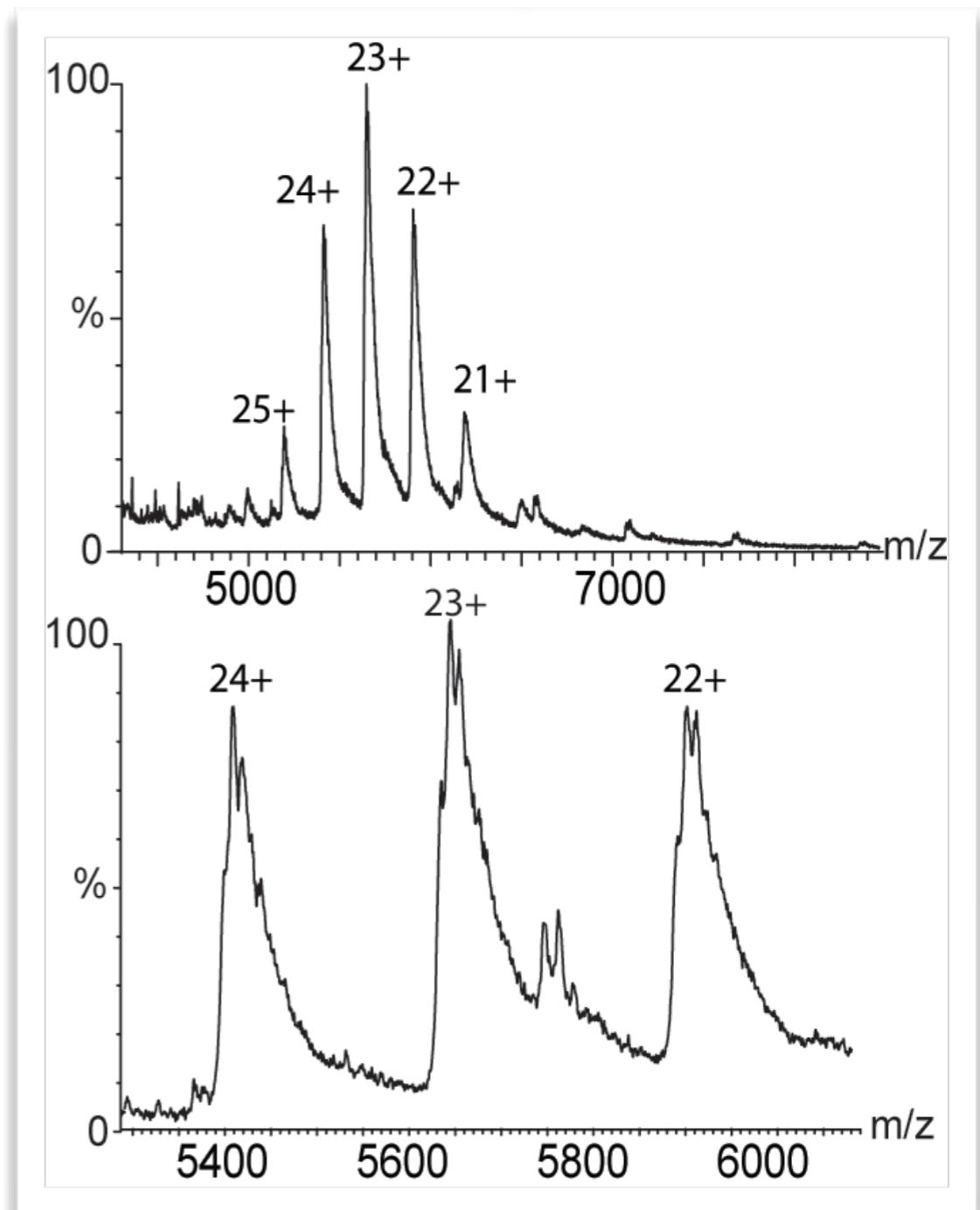


- Additional mass due to adducted solvent molecules and buffer ions
- Number of adducts inversely related to activation

Membrane protein assemblies are also tractable

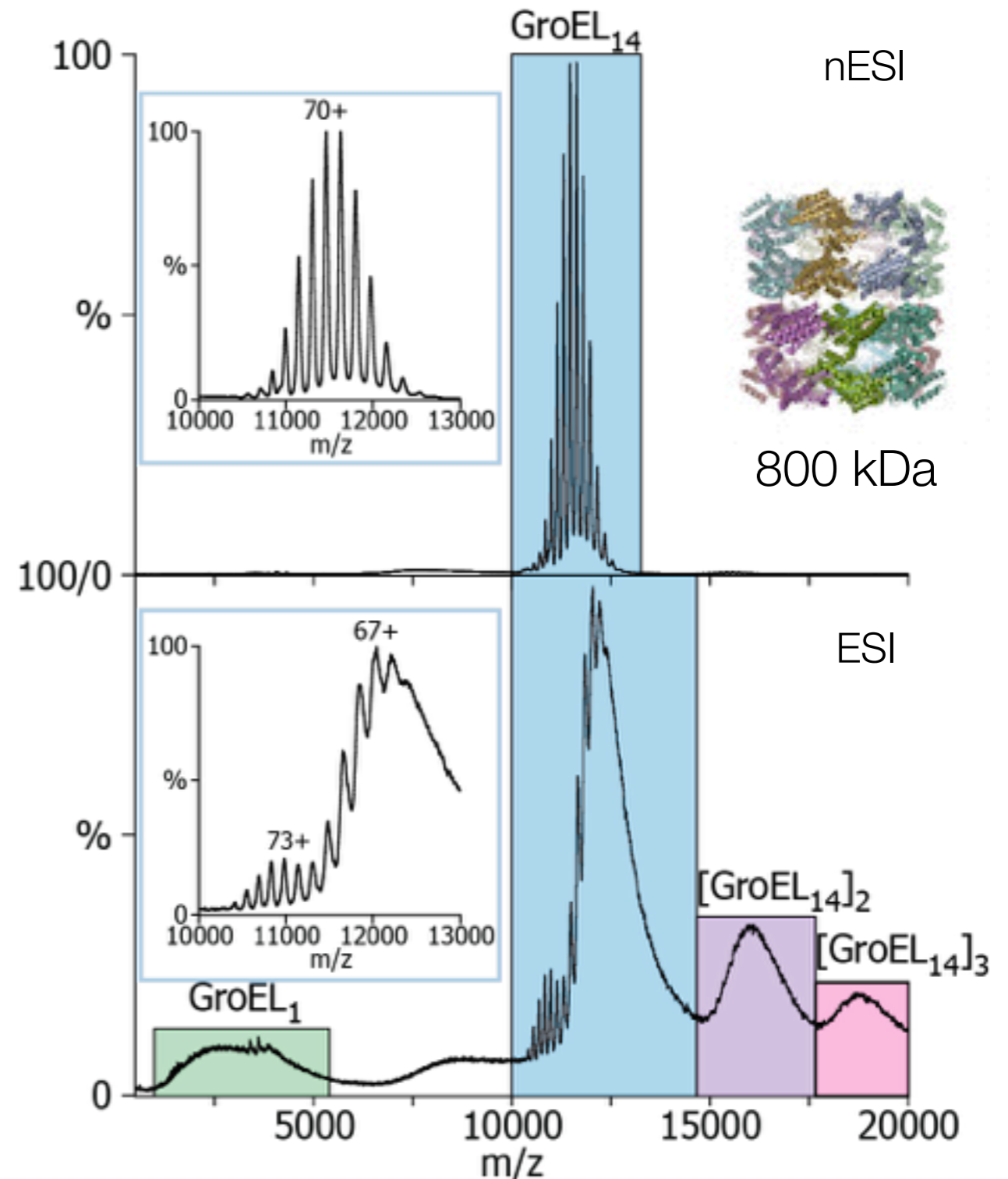


- Sample transferred into vacuum within detergent micelle
- Activation within the mass spectrometer removes detergent to leave “naked” protein assembly

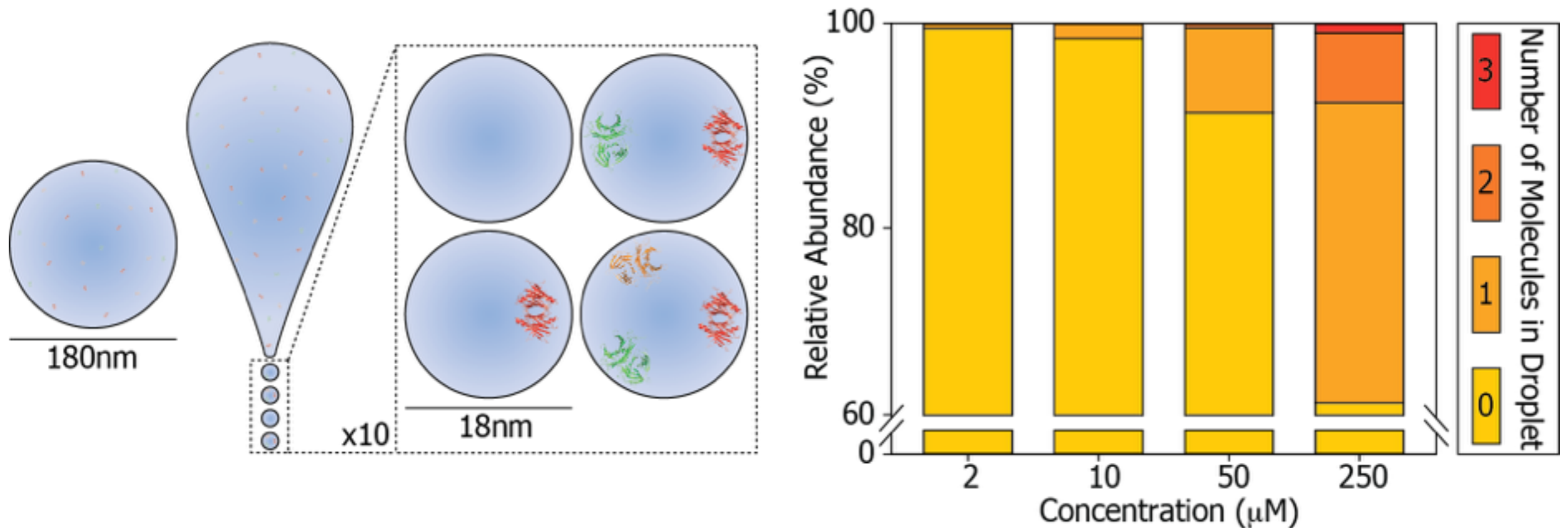


Benefits of nESI

- Lower sample amounts (flow rate approx 10nL/min, vs 5 μ L/min in ESI)
- Can use aqueous buffers and ambient temperatures
- Narrower charge states due to fewer adduction
- Less dissociation of oligomer
- Symmetrical charge state distribution indicative of a single conformation
- Fewer non-specific aggregates

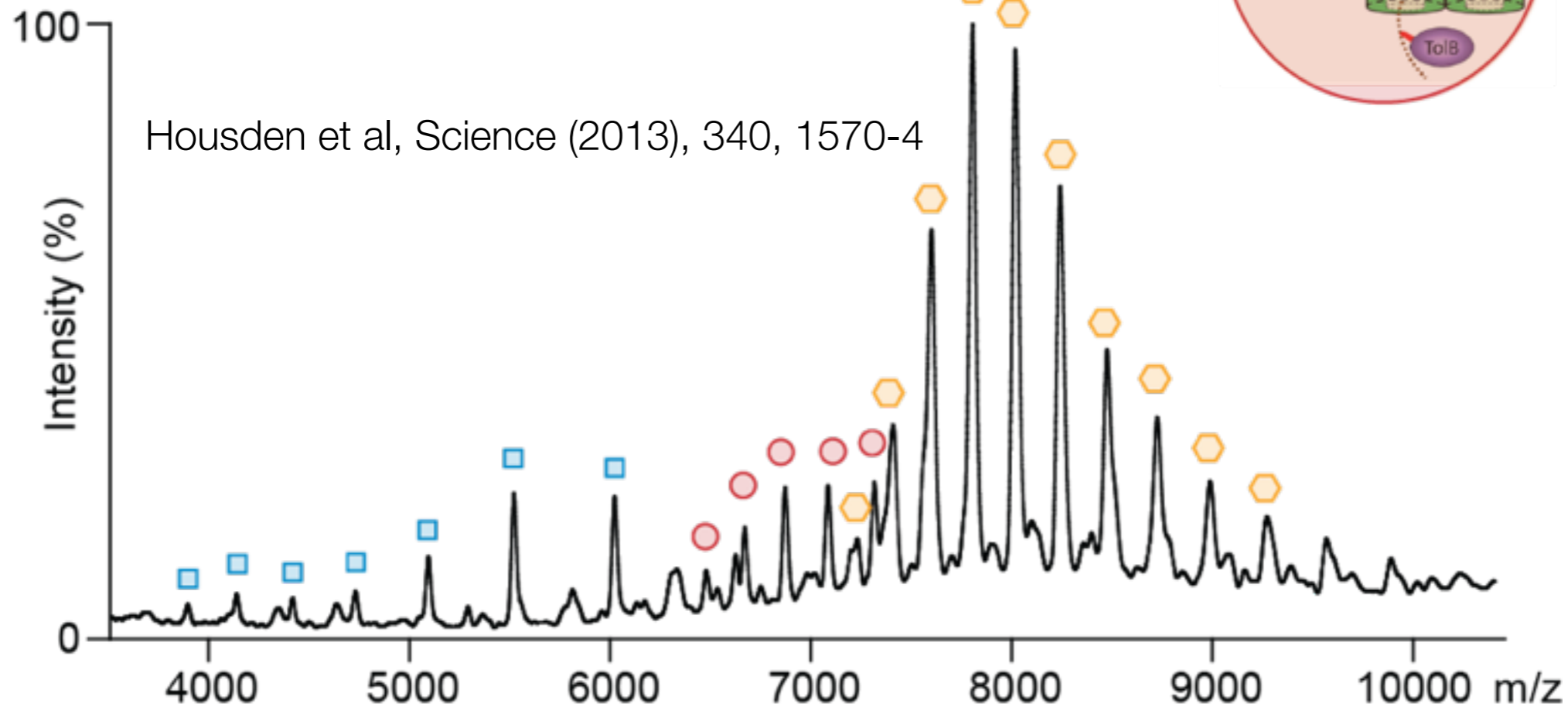
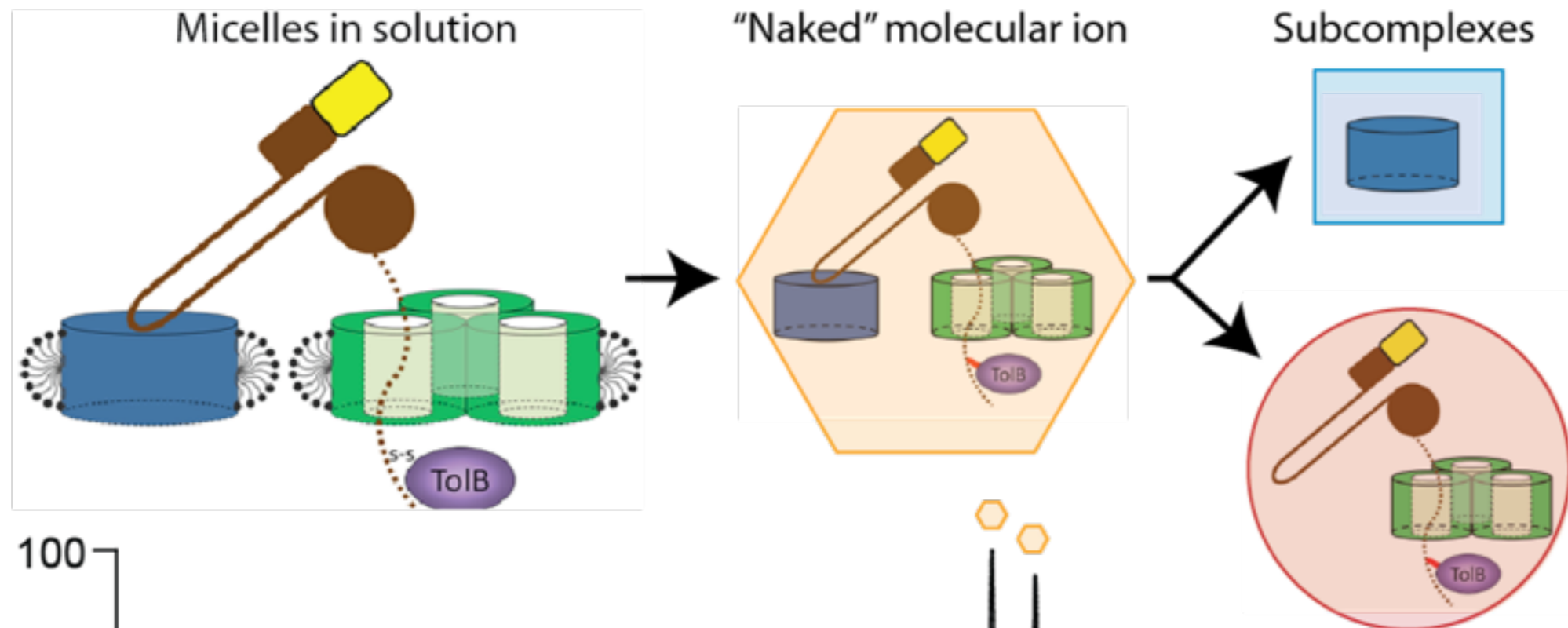


Non-specific associations during ESI

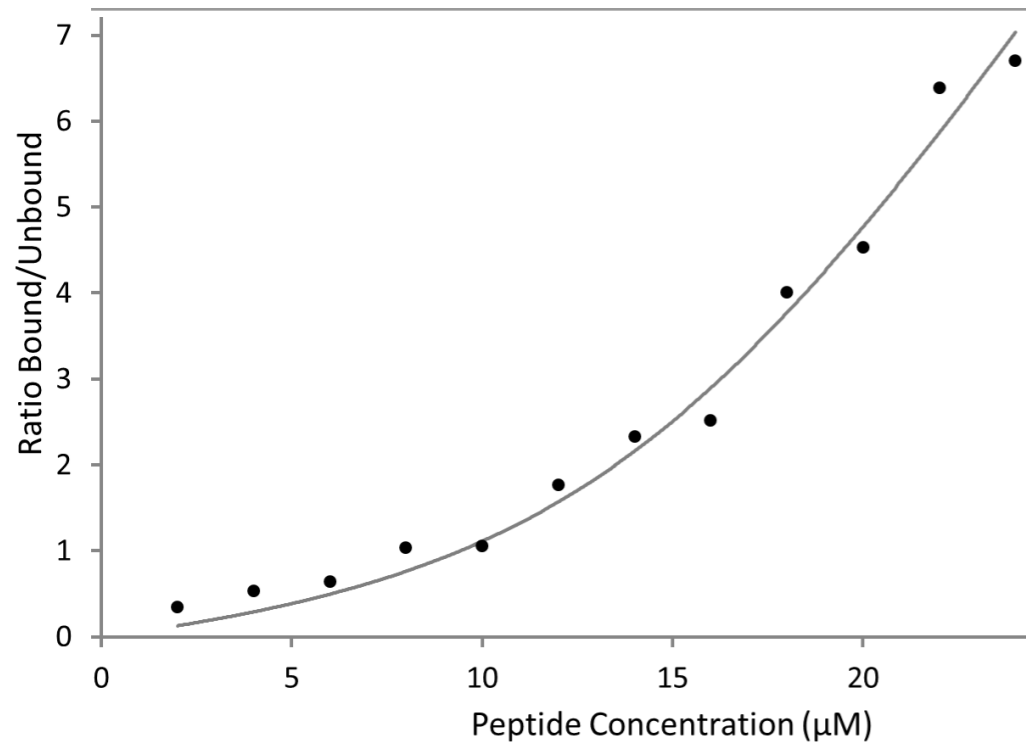


- Probability of there being >1 analyte molecules in 'final' ESI droplet
- Most droplets are empty, occupancy increases with concentration
- Decreased initial droplet size in nESI reduces prevalence of non-specific aggregates

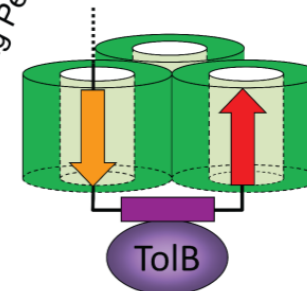
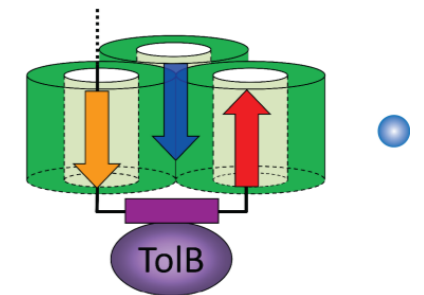
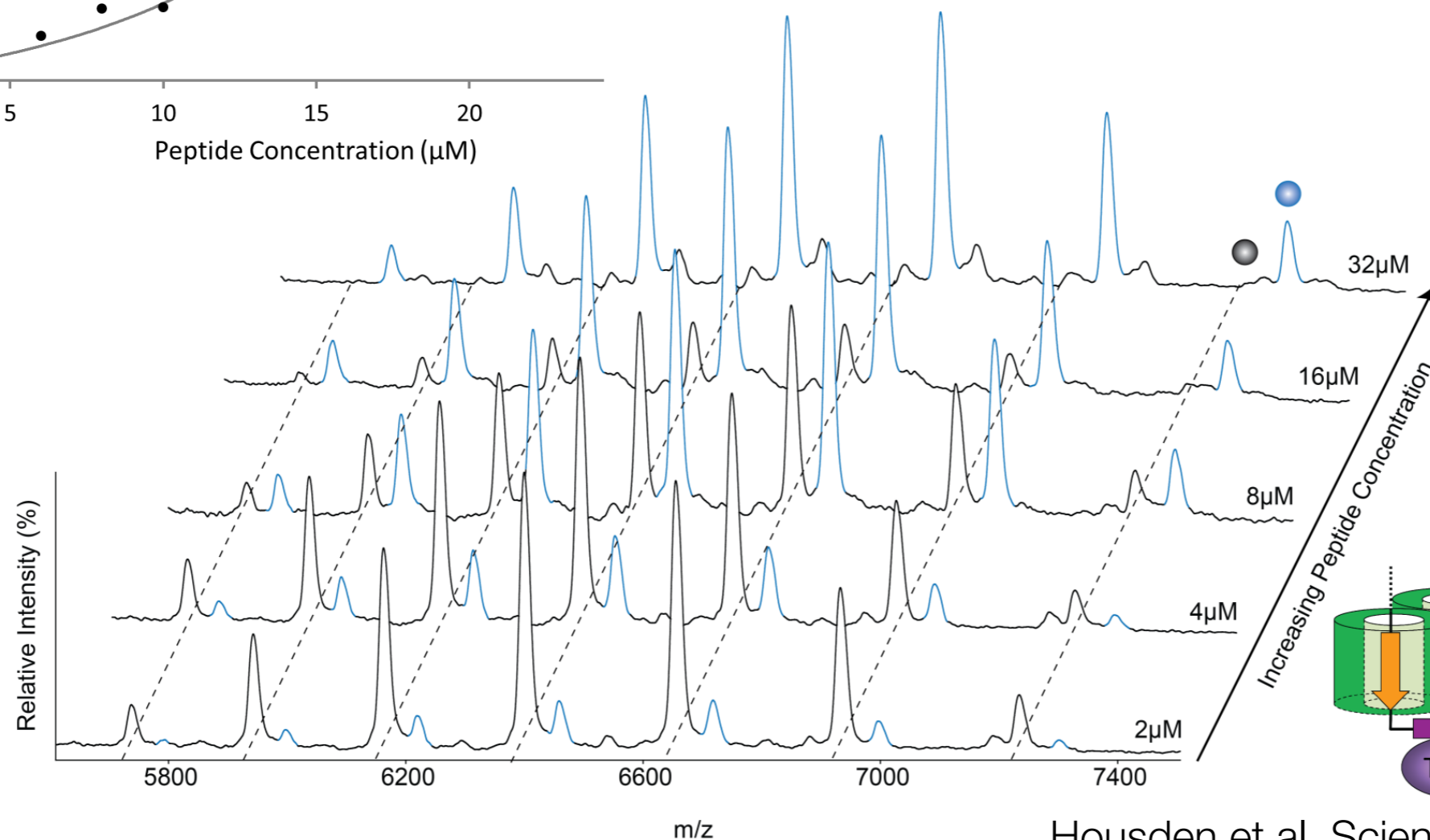
Intact membrane protein machines - Example



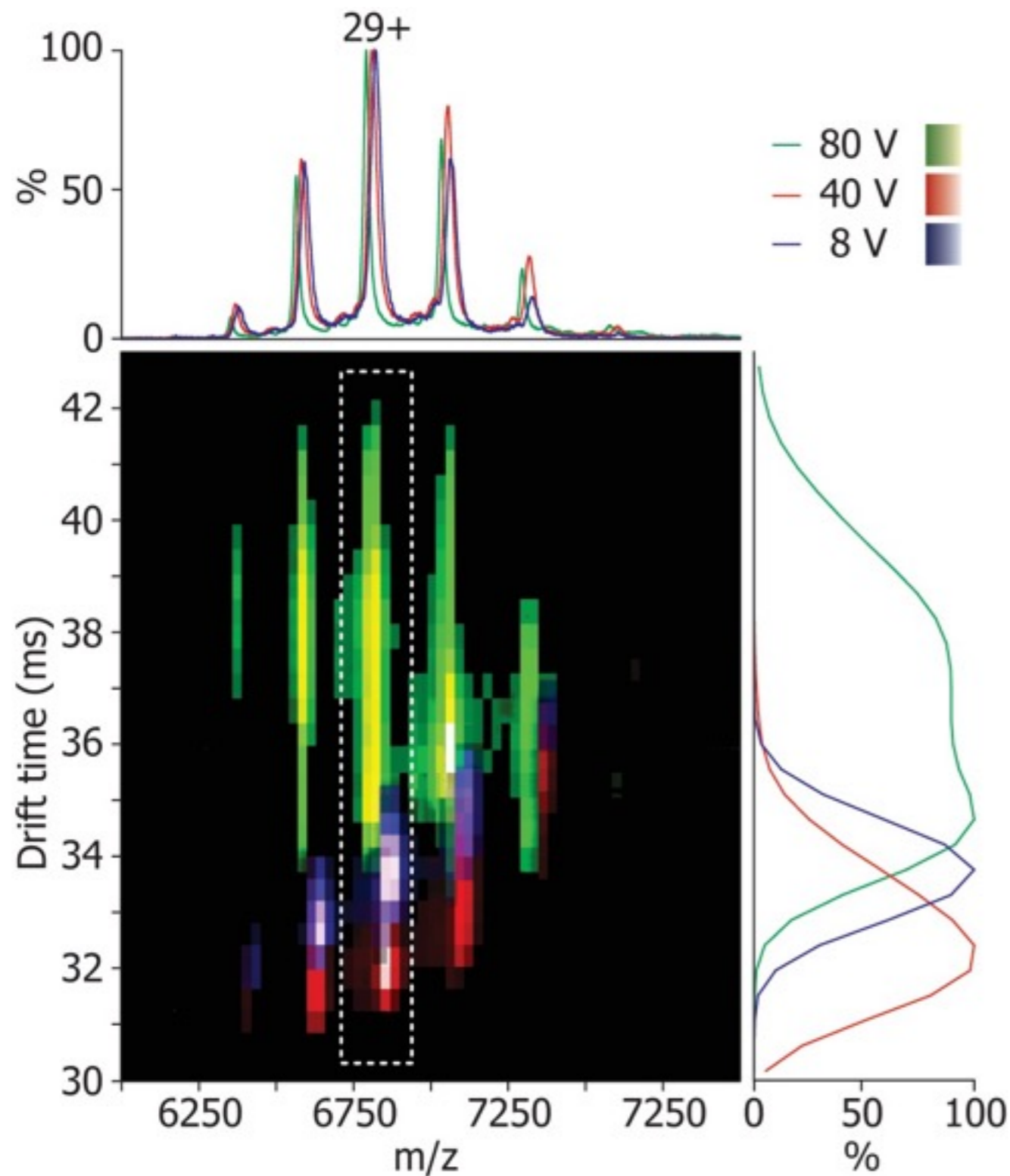
Determining binding affinities - Example



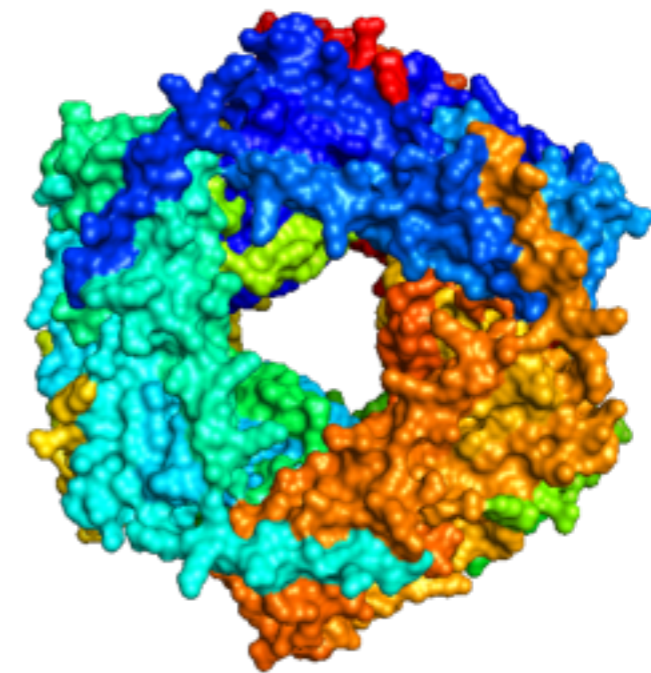
- Titration experiments allow the determination of binding affinity
- Practicable in the 100 nM - 1 mM range



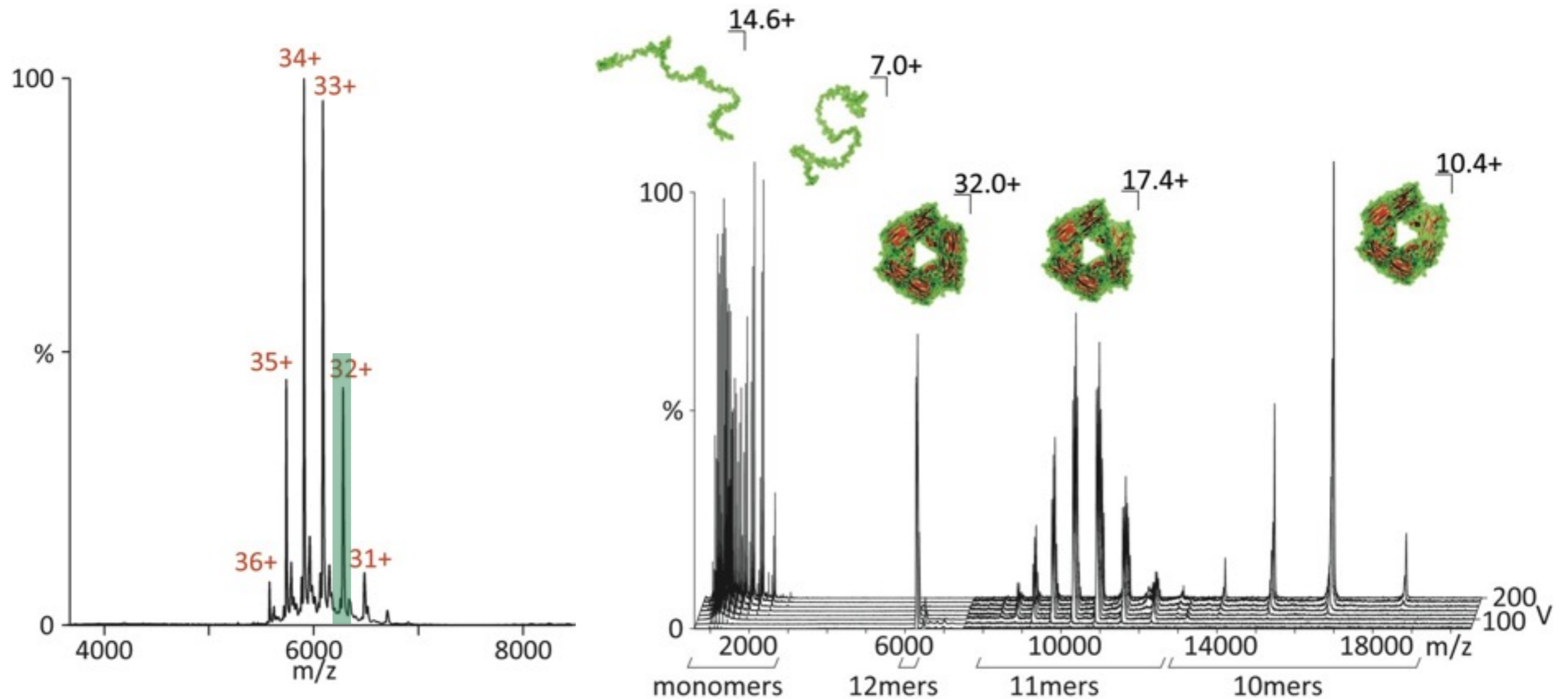
Effects of activation in the gas phase



- Activating conditions lead to high quality mass spectra
- Same activation can lead to
 - collapse (i.e. smaller CCS)
 - then unfolding (i.e. larger CCS)

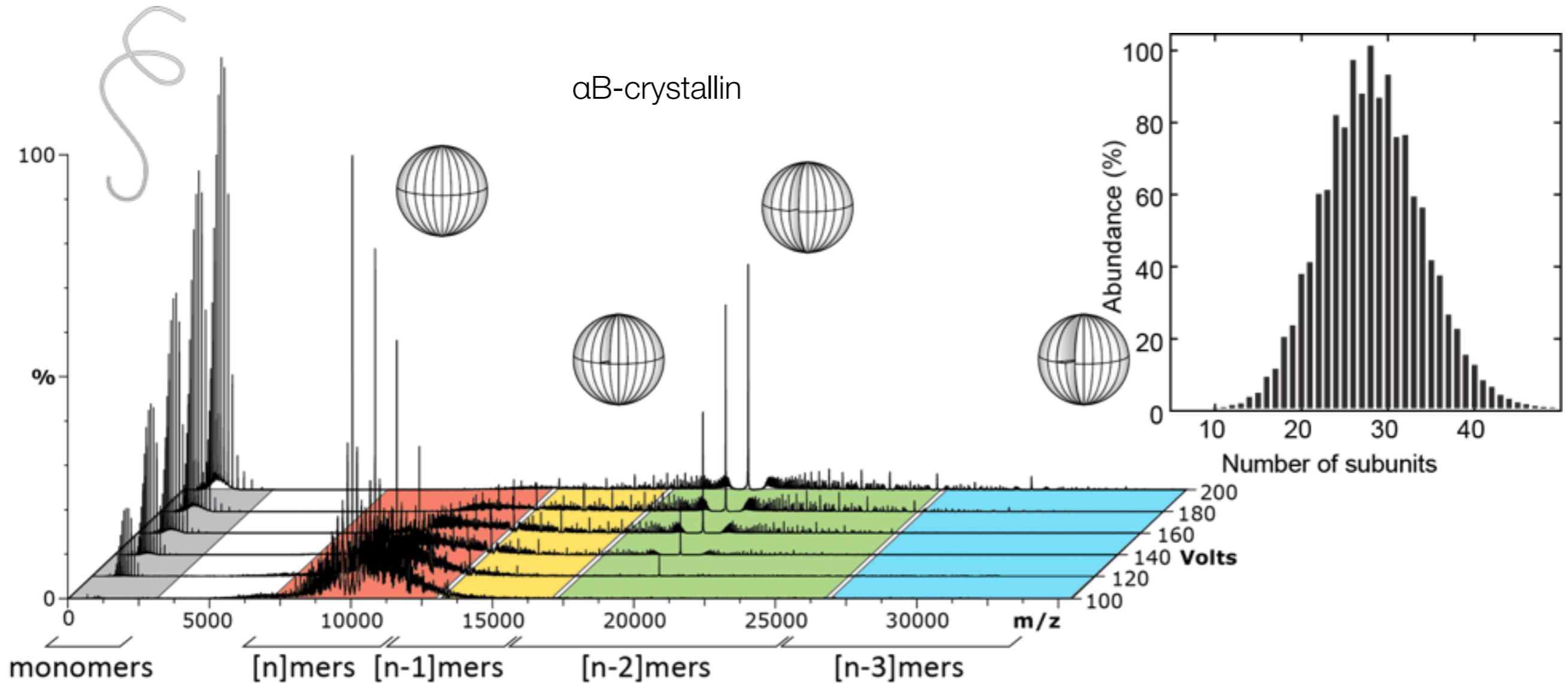


Collision induced dissociation of protein assemblies



- Dissociation is asymmetric with respect to mass
- Unfolded, highly charged monomers are removed sequentially

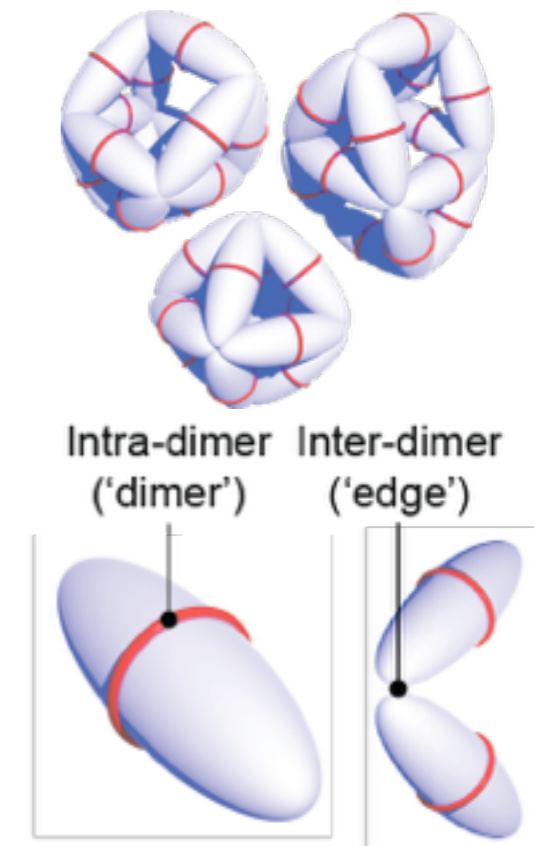
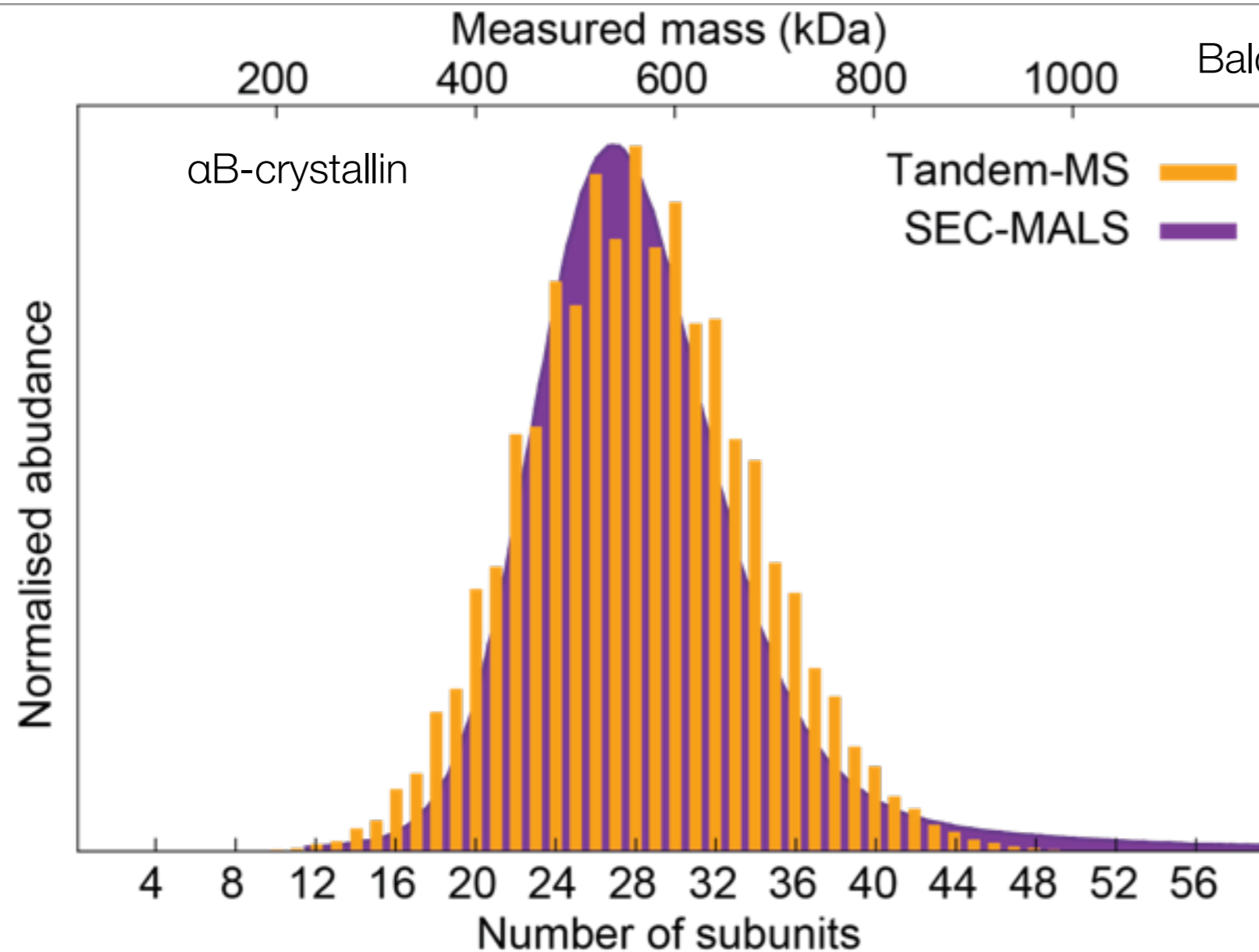
Deconvoluting heterogeneity with CID



- Peak separation is aided by the charge reduction afforded by CID
- Predictable nature of CID allows back calculation of oligomeric distribution

Quantifying stoichiometries

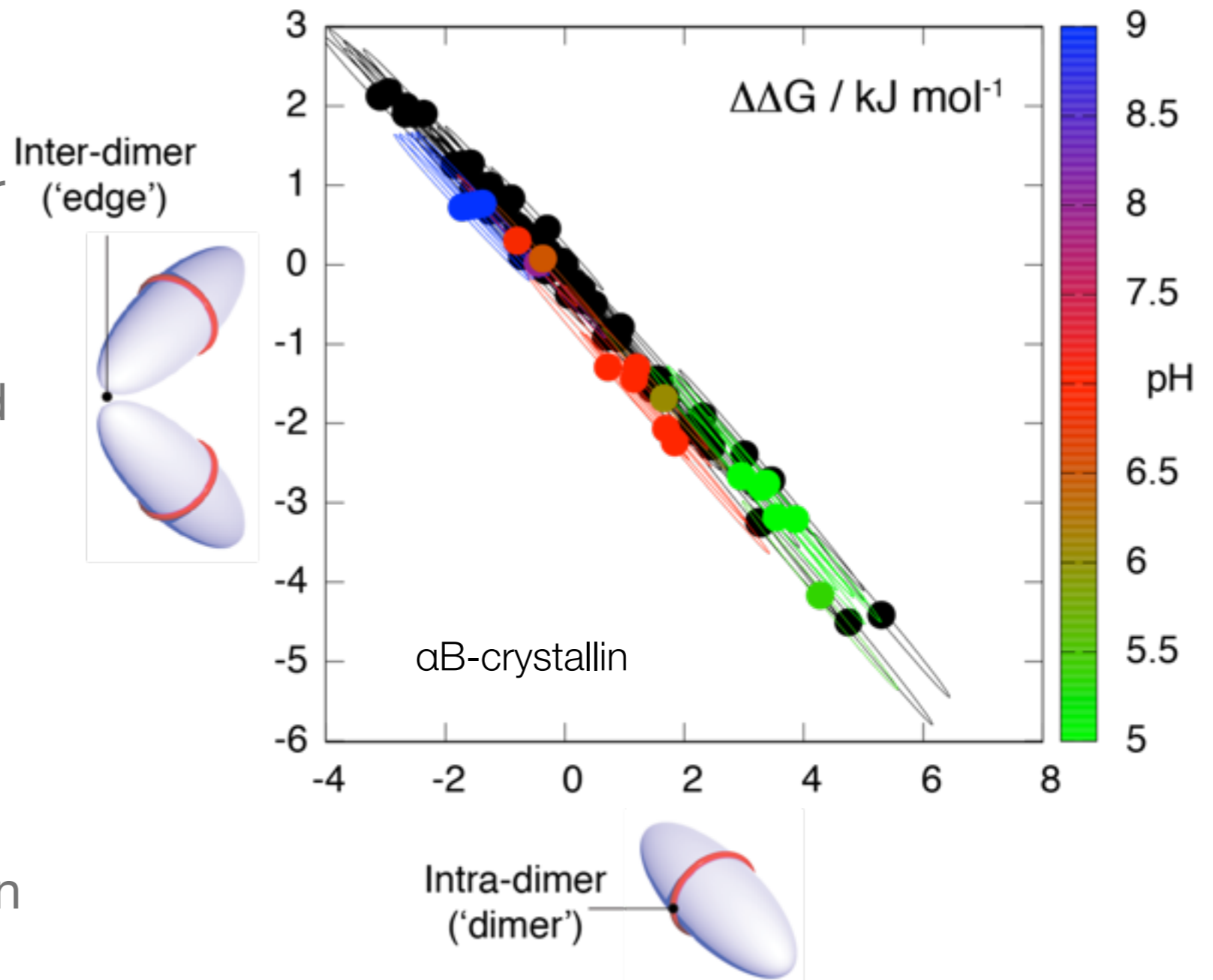
Baldwin et al, J Mol Biol (2011), 297-309



- MS versus size-exclusion chromatography with multi-angle light scattering
- For proteins of similar composition, abundances match solution values

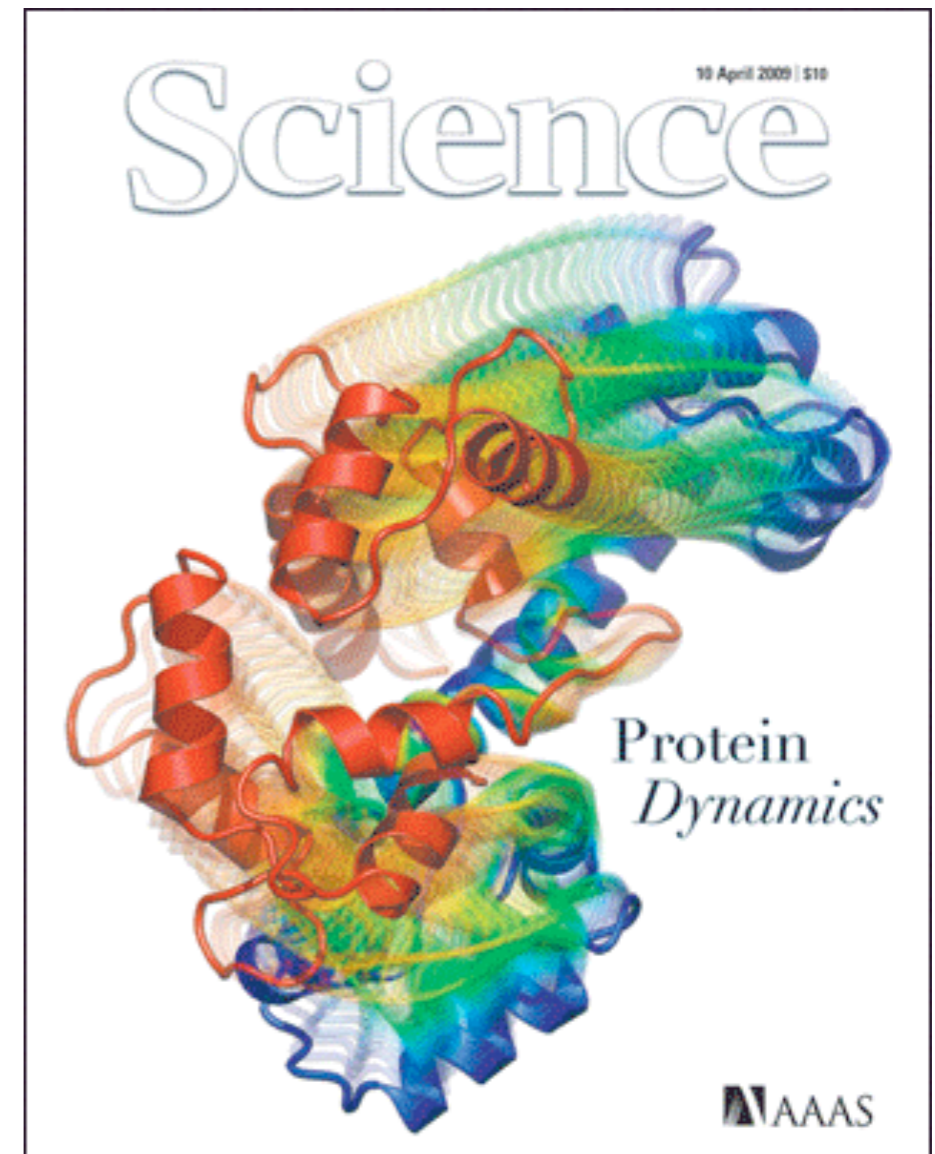
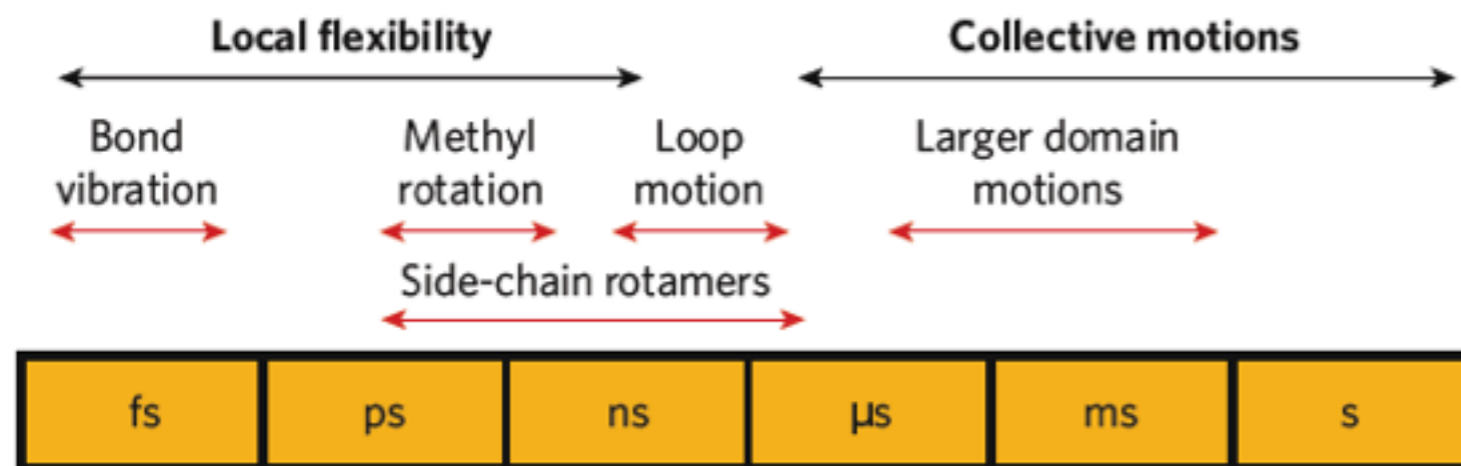
Free energies from MS measurements - Example

- α B-crystallin forms polydisperse oligomers via inter- and intra-dimer interfaces
- Abundances of stoichiometries are fitted to an appropriate biophysical model for oligomerisation
- Effect of mutation/ environment on free energies extracted
- Allosteric communication between interfaces



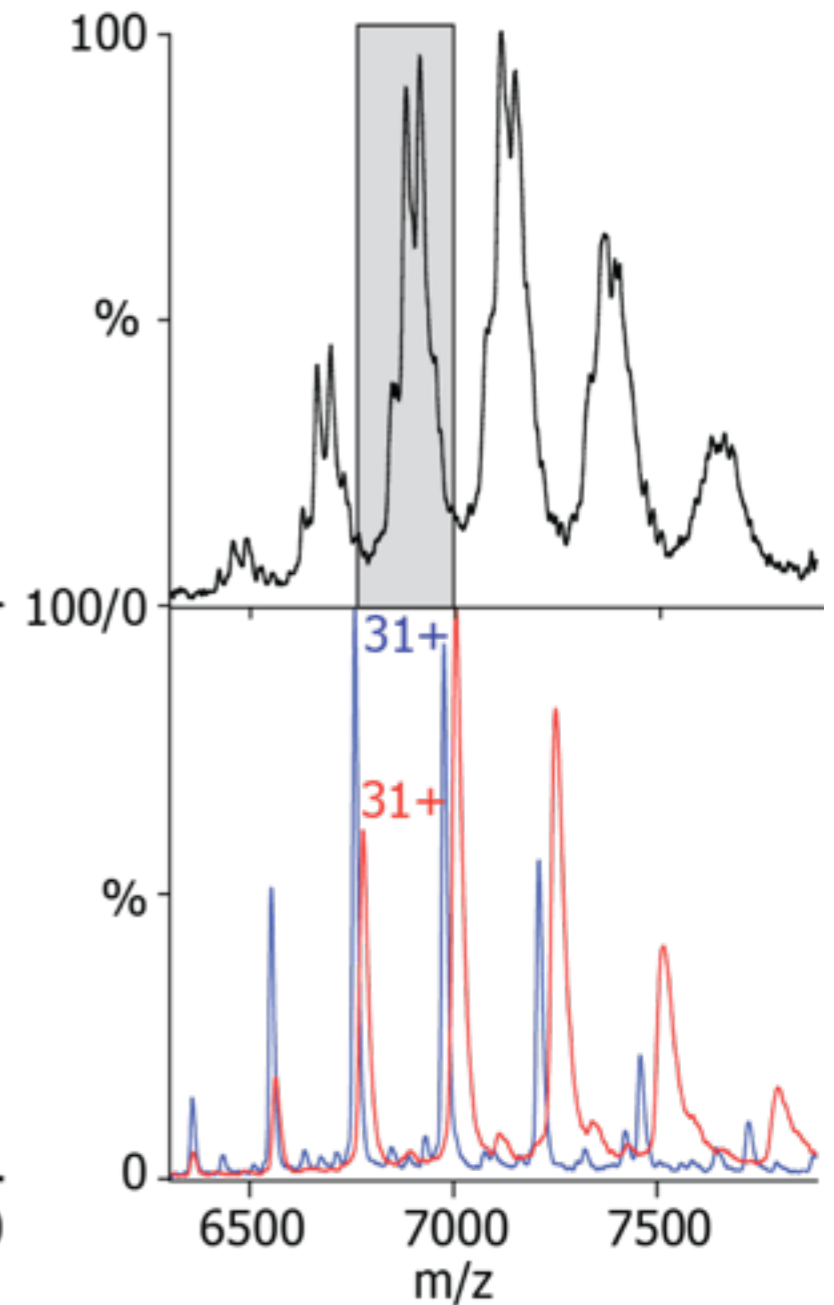
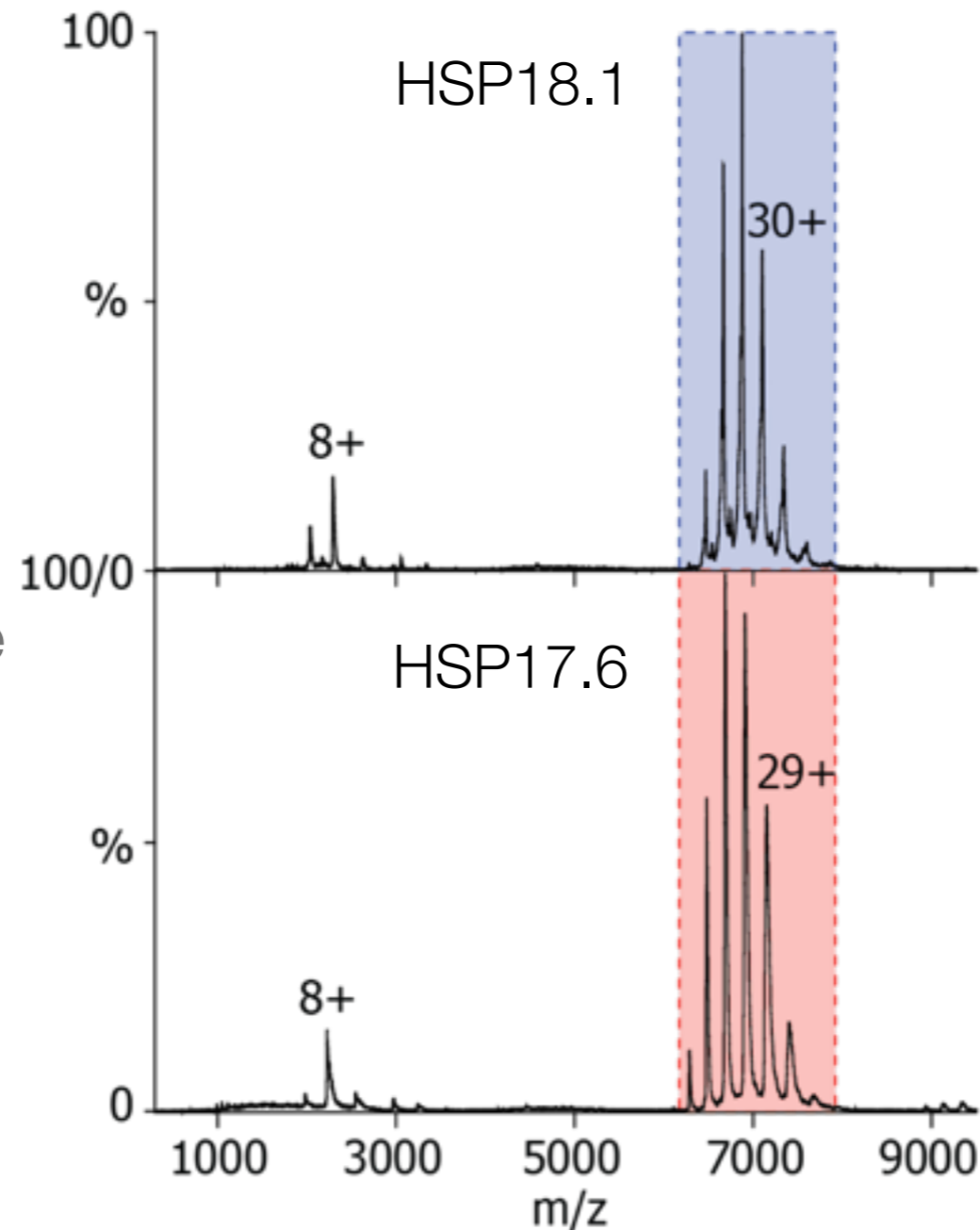
Protein dynamics

- Proteins are not static structures, but rather undergo fluctuations both at and before equilibrium
- Such 'protein dynamics' are crucial to their function in the cell
- These dynamics can span a wide range of amplitudes and timescales



Quaternary dynamics - Example

- Two homologous proteins from the same cellular compartment incubated
- Subunit exchange results in the appearance of hetero-oligomers

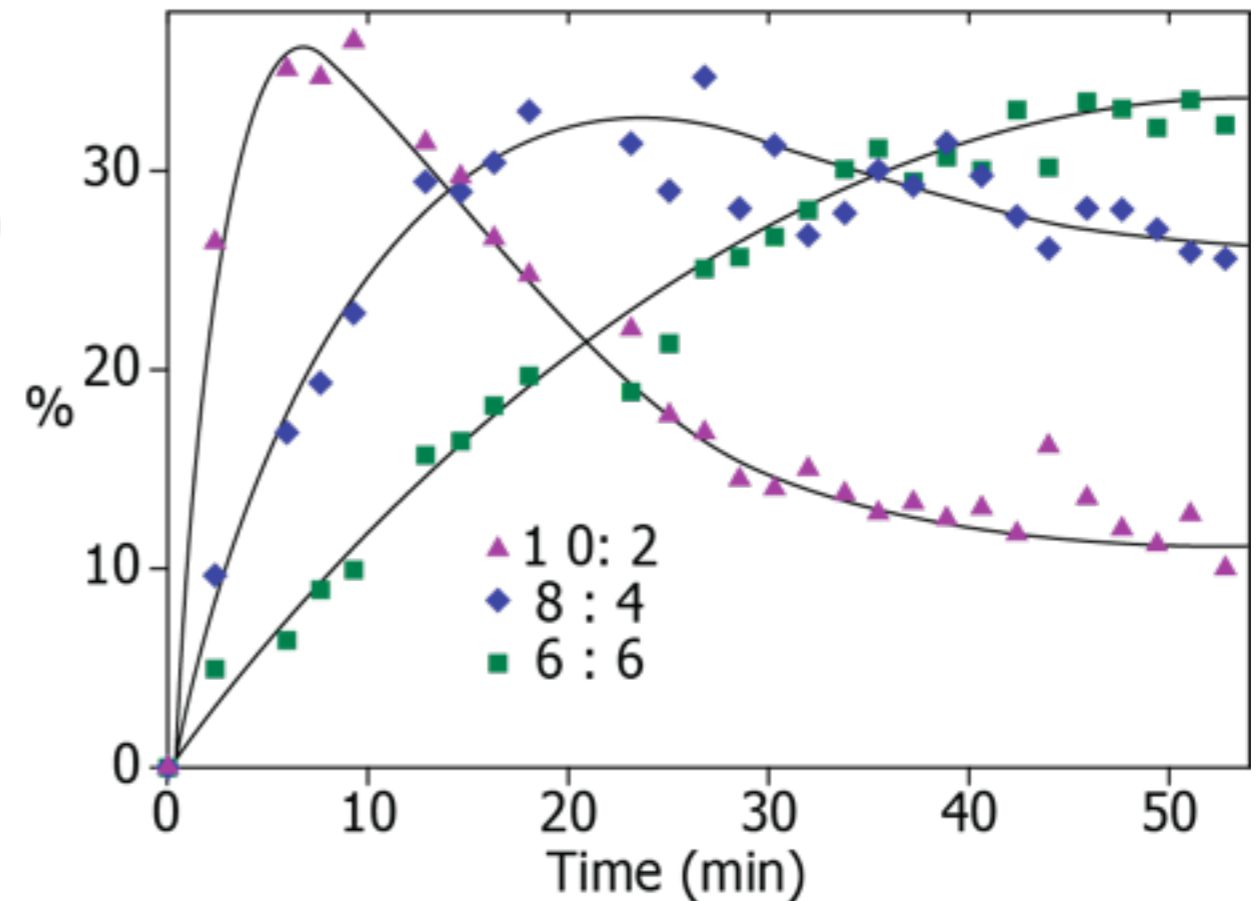
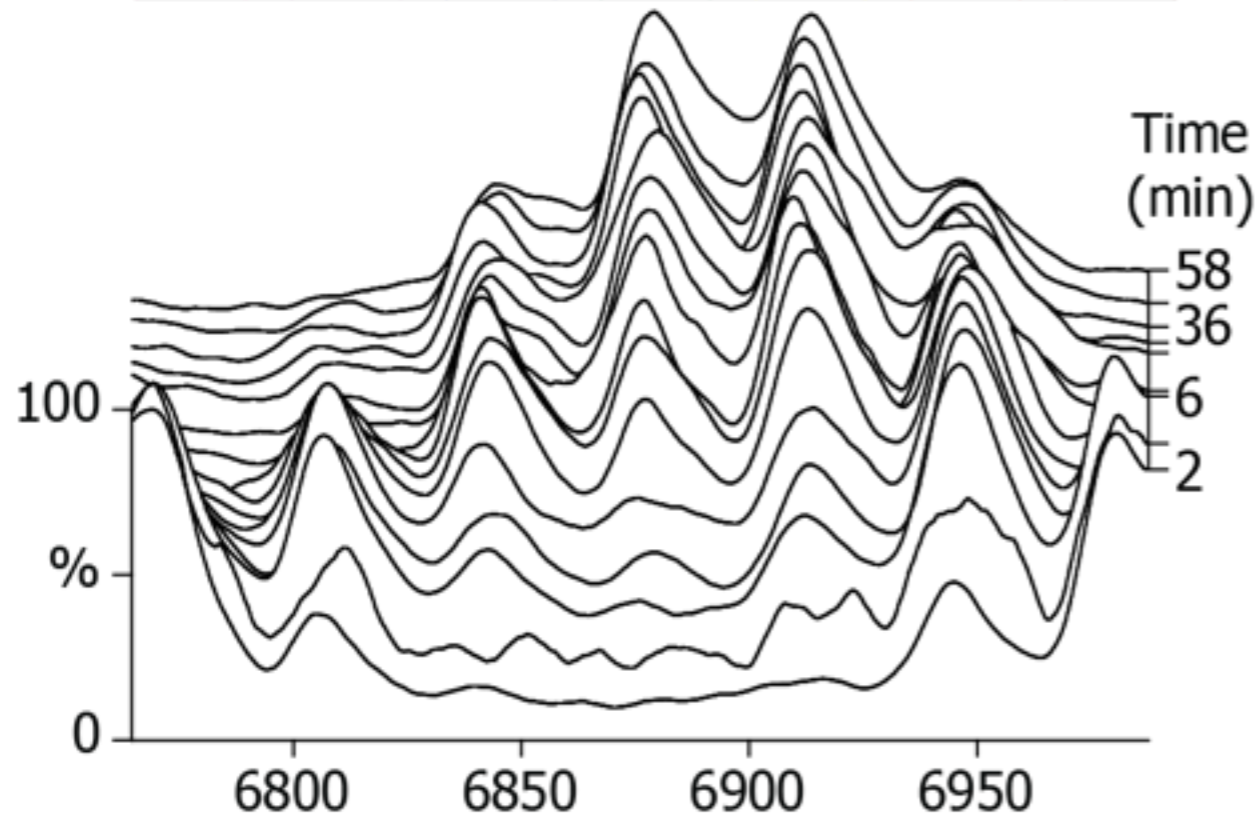


Quaternary dynamics - Example

HSP17.6

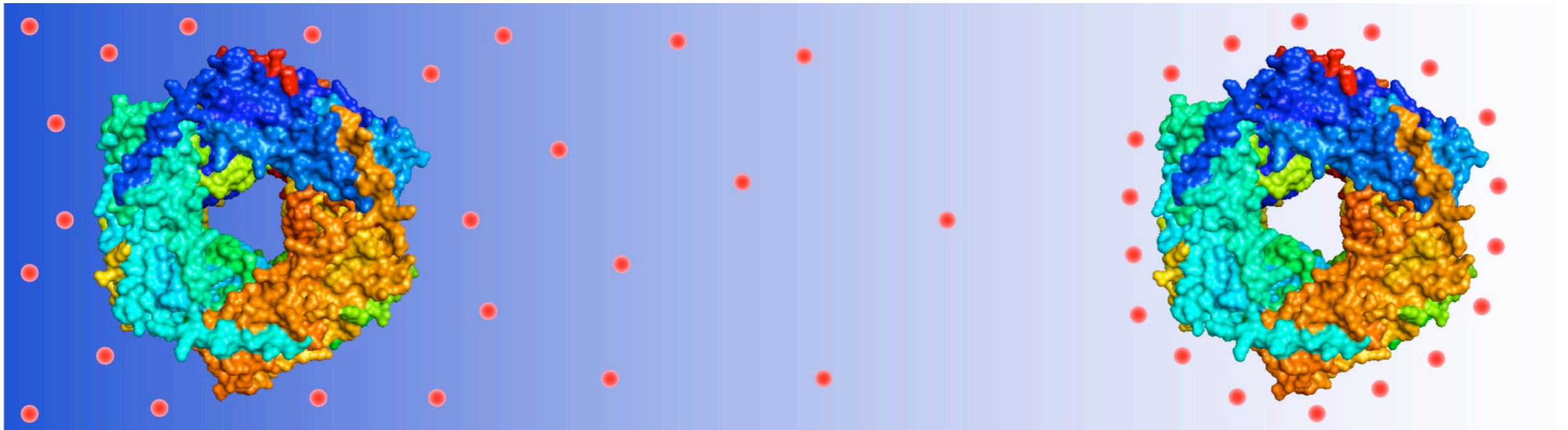
HSP18.1

12:0 10:2 8:4 6:6 4:8 2:10 0:12



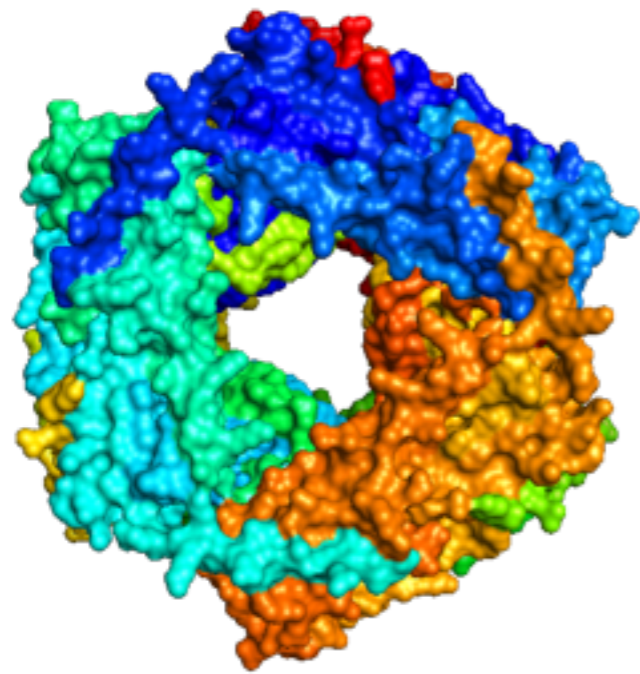
- Exchange proceeds via the movement of dimeric units
- Incorporation is via sequential incorporation of dimers into oligomers
- Hetero-assembly leads to a wide variety of possible oligomers

Preservation of structure



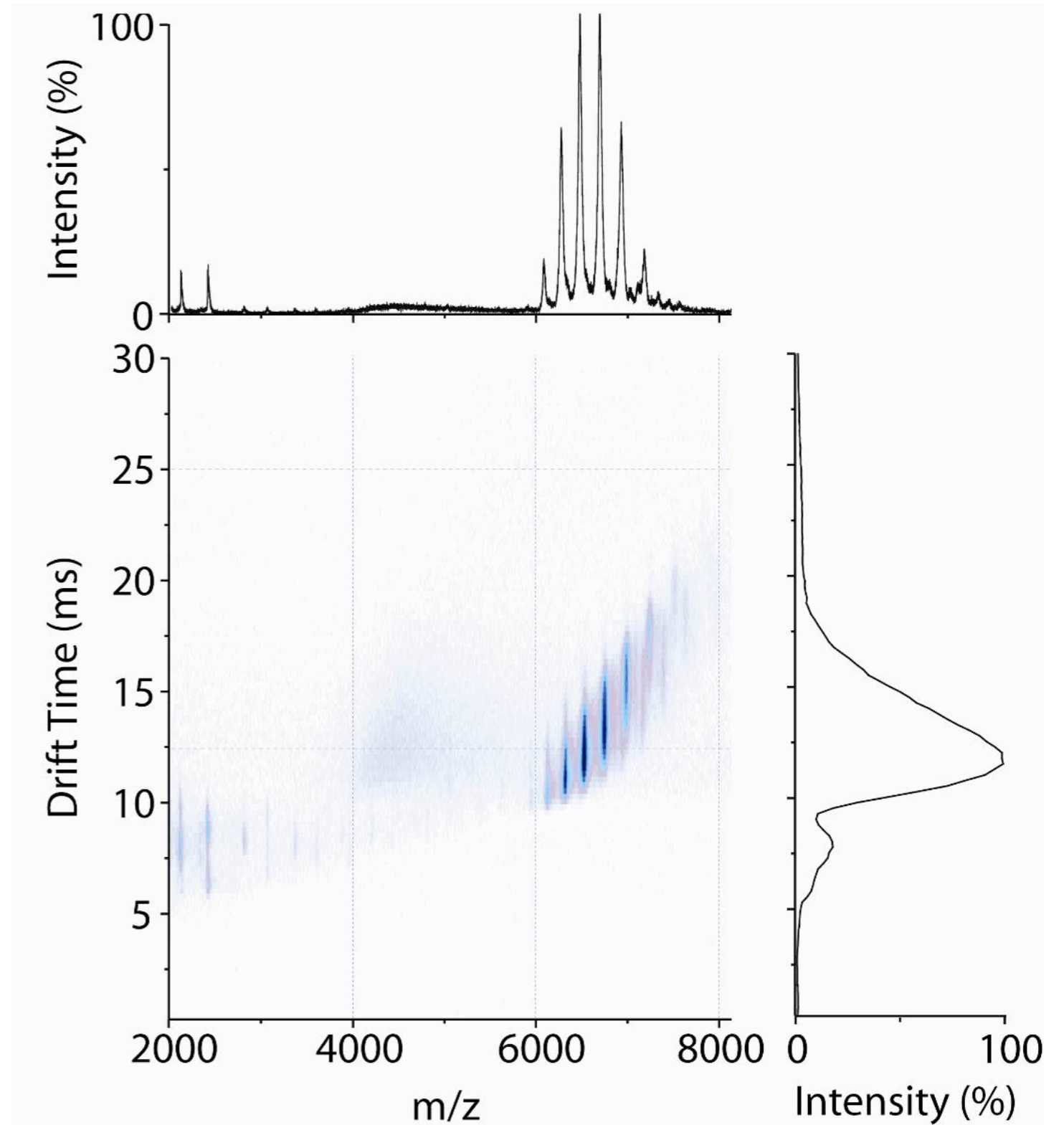
- It is clear stoichiometry is preserved in the mass spectrometer, but can we probe native structure?

IM-MS spectrum

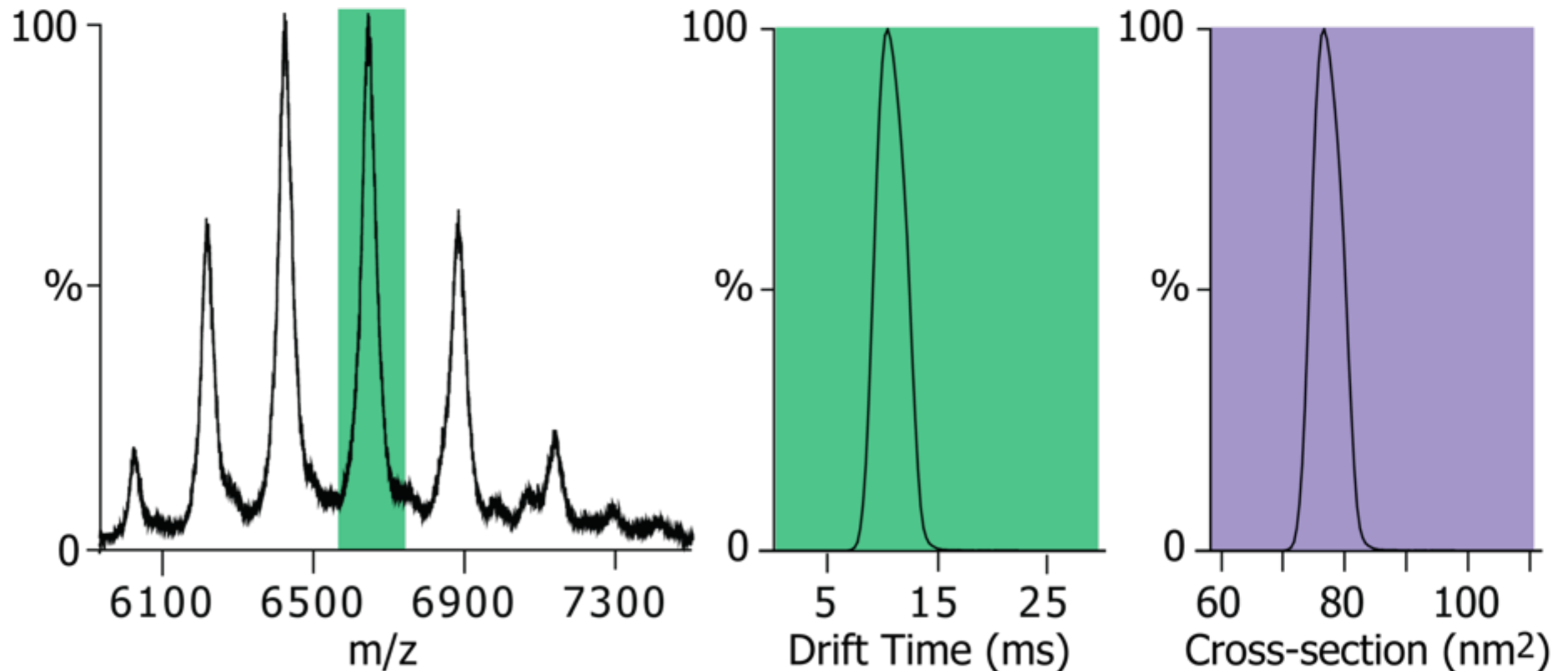


HSP16.9

- Plot of m/z versus drift time

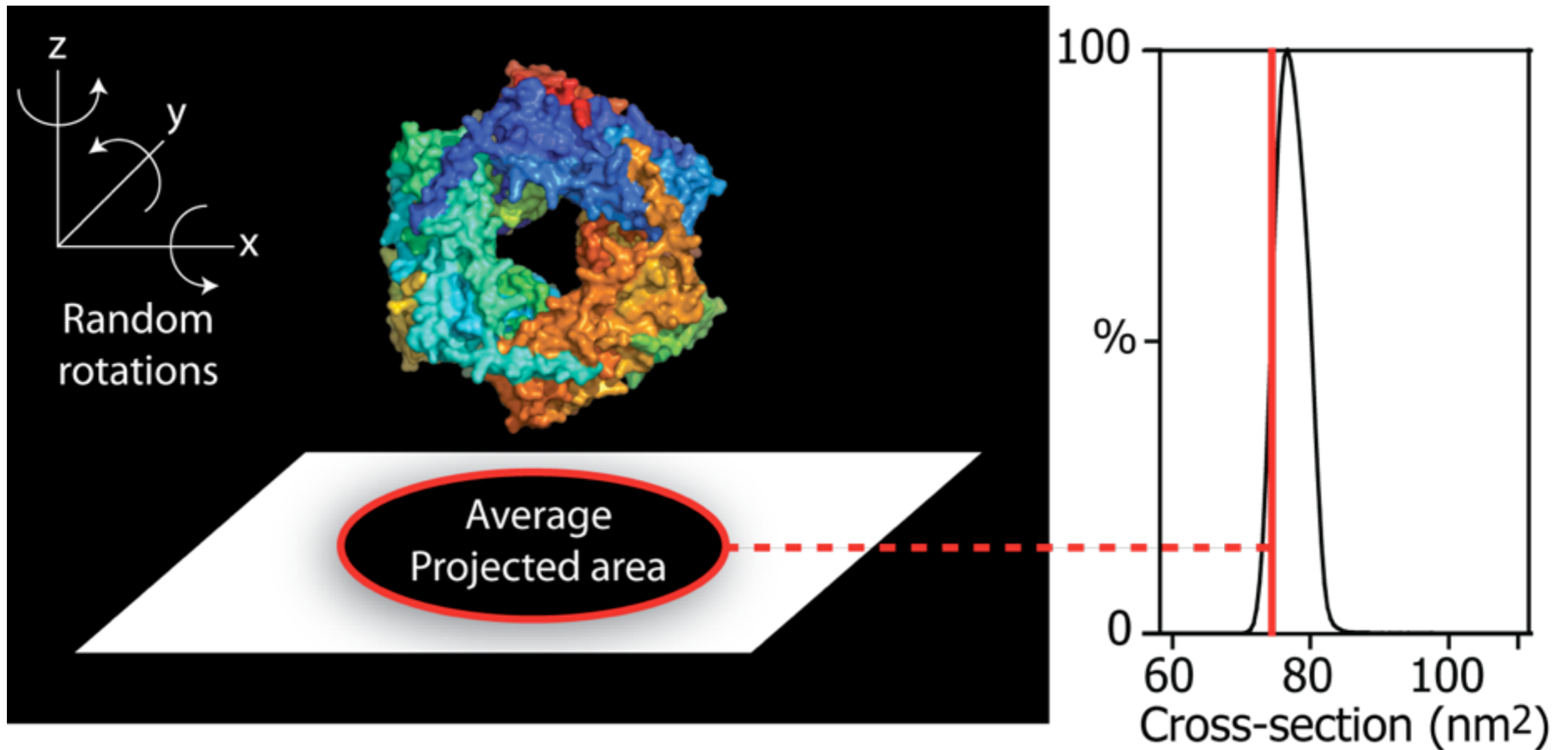


Obtaining an experimental CCS



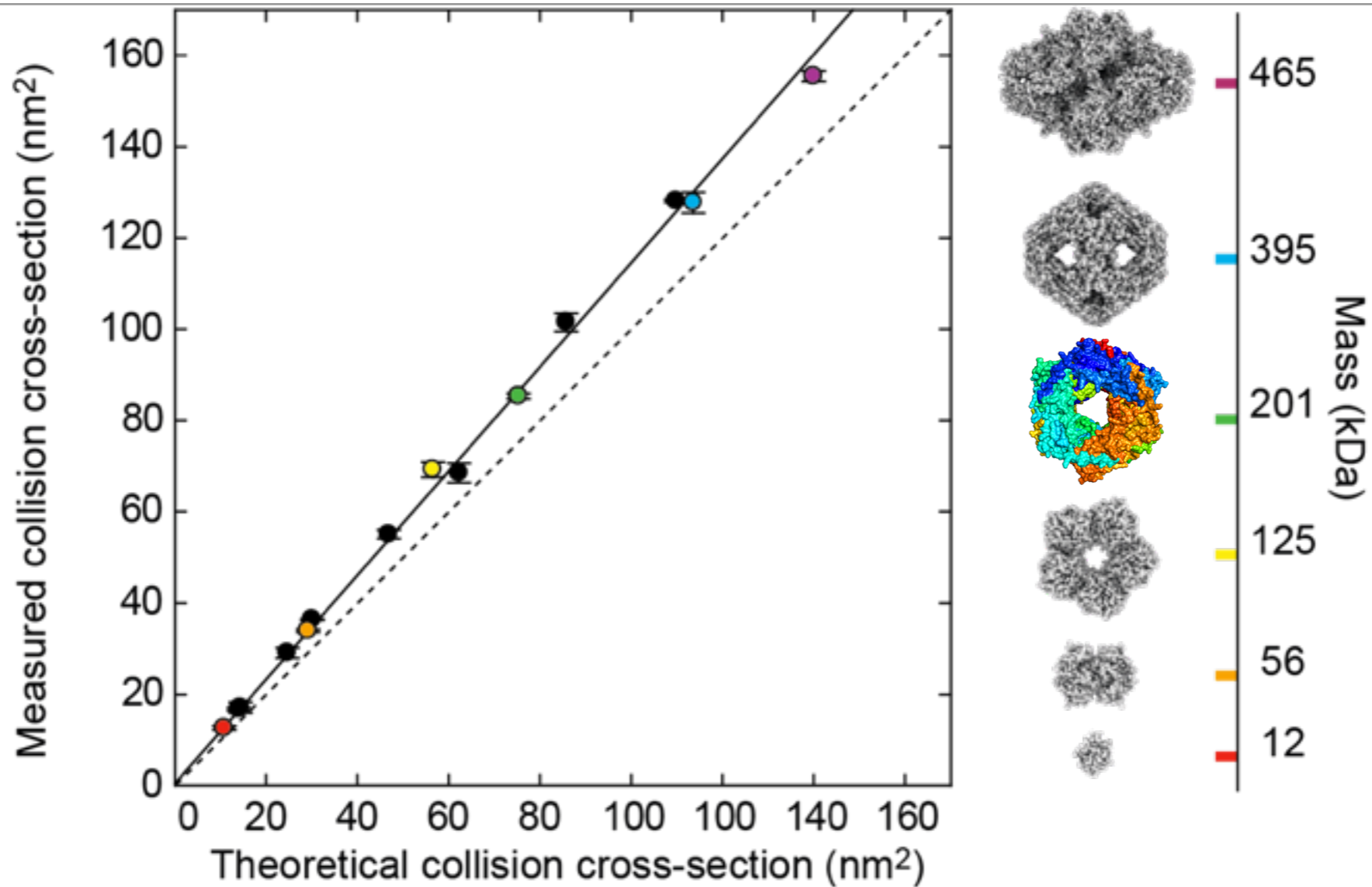
- Every feature resolved in m/z has an associated drift time distribution
- Drift time is converted into CCS either directly or via calibration

CCS values from protein structures



- Can approximate CCS as rotationally averaged projected area
- Determine 'theoretical' CCSs from solved protein structures

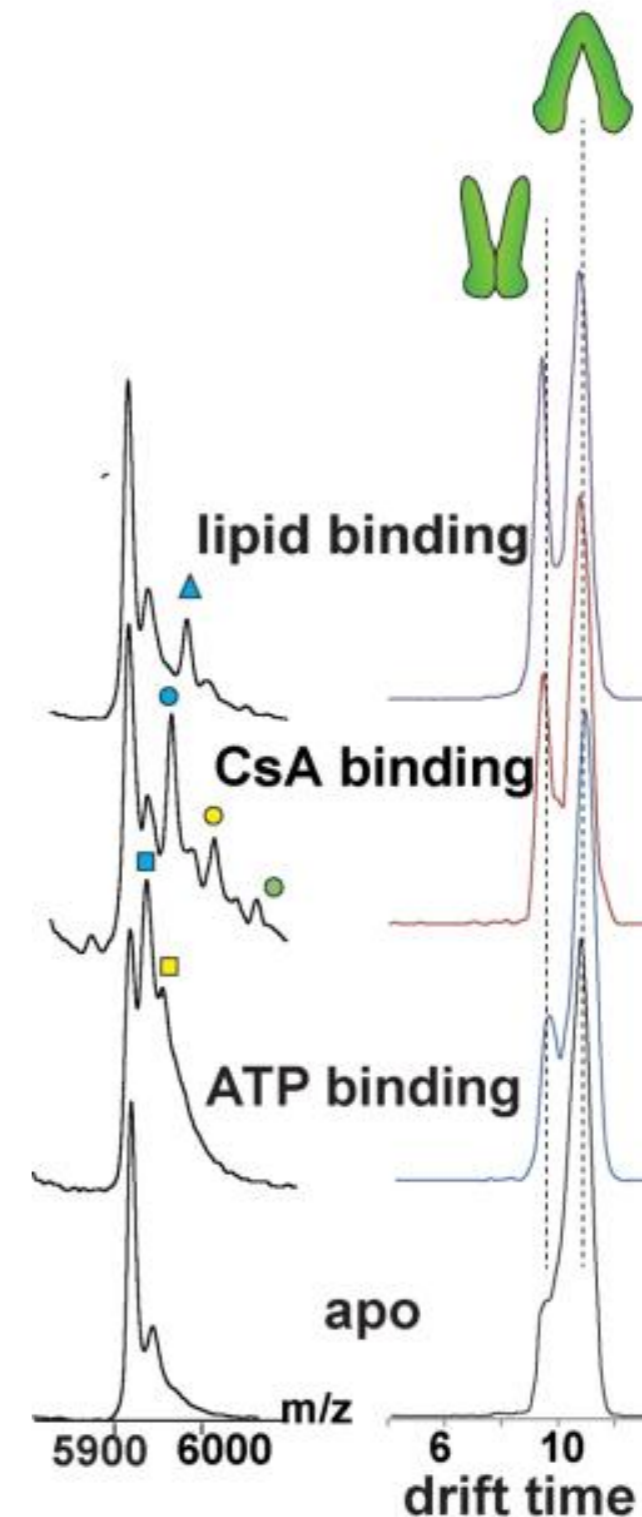
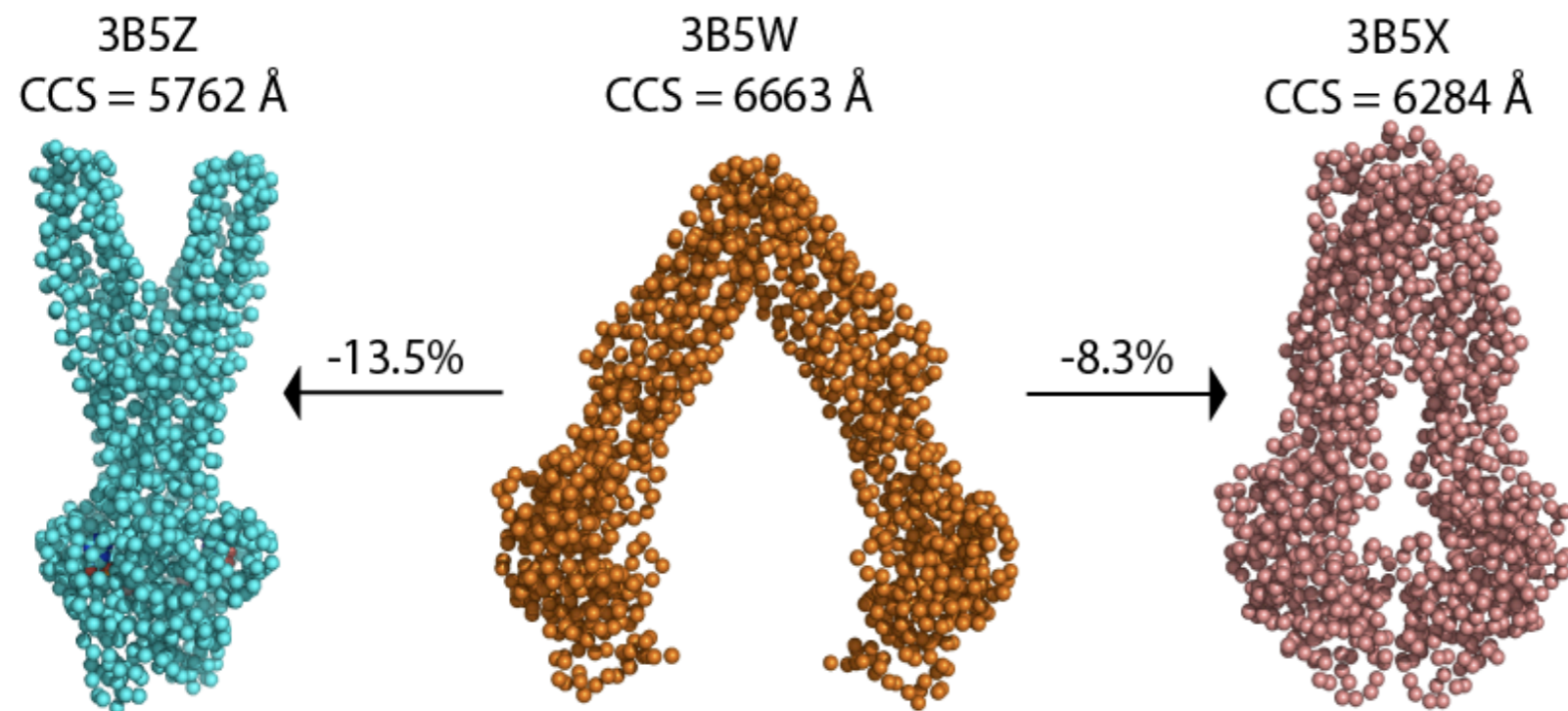
CCS comparison



- Excellent correlation between theoretical and measured values for globular proteins
- Discrepancy is due to simplicity of 'projection approximation'
- Correlation motivates use of IM measurements in assessing model structures

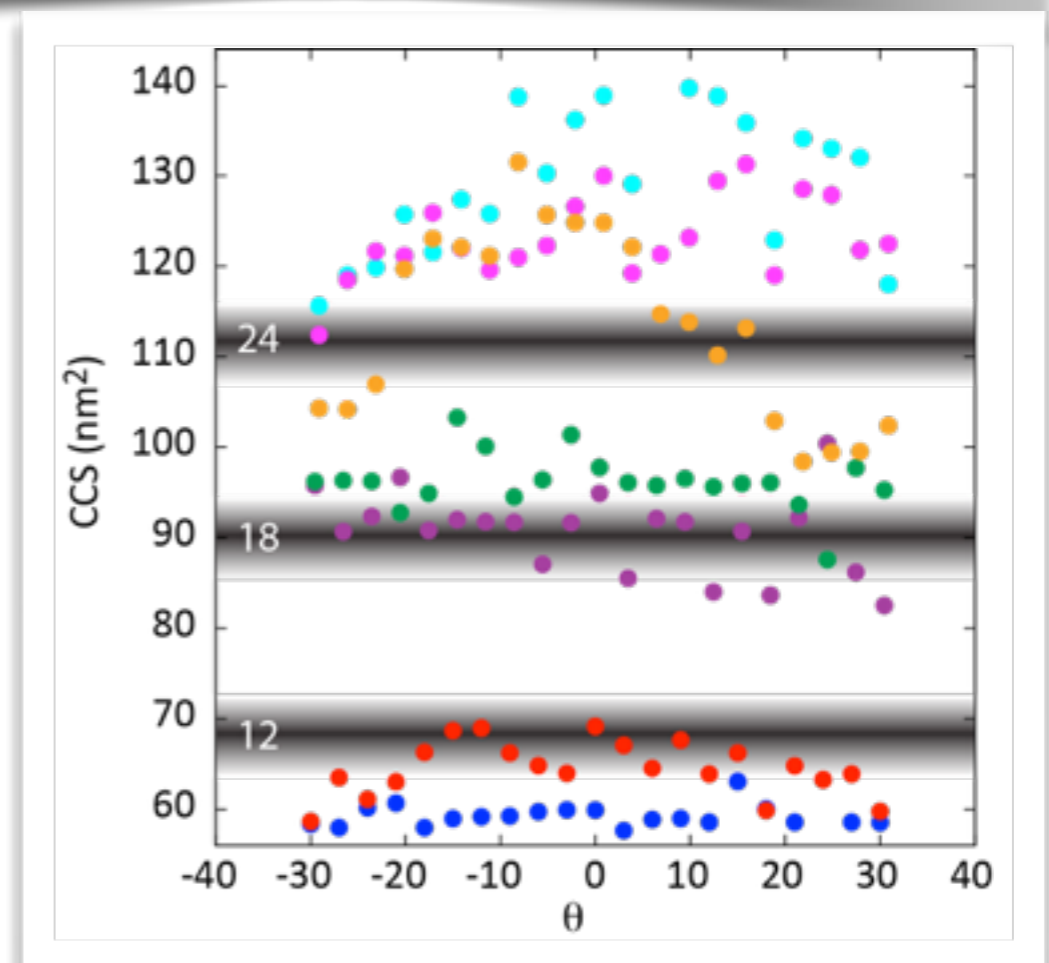
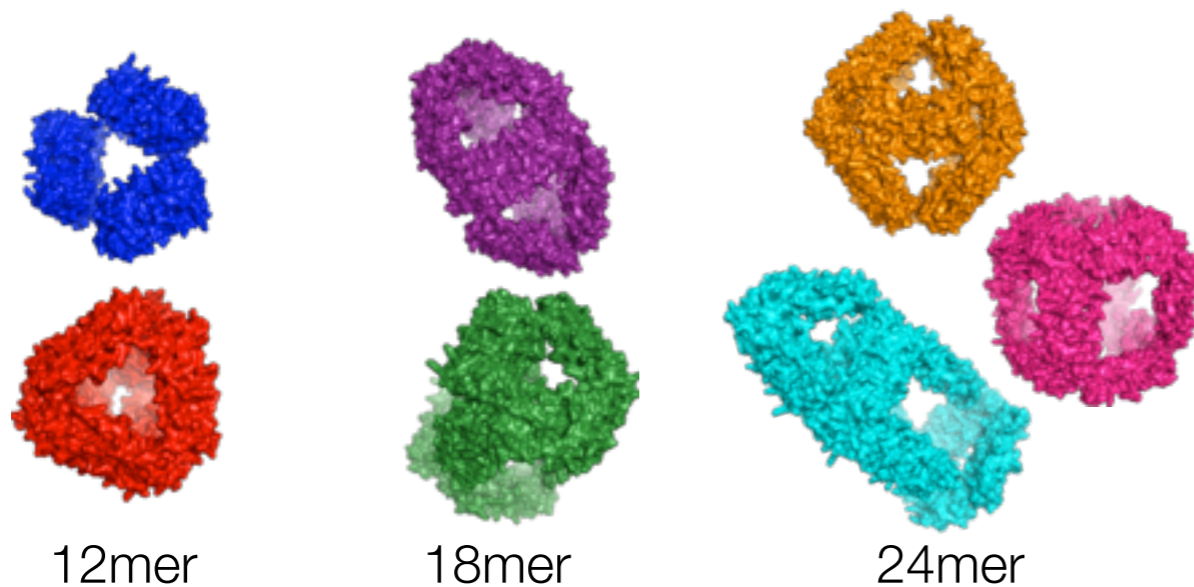
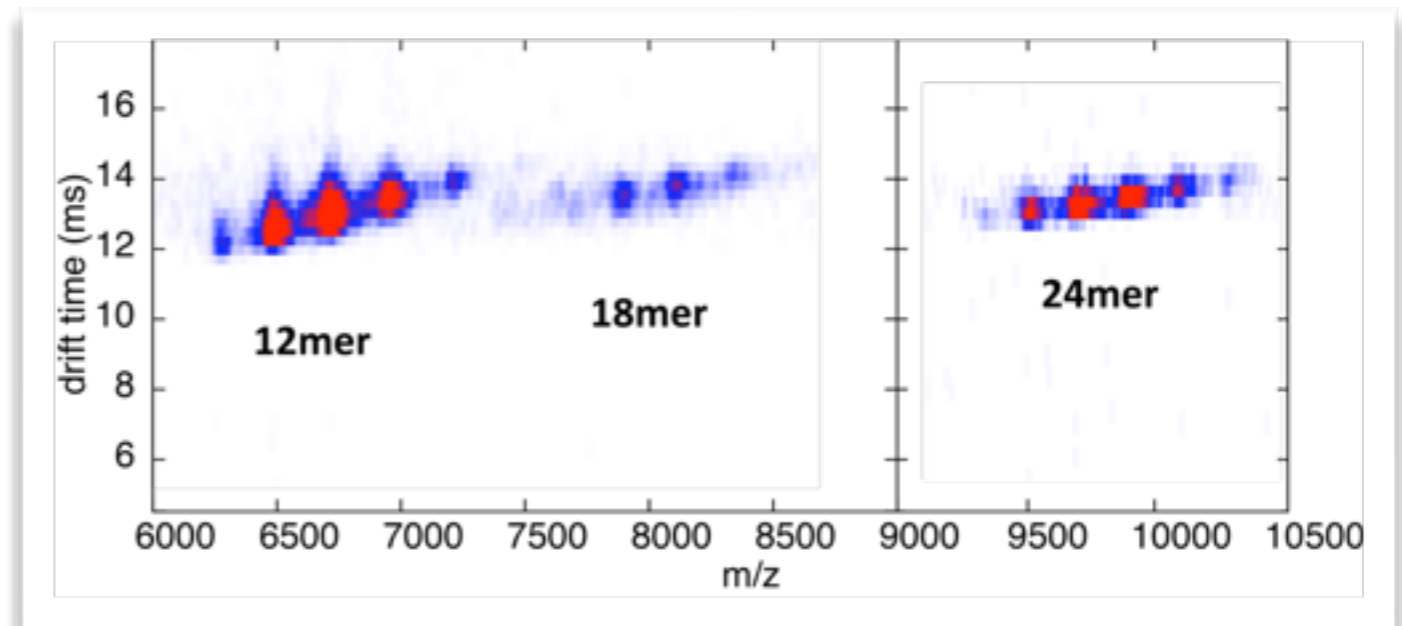
Using IM-MS to measure conformers - Example

- P-glycoprotein is a low specificity efflux pump which impairs drug delivery
- IM-MS allows the detection of different conformations - outward, inward open, inward closed (left to right)
- Small molecules affect conformational equilibrium



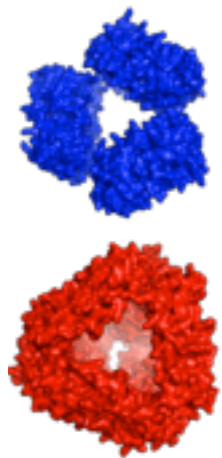
Using IM-MS to filter structures - Example

- Polydisperse oligomeric protein exists in three different stoichiometries
- Based on comparison with homologous proteins likely structures are polyhedral
- Different polyhedral models can be compared to the IMS measurements

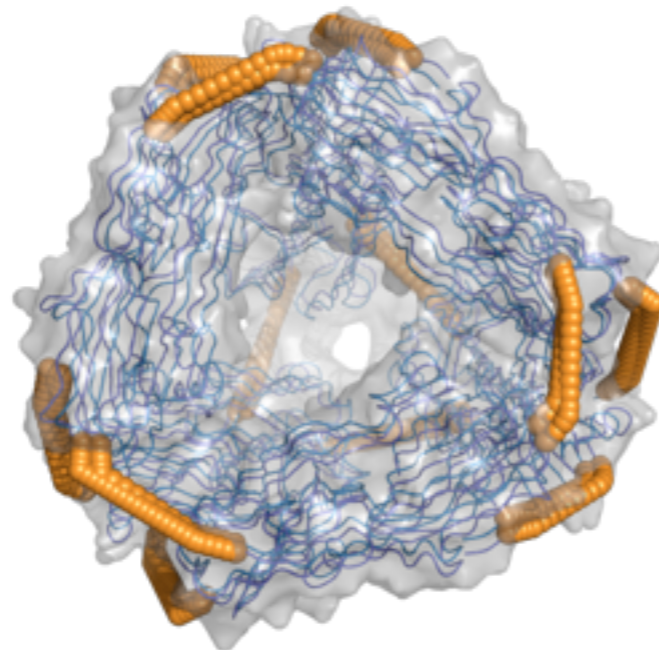


Using IM-MS to filter structures - Example

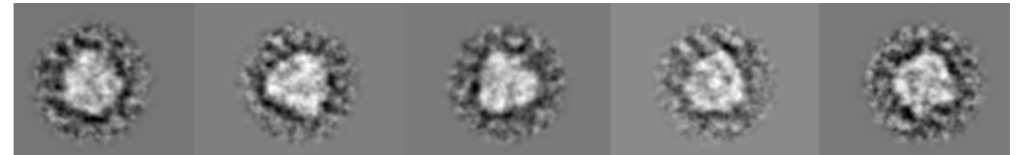
- Compare random rotations of models to TEM class averages
- Lower score is better fit
- Projected area from TEM is conceptually similar to CCS area from IM
- Combination of techniques reveals ensemble structures that



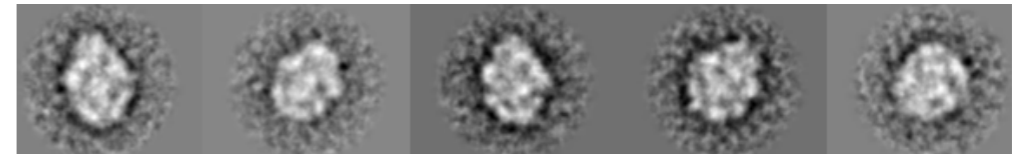
12mer



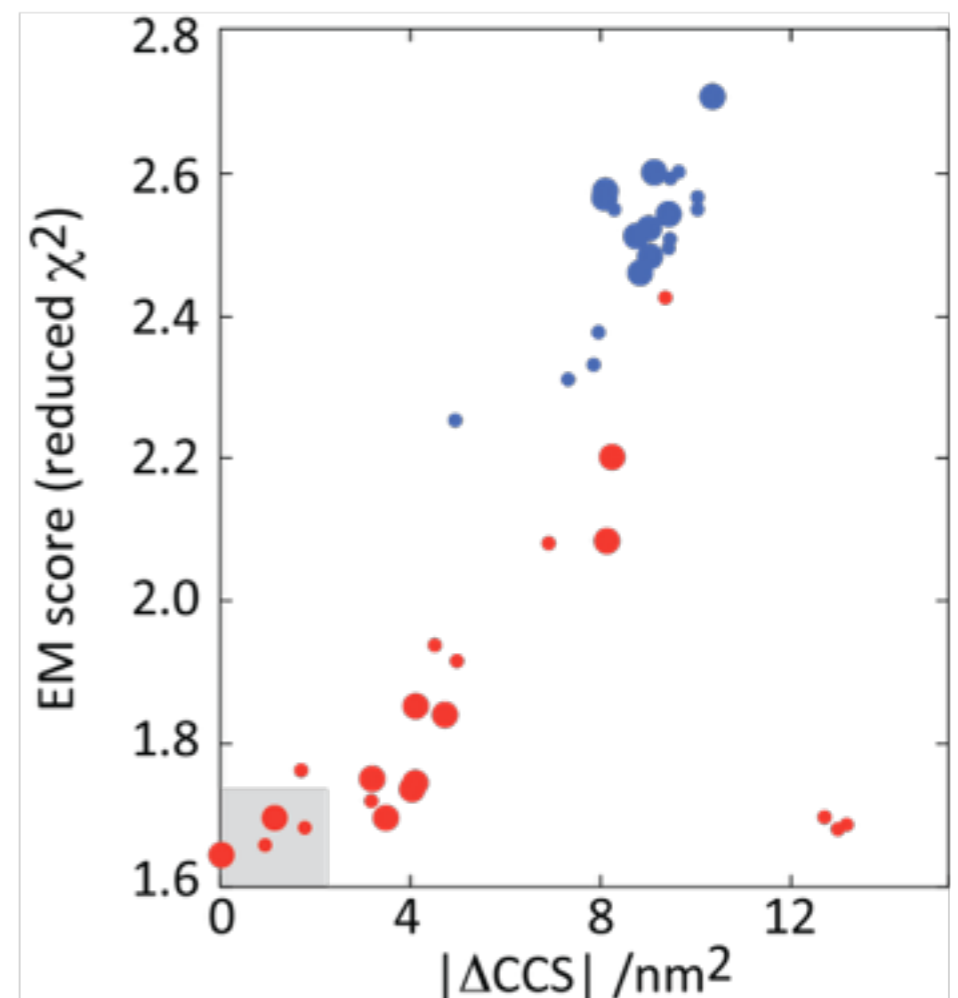
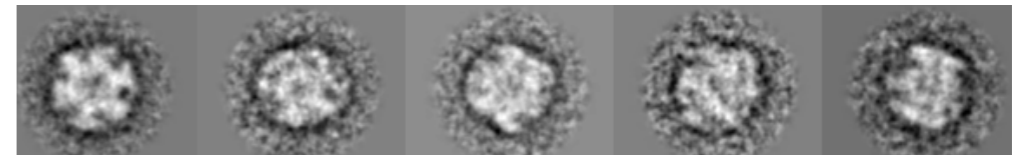
12mer



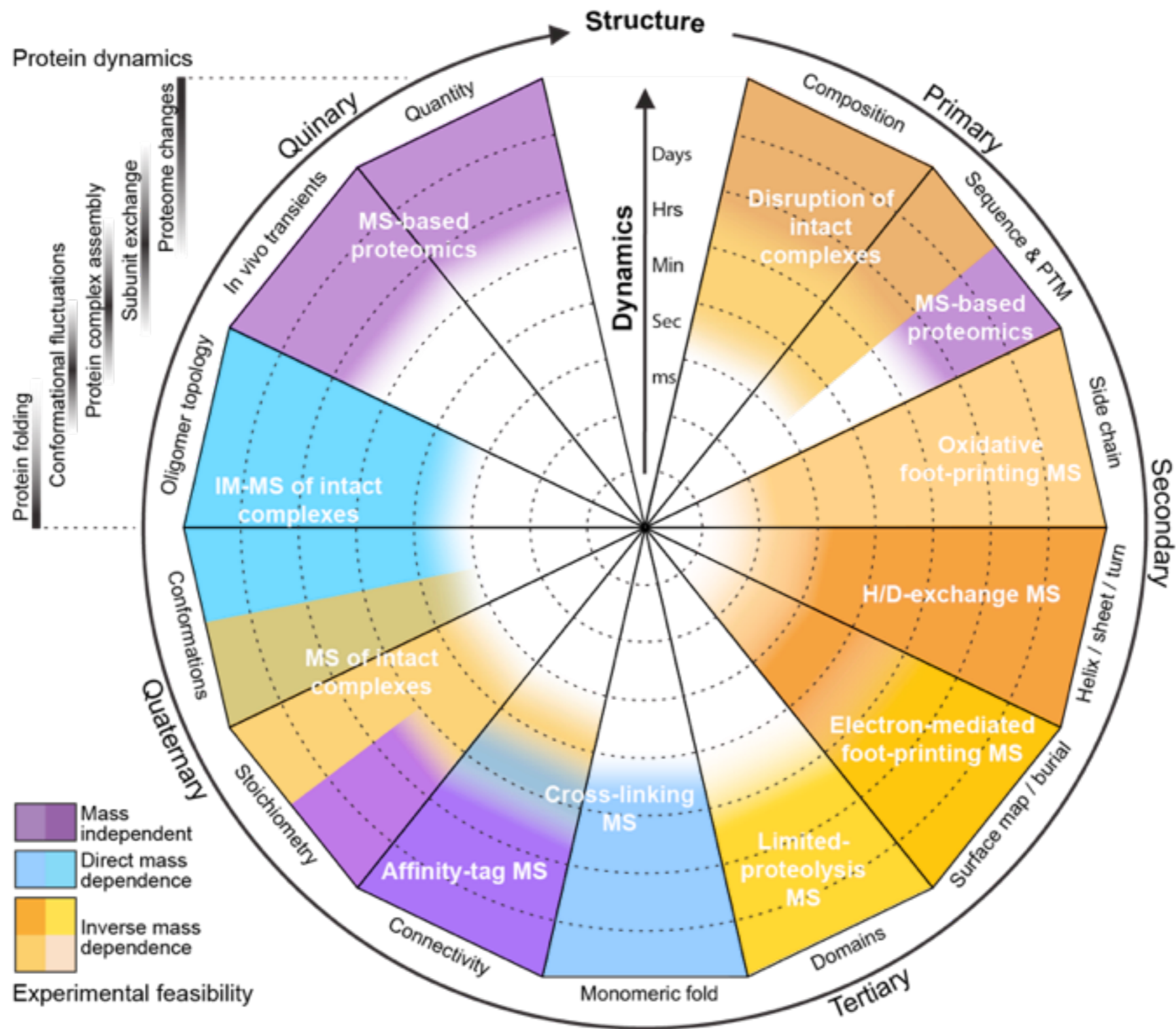
18mer



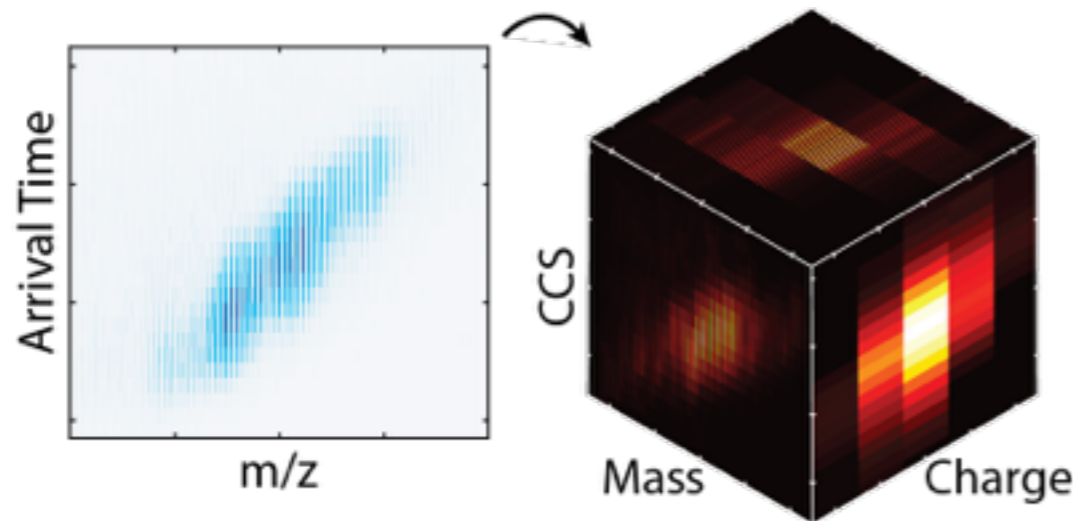
24mer



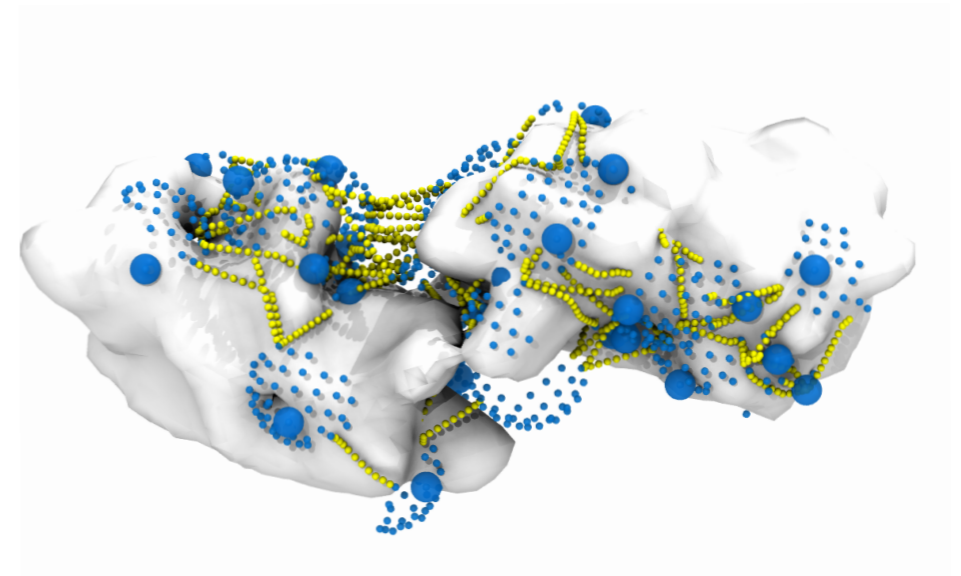
MS across wide range of time and length scales



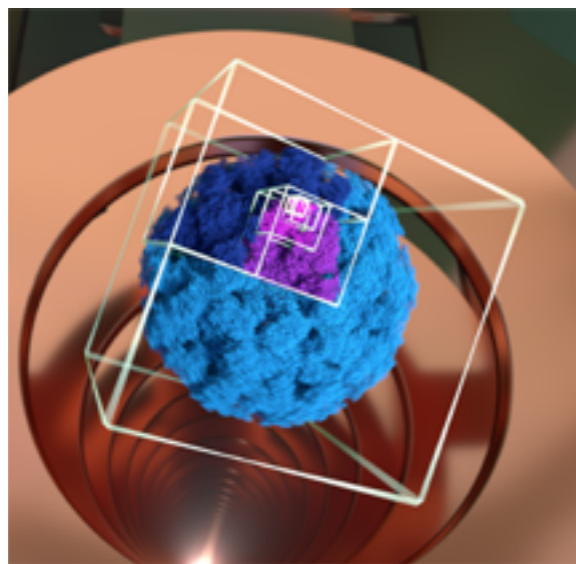
In-house software to enable quantification



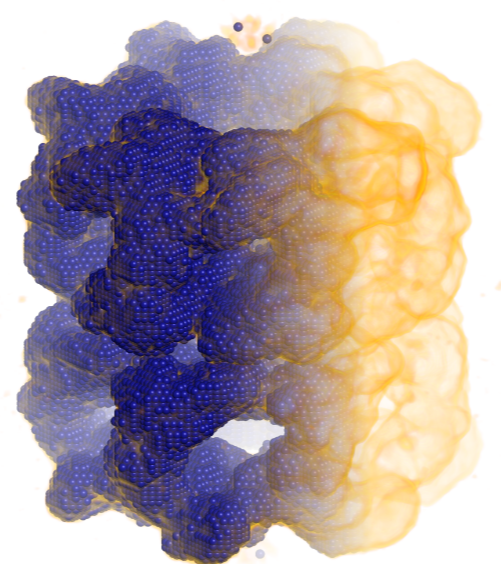
UniDec.chem.ox.ac.uk



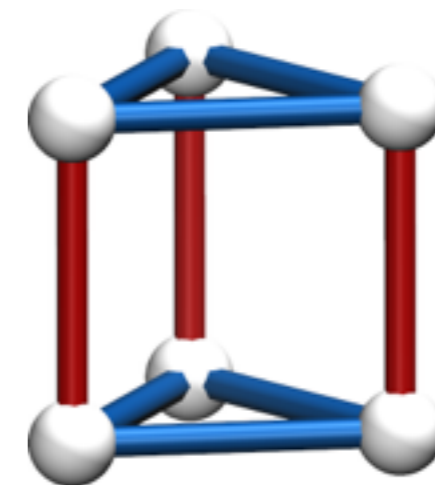
DynamixL



IMPACT.chem.ox.ac.uk



EMnIM.chem.ox.ac.uk

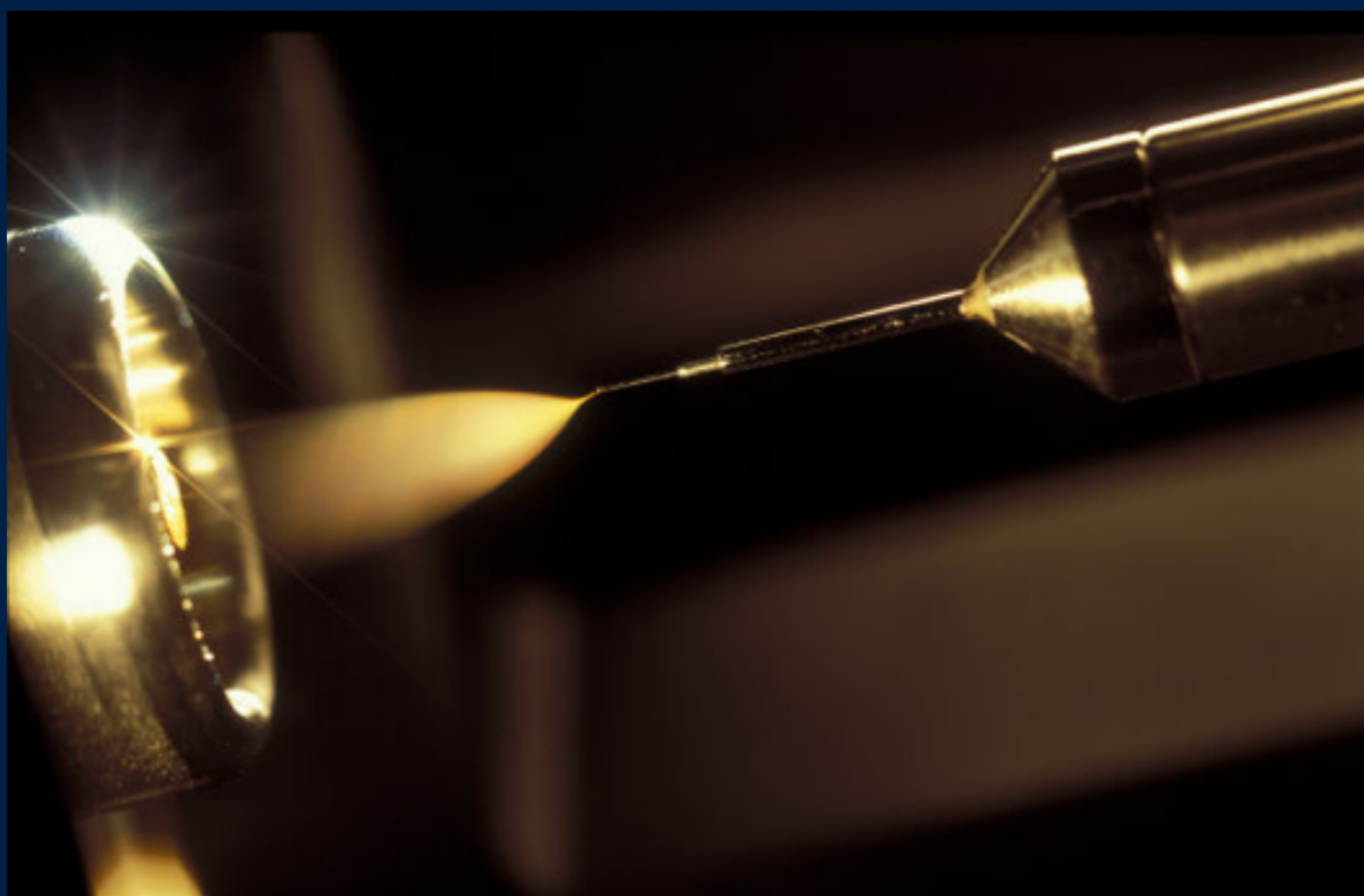


BioBOx

DEPARTMENT OF CHEMISTRY

GRADUATE COURSE IN MASS SPECTROMETRY: LECTURE 8

Mass spectrometry for biophysics and structural biology



Professor Justin Benesch, 23rd November 2016